ESSENTIALS
OF
CHEMICAL PHYSIOLOGY



HALLIBURTON


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CHEMICAL PHYSIOLOGY

By the same Author.

A TEXT-BOOK OF CHEMICAL PHYSIOLOGY AND PATHOLOGY.

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THE ESSENTIALS

OF

CHEMICAL PHYSIOLOGY

FOR THE USE OF STUDENTS

BY

W. D. HALLIBURTON, M.D., F.R.S.

FELLOW OF THE ROYAL COLLEGE OF PHYSICIANS
PROFESSOR OF PHYSIOLOGY IN KING'S COLLEGE, LONDON
AUTHOR OF 'TEXT-BOOK OF CHEMICAL PHYSIOLOGY AND PATHOLOGY'

FIFTH EDITION

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PREFACE

TO

THE FIFTH EDITION

In the present edition I have made considerable changes in the arrangement of the exercises. I have introduced a new first lesson on the detection of the elements contained in substances of physiological interest, and thus a link is made in the work of the student who passes from the study of pure chemistry to that of chemical physiology. The fats, which were somewhat meagrely treated in former editions, have now been made the subject of a separate lesson. By grouping together some of the shorter subsequent lessons, the number of those in the elementary course has still been limited to twelve. How much of each should be taken at each meeting of a practical class will necessarily be determined by individual teachers according to the time at their disposal.

A good many new exercises have been inserted in other places, and a few of the advanced lessons, *e.g.* those on milk, pancreatic digestion, muscle, and Kjeldahl's method, have been rewritten.

In carrying out the revision, my object has been to bring the work abreast of advances in science, and at the same time to keep it within moderate limits. The book does not pretend to be a complete laboratory guide. My endeavour has been to choose such exercises as can be readily performed in a practical class; work of subsidiary importance, or which demands long laboratory processes, has

therefore as a rule been excluded, and in cases where methods or tests are numerous I have selected those which I have found most trustworthy. The revision has involved a slight increase in the total number of pages, but I hope the book will still be found to contain what its title professes it to be, the Essentials of Chemical Physiology.

I have to thank Dr. H. Willoughby Lyle, who has again assisted me in reading proof-sheets and preparing the Index.

W. D. Halliburton.

King's College, 1904.

CONTENTS

PAGE

Introduction 1

ELEMENTARY COURSE

Lesson

I. The Elements contained in Physiological Compounds . . . 9

II. The Carbohydrates 13

III. The Fats 22

IV. The Proteids 27

V. The Proteids (continued) 28

VI. Foods 41

VII. The Digestive Juices—Saliva and Gastric Digestion . . . 53

VIII. The Digestive Juices (continued)—Pancreatic Digestion and Bile 69

IX. The Blood 92

X. Urine 121

XI. Urine (continued) 135

XII. Pathological Urine 146

Scheme for Detecting Physiological Proximate Principles . . 151

ADVANCED COURSE

Introduction 155

Lesson

XIII. Carbohydrates 156

XIV. Action of Malt upon Starch 159

XV. Crystallisation of Egg Albumin 160

XVI. Milk 161

XVII. The Albumoses 162

Lesson		page
XVIII.	Digestion	164
XIX.	Hæmoglobin and its Derivatives	168
XX.	Serum	172
XXI.	Coagulation of Blood	174
XXII.	Muscle and Nervous Tissue	176
XXIII.	Urea and Chlorides in Urine	183
XXIV.	Estimation of Phosphates and Sulphates in Urine	187
XXV.	Uric Acid and Creatinine	190
XXVI.	The Pigments of the Urine	192

APPENDIX

Hæmacytometers . . . 197

Hæmoglobinometers . . . 199

Polarisation of Light . . . 202

Polarimeters . . . 206

The Spectro-polarimeter . . . 209

Relation between Circular Polarisation and Chemical Constitution . 210

Mercurial Air Pumps . . . 211

Analysis of Gases . . . 214

Kjeldahl's Method of Estimating Nitrogen . . . 215

Solutions, Diffusion, Dialysis, Osmosis . . . 216

INDEX . . . 225

LIST OF ILLUSTRATIONS

FIG.			PAGE
1.	Dextrose Crystals	*Frey*	17
2.	Inosite Crystals	*Frey*	18
3.	Milk-sugar Crystals	*Frey*	19
4.	Section of Pea, showing Starch Grains	*Yeo, after Sachs*	20
5.	Fat Cells	*Schäfer*	23
6.	Simple Warm Bath		28
7.	Dialyser		31
8.	Dialyser		31
9.	Diagram of a Cell	*Schäfer*	37
10.	Milk	*Yeo*	44
11.	Colostrum Corpuscles	*Heidenhain*	44
12.	Yeast Cells	*Yeo's Physiology*	54
13.	Schizomycetes	*After Zopf.*	55
14.	Bacillus Anthracis	*Koch*	56
15.	Alveoli of Serous Gland	*Langley*	60
16.	Mucous Cells	*Langley*	60
17.	Submaxillary Gland	*Heidenhain*	60
18.	Cardiac Gland	*Klein*	62
19.	Pyloric Gland	*Ebstein*	62
20.	Cardiac Gland	*Langley*	63
21.	Alveolus of Pancreas	*Kühne and Lea*	71
22.	Leucine Crystals	*Kühne*	77
23.	Tyrosine Crystals	*Frey*	77
24.	Hæmatoidin Crystals	*Frey*	80
25.	Cholesterin Crystals	*Frey*	84

FIG.			PAGE
26.	Villus of Rat Killed during Fat Absorption	*Schäfer*	88
27.	Mucous Membrane of Frog's Intestine during Fat Absorption	*Schäfer*	89
28.	Fibrin Filaments and Blood Platelets	*Schäfer*	94
29.	Action of Reagents on Blood Corpuscles	*Schäfer*	99
30.	Oxyhæmoglobin Crystals	*Quain's Anatomy*	100
31.	Hæmin Crystals	*Preyer*	102
32.	Diagram of Spectroscope		104
33.	Figure of Spectroscope and Accessories	*McKendrick*	104
34.	Arrangement of Prisms in Direct-vision Spectroscope	*Gscheidlen*	105
35.	Stand for Direct-vision Spectroscope		106
36.	Absorption Spectra	*Rollett*	106
37.	Absorption Spectra		107
38.	Dupré's Urea Apparatus	*Gamgee*	122
39.	Urinometer	*McKendrick*	124
40.	Urea Crystals	*Frey*	124
41.	Urea Nitrate and Oxalate	*Frey*	126
42.	Triple Phosphate Crystals	*Frey*	134
43.	Uric Acid Crystals	*Frey*	136
44.	Hippuric Acid Crystals	*Frey*	140
45.	Creatine Crystals	*Frey*	142
46.	Creatinine Crystals	*Frey*	142
47.	Acid Sodium Urate	*Frey*	143
48.	Acid Ammonium Urate	*Frey*	143
49.	Envelope Crystals of Calcium Oxalate	*Frey*	144
50.	Cystin Crystals	*Frey*	144
51.	Triple Phosphate Crystals	*Bryant's Surgery*	144
52.	Calcium Phosphate Crystals	*Bryant's Surgery*	144
53.	Albuminometer of Esbach		146
54.	Two Burettes on Stand	*Sutton*	147
55.	Hot-air Oven with Gas Regulator	*Gscheidlen*	156
56.	Osazone Crystals	*Coloured plate to face*	157
57.	Absorption Spectra of Hæmoglobin, &c.		169
58.	Photographic Spectrum of Hæmoglobin and Oxyhæmoglobin	*Gamgee*	170

FIG.			PAGE
59.	Photographic Spectrum of Oxyhæmoglobin and Methæmoglobin	*Gamgee*	170
60.	Centrifugal Machine		174
61.	Absorption Spectra of Myohæmatin		178
62.	A Desiccator	*Gscheidlen*	178
63.	Absorption Spectra of Urinary Pigments	*After Hopkins*	194
64.	Gowers' Hæmacytometer		197
65.	Oliver's Hæmacytometer		198
66.	Gowers' Hæmoglobinometer		199
67.	Von Fleischl's Hæmometer		200
68.	Oliver's Hæmoglobinometer		201
69.	Model to Illustrate Polarised Light		203
70.	Model to Illustrate Polarised Light		203
71.	Model to Illustrate Polarised Light		204
72.	Diagram to Explain Polarisation of Light.		205
73.	Diagram to Explain Polarisation of Light.		206
74.	Soleil's Saccharimeter.		207
75.	Diagram of Optical Arrangements in Soleil's Saccharimeter		207
76.	Laurent's Polarimeter.		208
77.	Spectro-polarimeter of von Fleischl		209
78.	Diagram of Asymmetric Carbon Atoms.		211
79.	Diagram of Pflüger's Pump		212
80.	L. Hill's Air Pump		213
81.	Waller's Apparatus for Gas Analysis	*Waller*	214
82.	Kjeldahl's Method, Distilling Apparatus		215
83.	Diagram to Illustrate Osmosis		219

ESSENTIALS

OF

CHEMICAL PHYSIOLOGY

INTRODUCTION

Chemical Physiology or **Physiological Chemistry** deals with the chemical composition of the body and with the chemical changes it undergoes; it also deals with the composition of the food which enters, and the excretions which leave, the body.

When a chemist examines living things he is placed at a disadvantage when compared with an anatomist; for the latter can with the microscope examine cells, organisms, and structures in the living condition. The chemist, on the other hand, cannot at present state anything positive about the chemical structure of living matter, because the reagents he uses will destroy the life of the tissue he is examining. There is, however, no such disadvantage when he examines non-living matter, like food and urine, and it is therefore in the analysis of such substances that chemical physiology has made very important advances, and the knowledge so obtained is of the greatest practical interest to the student and practitioner of medicine.

The animal organism is in its earliest embryonic state a single cell; as development progresses it becomes an adherent mass of simple cells. In the later stages various tissues become differentiated from each other by the cells becoming grouped in different ways, by alteration in the shape of the cells, by deposition of intercellular matter between the cells, and by chemical changes in the living matter of the cells themselves. Thus in some situations the cells are grouped into the various epithelial linings; in others the

cells become elongated, and form muscular fibres; in the connective tissues we have a preponderating amount of intercellular material, which may become permeated with fibres, or be the seat of the deposition of calcareous salts, as in bone. Instances of chemical changes in the cells themselves are seen on the surface of the body, where the superficial layers of the epidermis become horny (*i.e.* filled with the chemical substance called keratin); in the mucous salivary glands, where the cells become filled with mucin, which they subsequently extrude; and in adipose tissue, where they become filled with fat.

In spite of these changes, the variety of which produces the great complexity of the adult organism, there are many cells which still retain their primitive structure; notable among these are the white corpuscles of the blood.

A cell may be defined as a mass of living material containing in its interior a more solid structure called the *nucleus*. The nucleus exercises a controlling influence over the nutrition and subdivision of the cell.

The living substance is usually pervaded with granules; one of these minute particles called the *centrosome* exercises an attractive influence on the granules and fibrils of the protoplasm in its neighbourhood, and the appearance so produced is called the attraction sphere. The attraction sphere becomes specially prominent, and divides into two when the cell is about to divide; this usually precedes the division of the nucleus.

Living material is called *protoplasm*, and protoplasm is characterised by (1) irritability—that is, the property of responding by some change under the influence of an external agent or stimulus: the most obvious of these changes is movement (amœboid movement, ciliary movement, muscular movement &c.); (2) its power of assimilation—that is, it is able to convert into protoplasm the nutrient material or food which is ingested; (3) its power of growth—this is a natural consequence of its power of assimilation; (4) its power of reproduction—this is a variety of growth; and (5) its power to excrete, to give out waste materials, the products of its other activities.

Of all the signs of life, those numbered 2 and 5 in the foregoing list are the most essential. Living material is in a continual state of unstable chemical equilibrium, building itself up on the one hand, breaking down on the other; the term used for the sum total of these intra-molecular rearrangements is **metabolism**. The chemical substances in the protoplasm which are the most important from this

point of view are the complex nitrogenous compounds called *Proteids*. So far as is at present known, proteid material is never absent from living substance, and is never present in anything else than that which is alive or has been formed by the agency of living cells. It may therefore be stated that **Proteid Metabolism** is the most essential characteristic of vitality.

The chemical structure of protoplasm can only be investigated after the protoplasm has been killed. The substances it yields are (1) **Water**; protoplasm is semi-fluid, and at least three-quarters of its weight, often more, are due to water. (2) **Proteids**. These are the most constant and abundant of the solids. A proteid or albuminous substance consists of carbon, hydrogen, nitrogen, oxygen, with sulphur and phosphorus in small quantities only. In *nuclein*, a proteid-like substance obtained from the nuclei of cells, phosphorus is more abundant. The proteid obtained in greatest abundance from the cell-protoplasm is *nucleo-proteid*: that is, a compound of proteid with varying amounts of *nuclein*. White of egg is a familiar instance of an albuminous substance or proteid, and the fact (which is also familiar) that this sets into a solid on boiling will serve as a reminder that the greater number of the proteids found in nature have a similar tendency to coagulate under the influence of heat and other agencies. (3) Various other substances occur in smaller proportions, the most constant of which are **lecithin**, a phosphorised fat; **cholesterin**, a monatomic alcohol; and **inorganic salts**, especially phosphates and chlorides of calcium, sodium, and potassium.

It will be seen from this rapid survey of the composition of the body how many are the substances which it is necessary we should study; the food from which it is built up is also complex, for animals do not possess to such an extent as plants do the power of building up complex from simple materials.

The substances out of which the body is built consist of chemical elements and of chemical compounds, or unions of these elements.

The **elements** found in the body are carbon, hydrogen, nitrogen, oxygen, sulphur, phosphorus, fluorine, chlorine, iodine, silicon, sodium, potassium, calcium, magnesium, lithium, iron, and occasionally manganese, copper, and lead.

Of these very few occur in the free state. Oxygen (to a small extent) and nitrogen are found dissolved in the blood-plasma; hydrogen is formed by putrefaction in the alimentary canal. With some few exceptions such as these, the elements enumerated above are found combined with one another to form compounds.

The **compounds**, or, as they are often termed in physiology, the *proximate principles*, found in the body are divided into—

(1) Mineral or inorganic compounds.

(2) Organic compounds, or compounds of carbon.

The **inorganic compounds** present are water, various acids (such as hydrochloric acid in the gastric juice), ammonia (as in the urine), and numerous salts, such as calcium phosphate in bone, sodium chloride in blood and urine, and many others.

The **organic compounds** are more numerous; they may be subdivided into—

1. Various groups of alcohols and organic acids, and their derivatives, such as the fats and carbohydrates

2. Various derivatives of ammonia, amides, amines, urea, &c.

3. Aromatic bodies, or derivatives of benzene.

4. Proteids, the most important of all, and substances allied to proteids, like the albuminoids, pigments, and ferments.

A more convenient practical method of grouping the proximate principles of the body and of food is the following:—

Inorganic		*Water.*
		Salts—*e.g.* chlorides and phosphates of sodium and calcium.
Organic	Nitrogenous	*Proteids*—*e.g.* albumin, myosin.
		Albuminoids—*e.g.* gelatin, keratin.
		Simpler nitrogenous bodies—*e.g.* lecithin, creatine, urea.
	Non-nitrogenous	*Fats*—*e.g.* butter, fats of adipose tissue.
		Carbohydrates—*e.g.* sugar, starch.
		Simple organic bodies—*e.g.* alcohol, cholesterin, vegetable acids and salts, lactic acid.

Many of the substances enumerated above only occur in small quantities. The most important are the inorganic substances, *water* and *salts*; and the organic substances, *proteids*, *carbohydrates*, and *fats*. It is necessary in our subsequent study of the principles of chemical physiology that we should always keep in mind this simple classification; the subdivision of organic substances into proteids, fats, and carbohydrates forms the starting point, the A B C, as one might say, of chemical physiology.

I will conclude this introductory chapter by giving a list of the apparatus and reagents necessary for a practical study of the subject, and some tables to which it will be often found convenient to refer.

The following set of reagents conveniently contained in 4 to 6 oz. glass stoppered bottles should be provided for each two students:—

Sulphuric acid, concentrated.
„ „ 25 per cent.

Sulphuric acid, 0·1 per cent.
Nitric acid, concentrated.
Fuming nitric acid.
Hydrochloric acid, concentrated.
„ „ 0·2 per cent.[1]
Acetic acid, glacial.
„ „ 20 per cent.
„ „ 2 „
Glyoxylic acid.
Caustic potash, 20 per cent.
„ „ 0·1 „
Ammonia.
Sodium carbonate, 1 per cent.
Ammonium sulphide solution.
Ammonium sulphate, saturated solution.
Silver nitrate, 1 per cent.
Barium chloride, saturated solution.
Ammonium molybdate solution.
Millon's reagent.[2]
Solution of ferrocyanide of potassium.
„ litmus
„ sodium phosphate.
„ iodine in potassium iodide.
„ ferrous sulphate.
„ ferric chloride.
„ sodium nitroprusside.
Alcohol.
Ether.
Esbach's reagent.[3]
Solution of copper sulphate, 1 per cent.
Fehling's solution.
Lime water.

The following additional reagents will be required by those taking the advanced course :—

Solution of mercuric chloride.
„ potassium ferricyanide.
Sodium carbonate, saturated solution.
„ chloride, saturated solution.
„ „ 10-per-cent. solution.
Magnesium sulphate, saturated solution.
Baryta mixture.[4]
Sodium acetate solution.[5]
Phosphoric acid, 0·5 per cent.

[1] Made by adding 994 c.c. of water to 6 c.c. of the concentrated hydrochloric acid of the British Pharmacopœia.

[2] Mercury is dissolved in its own weight of strong nitric acid. The solution so obtained is diluted with twice its volume of water. The decanted clear liquid is Millon's reagent.

[3] Ten grammes of picric acid and 20 grammes of citric acid are dissolved in 800 to 900 c.c. of boiling water, and then sufficient water added to make up a litre.

[4] Made by mixing 1 volume of barium nitrate solution with 2 of barium hydrate solution, both saturated *in the cold.*

[5] Prepared as follows :—Sodium acetate, 100 grammes; water, 900 c.c.; glacial acetic acid, 100 c.c.

In addition to these, there should be kept in stock in the laboratory, to be given out for the lessons in which they are used, the following :—

Solid sodium chloride.
„ magnesium sulphate.
„ ammonium sulphate.
„ sodio-magnesium sulphate.
Standard solution of uranium acetate or nitrate for estimating phosphates.[1]
Standard solution of mercuric nitrate for estimating urea.
„ „ „ „ „ chlorides.
„ „ silver „ „ „
Caustic soda, 40 per cent.
Bromine.
Solution of potassium bichromate.
Phenyl hydrazine hydrochloride.
Solid sodium acetate.
Phospho-tungstic acid.
Glacial phosphoric acid.
Dry cupric oxide.
Soda-lime.
Metallic sodium.

Each student should be provided with—

A Bunsen burner.
1 dozen test-tubes in test-tube stand.
2 or 3 4-oz. flasks.
2 flat porcelain dishes.
2 or 4 4-oz. beakers.
2 small glass funnels and a funnel stand.
A glass stirring rod and a small pipette.
1 burette.
An iron tripod with wire gauze.
Filter papers and litmus papers.
A 100-c.c. cylindrical measuring glass.
A thermometer marked in degrees Centigrade.
A urinometer.
A tin can on a stand to be used as a water-bath.

Apparatus which is not so frequently used, such as that employed in generating carbonic anhydride, carbonic oxide, or sulphuretted hydrogen, may be given out as required. The laboratory should also possess a good balance, with its accessories, water and air baths, kept at various temperatures, retorts, and analytical apparatus generally. The microscope, polarimeter, spectroscope, dialyser, are also frequently employed in chemico-physiological investigations. Apparatus and reagents for carrying out the Kjeldahl process are also necessary.

WEIGHTS AND MEASURES

The weights and measures usually employed in science are those of the metric system, but as in this country the practical physician still largely uses English grains and ounces, we may compare the two systems in the following way :—

[1] Instructions how to make standard solutions will be given in the lessons where they are used.

Weights

(English System)

1 grain	=	0·0648 gramme
1 ounce = 437·5 grains	=	28·3595 grammes
1 lb. = 16 oz. = 7,000 grains	=	453·5925 „

The scruple = 20 grains = 1·296 gramme, and the drachm = 60 grains = 3·888 grammes, are retained in use, but neither is an aliquot part of the ounce; though for practical purposes an ounce is considered to consist of 8 drachms.

(Metric System)

1 milligramme =	0·001	gramme	=	0·015432 grain
1 centigramme =	0·01	„	=	0·154323 „
1 decigramme =	0·1	„	=	1·543235 „
1 gramme			=	15·43235 grains
1 decagramme = 10		grammes	=	154·3235 „
1 hectogramme = 100		„	=	1543·235 „
1 kilogramme = 1,000		„	=	15432·35 „
			=	2 lb. 3 oz. 119·8 „

Measures of Length

(English System)

1 inch = 25·4 millimetres
1 foot = 12 inches = 304·8 millimetres

(Metric System)

The standard of length is the metre; subdivisions and multiples of which, with the prefixes milli-, centi-, and deci-, on the one hand, and deca-, hecto-, and kilo-, on the other, have the same relation to the metre as the subdivisions and multiples of the gramme, in the table just given, have to the gramme, thus:

1 millimetre = 0·001	metre	=	0·03937 inch
1 centimetre = 0·01	„	=	0·3937 „
1 decimetre = 0·1	„	=	3·93707 inches
1 metre		=	39·37079 „

Measures of Capacity

(English System)

1 minim	=	0·059 cubic centimetre
1 fluid drachm = 60 minins	=	3·549 cubic centimetres
1 fluid ounce = 8 fluid drachms	=	28·398 „
1 pint = 20 fluid ounces	=	567·936 „
1 gallon = 8 pints	=	4·54837 litres

(Metric System)

In the metric system the measures of capacity are intimately connected with the measures of length; we thus have cubic millimetres, cubic centimetres, and so forth. The standard of capacity is the litre, which is equal to 1,000 cubic centimetres; and each cubic centimetre is the volume of 1 gramme of distilled water at 4° C.[1]

[1] 4° C. is the temperature at which water has the greatest density. For practical purposes measures are more often constructed so that a cubic centimetre holds a gramme of water at 16° C., which is about the average temperature of rooms. The true cubic centimetre contains only 0·999 gramme at 16° C.

1 cubic centimetre (generally written c.c.) = 16·931 minims.
1 litre = 1,000 c.c. = 1 pint 15 oz. 2 drs. 11 m. = 35·2154 fluid ounces.
1 cubic inch = 16·386 c.c.

THERMOMETRIC SCALES

The scale most frequently used in this country is the Fahrenheit scale; in this the freezing-point of water is 32°, and the boiling-point 212°. On the Continent the Réaumur scale is largely employed, in which the freezing-point is 0°, and the boiling-point 80°. In scientific work the Centigrade scale has almost completely taken the place of these; in this system the freezing-point is 0°, and the boiling-point 100°.

To convert degrees Fahrenheit into degrees Centigrade, subtract 32 and multiply by $\frac{5}{9}$, or $C = (F-32)\frac{5}{9}$. Conversely, degrees Centigrade may be converted into degrees Fahrenheit by the following formula: $F = \frac{9}{5}C + 32$.

TENSION OF AQUEOUS VAPOUR IN MILLIMETRES OF MERCURY FROM 10° TO 25° C.

10°. 9·126	14°. 11·882	18°. 15·351	22°. 19·675
11°. 9·751	15°. 12·677	19°. 16·345	23°. 20·909
12°. 10·421	16°. 13·519	20°. 17·396	24°. 22·211
13°. 11·130	17°. 14·409	21°. 18·505	25°. 23·582

TABLE OF THE DENSITY OF WATER AT TEMPERATURES BETWEEN 0° AND 30° C.

0°. 0·99988	8°. 0·99988	16°. 0·99900	24°. 0·99738
1°. 0·99993	9°. 0·99982	17°. 0·99884	25°. 0·99714
2°. 0·99997	10°. 0·99974	18°. 0·99866	26°. 0·99689
3°. 0·99999	11°. 0·99965	19°. 0·99847	27°. 0·99662
4°. 1·00000	12°. 0·99955	20°. 0·99827	28°. 0·99635
5°. 0·99999	13°. 0·99942	21°. 0·99806	29°. 0·99607
6°. 0·99997	14°. 0·99930	22°. 0·99785	30°. 0·99579
7° 0·99994	15°. 0·99915	23°. 0·99762	

SYMBOLS AND ATOMIC WEIGHTS OF THE PRINCIPAL ELEMENTS

Aluminium	Al	27·02	Fluorine	F	19·1	Oxygen	O	16·0
Antimony	Sb	120·0	Gold	Au	197·0	Phosphorus	P	30·96
Arsenic	As	74·9	Hydrogen	H	1·0	Platinum	Pt	194·3
Barium	Ba	136·8	Iodine	I	126·53	Potassium	K	39·04
Bismuth	Bi	208·0	Iron	Fe	55·9	Silver	Ag	107·66
Boron	B	10·9	Lead	Pb	206·4	Silicon	Si	28·3
Bromine	Br	79·75	Magnesium	Mg	24·0	Sodium	Na	23·05
Cadmium	Cd	112·0	Manganese	Mn	55·0	Strontium	Sr	87·3
Calcium	Ca	39·9	Mercury	Hg	199·8	Sulphur	S	31·98
Carbon	C	12·0	Nickel	Ni	58·6	Tin	Sn	118·8
Chlorine	Cl	35·45	Nitrogen	N	14·01	Tungsten	W	183·6
Copper	Cu	63·2	Osmium	Os	200	Zinc	Zn	117·8

ELEMENTARY COURSE

LESSON I

THE ELEMENTS CONTAINED IN PHYSIOLOGICAL COMPOUNDS

1. Take a fragment of meat about the size of a pea and place it in a porcelain crucible over a Bunsen flame. Note that it chars, showing the presence of carbon, and that it gives off the unpleasant odour of burning flesh, which is due to the fact that it contains the nitrogenous substances called proteids. In course of time the organic material is completely burnt up, and a small amount of white ash or inorganic material is left behind.

2. Repeat the experiment with a pure organic substance like sugar. Note that no ash is left. Charring, as before, indicates the presence of carbon, but there is no characteristic smell of burning nitrogenous substances (absence of nitrogen).

3. The tests for carbon depend on the fact that when this element is oxidised it gives rise to carbon dioxide; the test for hydrogen depends on the fact that when this element is oxidised it gives rise to water. If all the carbon dioxide and water formed by oxidation from a weighed amount of any organic substance under examination are collected and estimated, the amount of carbon and hydrogen respectively which it contains can be easily calculated. The following exercises, however, deal only with the qualitative detection of these elements.

4. *Tests for Carbon.*—The following tests can be carried out with sugar.

(*a*) When burnt in the air it chars and subsequently the carbon entirely disappears, passing off in combination with oxygen as carbon dioxide (carbonic acid gas).

(*b*) Mix some of the powdered sugar in a dry mortar with about ten times the quantity of cupric oxide (which has been freed from water by previous heating); place the mixture in a dry test-tube provided with a rubber cork perforated by a bent glass tube which dips into either lime water or baryta water. Heat the tube over a Bunsen flame, and as the carbon of the sugar becomes oxidised carbon dioxide comes off and causes a white precipitate of calcium or barium carbonate, as the case may be.

5. *Test for Hydrogen.*—In the experiment just described (4*b*) note that drops of water due to oxidation of hydrogen condense in the colder parts of the test-tube.

6. *Tests for Nitrogen.*—The greater number of tests for this element are due to the circumstance that on the breaking up of organic substances which contain it, it is given off as ammonia. If the ammonia is all collected and estimated, the amount of nitrogen can be easily calculated. Kjeldahl's method for carrying out this quantitative analysis is described in the Appendix. The following exercises, however, are qualitative only.

(*a*) The characteristic odour of burning flesh, horn, hair, feathers, &c. has been already noted, and, though only a rough test, is very trustworthy.

(*b*) Take a little dried albumin and mix it thoroughly in a mortar with about twenty times the amount of soda-lime and heat in a test-tube over a Bunsen flame. Ammonia comes off in the vapours produced, and may be recognised by (i.) its odour; (ii.) it turns moistened red litmus paper (held over the mouth of the tube) blue; (iii.) it gives off white fumes with a glass rod (held over the mouth of the tube) which has been dipped in hydrochloric acid.

(*c*) Lassaigne's test for nitrogen is thus described by Milroy:—

Take about 0·1 gramme of dried albumin in a dry test-tube and add about 1 gramme of metallic sodium (freed from petroleum by dipping in ether). Heat gradually until the mass is glowing red. Dip the tube while still glowing into a beaker containing a few c.c. of distilled water; the tube will break and its contents mix with the water. Filter and label the filtrate A; add to this filtrate a little strong solution of potash, one or two drops of cold saturated solution of ferrous sulphate and a drop of ferric chloride solution. Bring the mixture to boiling point, then cool and acidify with hydrochloric acid. The fluid becomes bluish green, and gradually a precipitate of Prussian blue separates out. This test is due to the fact that some of the nitrogen is fixed as sodium cyanide, and this gives the Prussian blue reaction with the reagents added.

7. *Tests for Sulphur.*

(*a*) In the foregoing test (6*c*) the sulphur of the albumin combines with the sodium to form sodium sulphide. This may be detected by taking some of the filtrate A and adding freshly prepared solution of sodium nitroprusside; a reddish violet colour forms.

(*b*) Some of the sulphur in proteid is firmly combined and some loosely combined. The loosely combined sulphur gives the following test:—

Boil some albumin solution with twice its volume of 20-per-cent. potash solution, and evaporate the mixture nearly to dryness in a porcelain capsule. Dissolve the residue in water and place it in a flask; acidify with sulphuric acid and sulphuretted hydrogen comes off, which may be recognised by its odour and also by the blackening of a piece of filter paper (placed in the neck of the flask) which has been previously moistened with lead acetate and ammonium carbonate.

(*c*) Take some dried albumin and fuse with a mixture of potash and potassium nitrate. Cool; dissolve in water and filter. The filtrate will give the following test for sulphates:—Acidulate with hydrochloric acid and add barium chloride; a white precipitate of barium sulphate is produced.

(*d*) Take some solution of albumin and heat in an open dish in a fume cupboard for at least an hour with large excess of fuming nitric acid, renewing the acid from time to time as necessary. The resulting fluid will give the test for sulphate as in *c*.

8. *Tests for Phosphorus.*—The two tests just described (7*c* and *d*) may be repeated with some substance, like nucleo-proteid or lecithin, which contains phosphorus in organic combination. The resulting fluid in each case gives the following test for phosphates:—Mix it with half its volume of nitric acid; add ammonium molybdate in excess and boil; a yellow crystalline precipitate falls.

The reactions described in the foregoing exercises show how the processes of pure chemistry may be employed for the detection of some of the most important elements that occur in substances of physiological importance, and thus form a fitting introduction to a study of physiological chemistry.

They show, in the first instance, how the substances with which we have to deal fall under the two main categories of organic and inorganic. In some of the tissues of the body, like bone and tooth, the inorganic or mineral material is in excess, but in the softer portions of the organism the organic compounds are in great preponderance.

Organic chemistry is sometimes defined as the chemistry of the carbon compounds; carbon is in all cases present, and is usually the most abundant element.

The most important of the nitrogenous substances are the proteids, as already explained in the introductory chapter, and the detection and estimation of nitrogen is thus an exercise of the highest interest.

All the proteids contain a small amount of sulphur; keratin or horny material contains more than an ordinary proteid does.

Phosphorus is another element of considerable importance, being present in nuclein and nucleo-proteids, and also in certain complex fats, of which lecithin may be taken as a type. Iodine occurs united to proteid material in the colloid substance of the thyroid gland; iron in the pigment of the blood called hæmoglobin; sodium, calcium, potassium, and other metals in the inorganic substances of the body. It would, however, lead us too far into the regions of pure chemistry to undertake exercises for the detection of these and other other elements which might be mentioned, and have been already commented upon. The teacher of physiological chemistry is bound to assume that the students who come before him have already passed through a course of ordinary chemistry.

The main interest of the exercises selected as types lies in their physiological application. As a rule an element is detected by breaking up or oxidising the more or less complex molecule in which it occurs into substances of simpler nature, and then performing tests for these simpler products. Thus carbon is identified by the formation of carbon dioxide, nitrogen by the formation of ammonia, and so forth.

A great many reactions which can be performed in the test-tube imitate those which are performed in the body. Reactions *in vitro* and *in vivo*, to use the technical phrases, often, though not always, run parallel. Life, from one point of view, is a process of combustion or oxidation; the fuel is supplied by the food; this becomes assimilated, and so forms an integral part of the living substance of the body; it is then burnt up by the oxygen brought to it by the blood-stream, giving rise to animal heat and other

manifestations of energy; and finally the simple products of oxidation or chemical breakdown are carried to the organs of excretion (lung, skin, kidney, &c.), where they are discharged from the body.

A candle consists principally of carbon and hydrogen; when it is burnt the products are carbonic acid gas and water; the former may be detected by means of lime water, the latter, by holding a dry beaker upside down for a few moments over the burning candle, when the moisture will condense on the cold glass.

The body is more complex than a candle, but so far as its carbon and hydrogen are concerned the main products of combustion are the same. The carbon dioxide is discharged by the expired air, as may be proved by blowing it into lime water. The water finds an outlet by several channels, lungs, skin, and kidneys. The presence of nitrogen in the body is perhaps the most striking chemical distinction between it and a candle, and here again the process of metabolism runs a course analogous in some degree to our experiments *in vitro*. Here again the most important and abundant substance which contains the waste nitrogen is the simple material ammonia, but ammonia is only discharged as such to a very small extent in health. It unites with carbon and oxygen to form the body called urea (CON_2H_4), which finds its way out of the body *via* the urine. The urine also contains the sulphates, due to the oxidation of the sulphur of the proteid constituents of protoplasm, and the phosphates due to the similar oxidation of the phosphorus of such substances as lecithin and nuclein. Some of the salts of the urine, however, in particular the chlorides, come directly from the food. This we shall discuss at the proper place when we come to the study of the urine.

LESSON II

THE CARBOHYDRATES

1. Note the general appearance of the specimens of grape sugar or dextrose, cane sugar, dextrin, and starch which are given round.

2. Put some of each into cold water. Starch is insoluble; dextrose, cane sugar, and dextrin dissolve after a time, but more readily in hot water.

3. *Trommer's test.*—Put a few drops of copper sulphate solution into a test-tube, then solution of dextrose, and then strong caustic potash. On adding the caustic potash a precipitate is first formed, which, owing to the presence of the sugar, rapidly redissolves, forming a blue solution. On boiling this a yellow or red precipitate (cuprous hydrate or oxide) forms.

4. *Fehling's test.*—Fehling's solution is a mixture of copper sulphate, caustic soda, and Rochelle salt of a certain strength. It is used for estimating dextrose quantitatively (see Lesson XII.). It may be used as a qualitative test also. Boil some Fehling's solution; if it remains clear it is in good condition; add to it an equal volume of solution of dextrose and boil again. Reduction, resulting in the formation of cuprous hydrate or oxide, takes place as in Trommer's test.

5. *Moore's test.*—Add to the dextrose solution about half its volume of 20-per-cent. potash and heat. The solution becomes yellowish brown. Add to this some sulphuric acid (25 per cent.) and the odour of caramel becomes apparent.

6. *Fermentation test.*—Add a fragment of dried yeast to the dextrose solution in a test-tube; fill the test-tube up with mercury, and invert it over mercury in a trough. Place it in an incubator at body temperature for 24 hours. The sugar is broken up into alcohol and carbon dioxide; the latter gas collects in the upper part of the test-tube.

7. *Cane Sugar.*—(*a*) The solution of cane sugar when mixed with copper sulphate and caustic potash gives a blue solution. But on boiling no reduction occurs.

(*b*) Take some of the cane sugar solution and boil it with a few drops of 25-per-cent. sulphuric acid. This converts it into equal parts of dextrose and levulose. It then gives Trommer's or Fehling's test in the typical way.

(*c*) Boil some of the cane sugar solution with an equal volume of concentrated hydrochloric acid. A deep red solution is formed. Dextrose, lactose, and maltose do not give this test.

8. *Starch.*—(*a*) Examine microscopically the scrapings from the surface of a freshly cut potato. Note the appearance of the starch grains with their concentric markings.

(*b*) On boiling starch with water an opalescent solution is formed, which, if strong, gelatinises on cooling.

(*c*) Add iodine solution. An intense blue colour is produced, which disappears on heating, and if not heated too long reappears on cooling. N.B.—Prolonged heating drives off the iodine, and consequently no blue colour returns after cooling.

(*d*) Conversion into dextrin and dextrose. To some starch solution in a flask add a few drops of 25-per-cent. sulphuric acid, and boil for 15 minutes. Take some of the liquid, which is now clear, and show the presence of dextrin and dextrose.

9. *Dextrin.*—Add iodine solution to solution of dextrin; a reddish-brown colour is produced. The colour disappears on heating and reappears on cooling.

10. *Glycogen.*—Solution of glycogen is given round: (*a*) it is opalescent like that of starch.

(*b*) With iodine solution it gives a brown colour very like that given by dextrin. The colour disappears on heating and reappears on cooling.

(*c*) By boiling with 25-per-cent. sulphuric acid for 15 to 20 minutes it is converted into grape sugar.

The carbohydrates are found chiefly in vegetable tissues, and many of them form important foods. Some carbohydrates are, however, found in or formed by the animal organism. The most important of these are *glycogen*, or animal starch; *dextrose*; and *lactose*, or milk sugar.

The carbohydrates may be conveniently defined as compounds of carbon, hydrogen, and oxygen, the two last named elements being in the proportion in which they occur in water. But this definition is only a rough one, and if pushed too far would include many substances, like acetic acid, lactic acid, and inosite, which are not carbohydrates. Research has shown that the chemical constitution of the simplest carbohydrates is that of an aldehyde, or a ketone, and that the more complex carbohydrates are condensation products of the simple ones. In order, therefore, that we may understand the constitution of these substances, it is first necessary that we should understand what is meant by the terms aldehyde and ketone.

A primary alcohol is one in which the hydroxyl (OH) is attached to the last carbon atom of the chain; its end group is CH_2OH. Thus the formula for common alcohol (primary ethyl alcohol) is

$$CH_3.CH_2OH.$$

The formula for the next alcohol of the same series (primary propyl alcohol) is

$$CH_3.CH_2.CH_2OH.$$

If a primary alcohol is oxidised, the first oxidation product is called an *aldehyde*; thus ethyl alcohol yields acetic aldehyde:—

$$\underset{\text{[Ethyl alcohol]}}{CH_3.CH_2OH} + O = \underset{\text{[Acetic aldehyde]}}{CH_2.CHO} + H_2O.$$

The typical end-group CHO of the aldehyde is not stable, but is easily oxidisable to form the group COOH, and the compound so formed is called an acid; in this way acetic aldehyde forms acetic acid:—

$$\underset{\text{[acetic aldehyde]}}{CH_3.CHO} + O = \underset{\text{[acetic acid]}}{CH_3.COOH}.$$

The majority of the simple sugars are aldehydes of more complex alcohols than this; they are spoken of as *aldoses*. The readiness with which aldehydes are oxidisable renders them powerful reducing agents, and this furnishes us with some of the tests for the sugars.

Let us now turn to the case of the ketones. A secondary alcohol is one in which the OH group is attached to a central carbon atom; thus secondary propyl alcohol has the formula

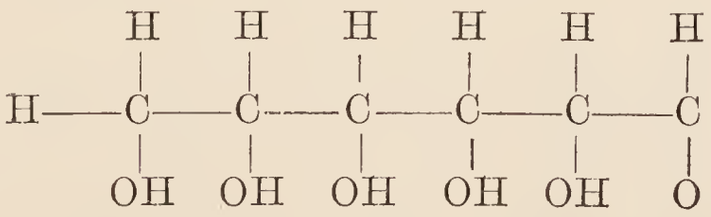
$$CH_3.CHOH.CH_3.$$

Its typical group is therefore CHOH. When this is oxidised, the first oxidation product is called a ketone, thus :—

$$\underset{\text{[secondary propyl alcohol]}}{CH_3.CHOH.CH_3} + O = \underset{\text{[propyl ketone]}}{CH_3.CO.CH_3} + H_2O.$$

It therefore contains the group CO in the middle of the chain. Some of the sugars are ketones of more complex alcohols; these are called *ketoses*. The only one of these which is of physiological interest is levulose.

The alcohols of which we have already spoken are called *monatomic*, because they contain only one OH group. Those which contain two OH groups (like glycol) are called diatomic; those which contain three OH groups (like glycerin) are called triatomic; and so on. The hexatomic alcohols are those which contain six OH groups. Three of these hexatomic alcohols with the formula $C_6H_8(OH)_6$ are of physiological interest; they are isomerides, and their names are sorbite, mannite, and dulcite. By careful oxidation their aldehydes and ketones can be obtained; these are the simple sugars; thus, dextrose is the aldehyde of sorbite; mannose is the aldehyde of mannite; levulose is the ketone of mannite; and galactose is the aldehyde of dulcite. These sugars all have the empirical formula $C_6H_{12}O_6$. The constitutional formula for dextrose is :—



By further oxidation, the sugars yield acids with various names. If we take such a sugar as a typical specimen, we see that their general formula is

$$C_nH_{2m}O_m$$

and as a general rule $n=m$; that is, the number of oxygen and carbon atoms are equal. This number in the case of the sugars already mentioned is six. Hence they are called *hexoses*.

Sugars are known to chemists, in which this number is 3, 4, 5, 7, &c., and these are called trioses, tetroses, pentoses, heptoses, &c. The majority of these have no physiological interest. It should, however, be mentioned that a pentose has been obtained from the nucleoproteid of the pancreas, of the liver, and of yeast. If the pentoses that are found in various plants are given to an animal, they are excreted in great measure unchanged in the urine.

The hexoses are of great physiological importance. The principal ones are dextrose, levulose, and galactose. These are called *monosaccharides*. Another important group of sugars are called *disaccharides*; these are formed by what is called condensation; that is, two molecules of monosaccharide combine together with the loss of a molecule of water, thus:—

$$C_6H_{12}O_6 + C_6H_{12}O_6 = C_{12}H_{22}O_{11} + H_2O.$$

The principal members of the disaccharide group are cane sugar, lactose, and maltose. If more than two molecules of the monosaccharide group undergo a corresponding condensation, we get what are called *polysaccharides*.

$$nC_6H_{12}O_6 = (C_6H_{10}O_5)_n + nH_2O.$$

The polysaccharides are starch, glycogen, various dextrins, cellulose, gums, &c. We may, therefore, arrange the important carbohydrates of the hexose family in a tabular form as follows:—

1. Monosaccharides or Glucoses, $C_6H_{12}O_6$	2. Disaccharides, Sucroses, or Saccharoses, $C_{12}H_{22}O_{11}$	3. Polysaccharides or Amyloses $(C_6H_{10}O_5)n$
+ Dextrose	+ Cane sugar.	+ Starch.
− Levulose.	+ Lactose.	+ Glycogen.
+ Galactose.	+ Maltose.	+ Dextrin.
		Cellulose.

The signs + and − in the above list indicate that the substances to which they are prefixed are dextro- and levo-rotatory respectively as regards polarised light.[1] The formulæ given above are merely empirical; the quantity n in the starch group is variable and often large. The following are the chief facts in relation to each of the principal carbohydrates.

[1] For a description of polarised light and polarimeters see Appendix. This and the other matter in the Appendix are placed there for convenience, not because they are unimportant. Students are therefore urged to refer to and carefully study these subjects.

MONOSACCHARIDES

Dextrose or Grape Sugar.—This carbohydrate is found in fruits, honey, and in minute quantities in the blood (0·12 per cent.) and numerous tissues, organs, and fluids of the body. It is the form of sugar found in large quantities in the blood and urine in the disease known as diabetes.

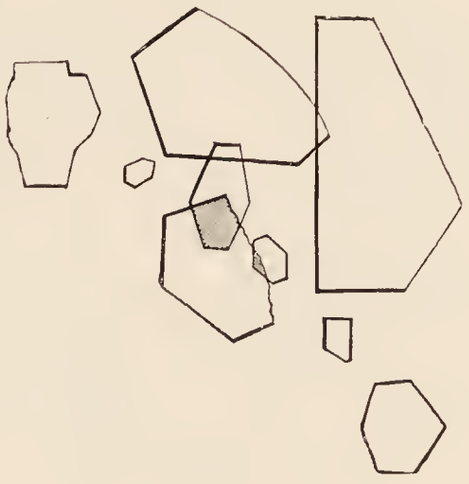


FIG. 1.—Dextrose crystals.

Dextrose is soluble in hot and cold water and in alcohol. It is crystalline (see fig. 1), but not so sweet as cane sugar. When heated with strong potash certain complex acids are formed which have a yellow or brown colour. This constitutes *Moore's test* for sugar. In alkaline solutions dextrose reduces salts of silver, bismuth, mercury, and copper. The reduction of cupric hydrate to cuprous hydrate or oxide constitutes *Trommer's test*, which has been already described at the head of the lesson. On boiling it with an alkaline solution of picric acid, a dark red opaque solution due to reduction of the picric to picramic acid is produced. Another important property of grape sugar is that under the influence of yeast it is converted into alcohol and carbonic acid ($C_6H_{12}O_6 = 2C_2H_6O + 2CO_2$).

Dextrose may be estimated by the fermentation test, by the polarimeter, and by the use of Fehling's solution. The last method is the most important; it rests on the same principles as Trommer's test, and we shall study it and other methods of estimating sugar in connection with diabetic urine (see Lesson XII.).

Levulose.—When cane sugar is treated with dilute mineral acids it undergoes a process known as inversion—*i.e.* it takes up water and is converted into equal parts of dextrose and levulose. The previously dextro-rotatory solution of cane sugar then becomes levo-rotatory, the levo-rotatory power of the levulose being greater than the dextro-rotatory power of the dextrose formed. Hence the term *inversion*. The same hydrolytic change is produced by certain ferments, such as the invert ferment of the intestinal juice, and of yeast.

Pure levulose can be crystallised, but so great is the difficulty of obtaining crystals of it that one of its names was 'uncrystallisable sugar.' Small quantities of levulose have been found in blood, urine, and muscle. It has been recommended as an article of diet in diabetes in place of ordinary sugar; in this disease it does not

appear to have the harmful effect that other sugars produce. Levulose gives the same general reactions as dextrose.

Galactose is formed by the action of dilute mineral acids or inverting ferments on lactose or milk sugar. It resembles dextrose in being dextro-rotatory, in reducing cupric hydrate in Trommer's test, and in being directly fermentable with yeast. When oxidised by means of nitric acid it, however, yields an acid called mucic acid ($C_6H_{10}O_8$), which is only sparingly soluble in water. Dextrose when treated in this way yields an isomeric acid—*i.e.* an acid with the same empirical formula, called saccharic acid, which is readily soluble in water.

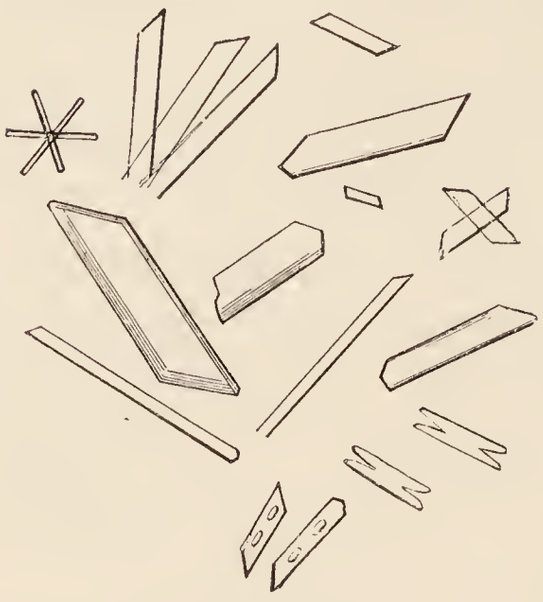


Fig. 2.—Inosite crystals.

Inosite, formerly called muscle sugar, is found in muscle, kidney, liver, and other parts of the body in small quantities. It is also largely found in the vegetable kingdom. It is a crystallisable substance (see fig. 2) and has the same formula ($C_6H_{12}O_6$) as the glucoses. It is, however, not a sugar. It gives none of the sugar tests, and careful analysis has shown it has quite a different chemical constitution from the true sugars. It belongs to the aromatic series, and is only included here for convenience.

DISACCHARIDES

Cane Sugar.—This sugar is generally distributed throughout the vegetable kingdom in the juices of plants and fruits, especially the sugar cane, beetroot, mallow, and sugar maple. It is a substance of great importance as a food. After abundant ingestion of cane sugar traces may appear in the urine, but the greater part undergoes inversion in the alimentary canal.

Pure cane sugar is crystalline and dextro-rotatory. It holds cupric hydrate in solution in an alkaline liquid—that is, with Trommer's test it gives a blue solution. But no reduction occurs on boiling. After inversion it is strongly reducing.

Inversion may be brought about readily by boiling with dilute mineral acids, or by means of an inverting ferment, such as that occurring in the succus entericus or intestinal juice. It then takes up water and is split into equal parts of dextrose and levulose (see p. 17).

$$\underset{\text{[cane sugar]}}{C_{12}H_{22}O_{11}} + H_2O = \underset{\text{[dextrose]}}{C_6H_{12}O_6} + \underset{\text{[levulose]}}{C_6H_{12}O_6}$$

With yeast, cane sugar is first inverted by means of a special soluble ferment produced by the yeast cells, and then there is an alcoholic fermentation of the glucoses so formed.

Lactose, or **milk sugar**, occurs in milk. It has also been described as occurring in the urine of women in the early days of lactation or after weaning.

It crystallises in rhombic prisms (see fig. 3). It is much less soluble in water than cane sugar or dextrose, and has only a slightly sweet taste. It is insoluble in alcohol and ether; aqueous solutions are dextro-rotatory.

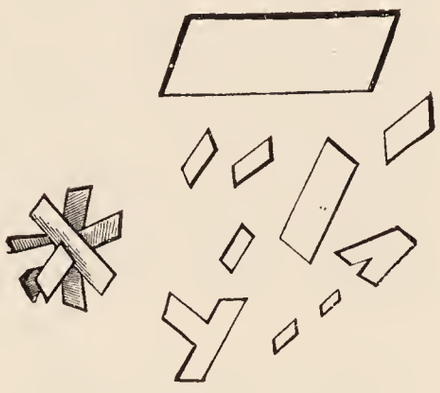


Fig. 3.—Milk-sugar crystals.

Solutions of lactose give Trommer's test, but when the reducing power is tested quantitatively by Fehling's solution it is found to be a less powerful reducing agent than dextrose. If it required seven parts of a solution of dextrose to reduce a given quantity of Fehling's solution, it would require ten parts of a solution of lactose of the same strength to reduce the same quantity of Fehling's solution.

Lactose, like cane sugar, can be hydrolysed by the same agencies as those already enumerated in connection with cane sugar. The glucoses formed are dextrose and galactose.

$$\underset{\text{[lactose]}}{C_{12}H_{22}O_{11}} + H_2O = \underset{\text{[dextrose]}}{C_6H_{12}O_6} + \underset{\text{[galactose]}}{C_6H_{12}O_6}$$

With yeast it is first inverted, and then alcohol is formed. This, however, occurs slowly.

With the lactic-acid organisms which bring about the souring of milk the lactic-acid fermentation is produced. This may also occur as the result of the action of putrefactive bacteria in the alimentary canal. The two stages of the lactic-acid fermentation are represented by the following equations:—

(1) $$\underset{\text{[lactose]}}{C_{12}H_{22}O_{11}} + H_2O = 4\underset{\text{[lactic acid]}}{C_3H_6O_3}$$

(2) $$\underset{\text{[lactic acid]}}{4C_3H_6O_3} = \underset{\text{[butyric acid]}}{2C_4H_8O_2} + 4CO_2 + 4H_2$$

Maltose is the chief end product of the action of malt diastase on starch, and is also formed as an intermediate product in the action of dilute sulphuric acid on the same substance. It is also the chief sugar formed from starch by the diastatic ferments contained in the saliva (ptyalin) and pancreatic juice (amylopsin). It can be obtained

in the form of acicular crystals; it is strongly dextro-rotatory. It gives Trommer's test; but its reducing power, as measured by Fehling's solution, is one-third less than that of dextrose.

By prolonged boiling with water, or, more readily, by boiling with a dilute mineral acid, or by means of an inverting ferment, such as occurs in the intestinal juice, it is converted into dextrose.

$$\underset{\text{[maltose]}}{C_{12}H_{22}O_{11}} + H_2O = \underset{\text{[dextrose]}}{C_6H_{12}O_6} + \underset{\text{[dextrose]}}{C_6H_{12}O_6}$$

It undergoes readily the alcoholic fermentation.

The three important physiological sugars (dextrose, lactose, and maltose) may be distinguished from one another by their relative reducing action on Fehling's solution (1·0 : 0·71 : 0·63), by their rotatory power, or by the phenyl-hydrazine test described in Lesson XIII.

POLYSACCHARIDES

Starch is widely diffused through the vegetable kingdom. It occurs in nature in the form of microscopic grains, varying in size and appearance, according to their source. Each consists of a central spot (*hilum*) round which more or less concentric envelopes of starch proper or granulose alternate with layers of cellulose. Cellulose has very little digestive value, but starch is a most important food.

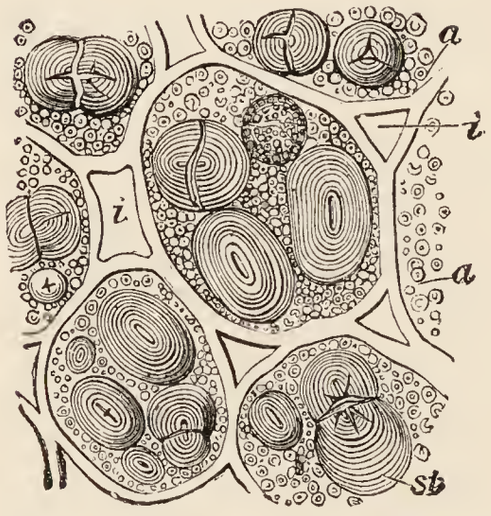


Fig. 4.—Section of pea showing starch and aleurone grains embedded in the protoplasm of the cells: *a*, aleurone grains; *st*, starch grains; *i*, intercellular spaces. (Yeo, after Sachs.)

Starch is insoluble in cold water; it forms an opalescent solution in boiling water, which if concentrated gelatinises on cooling. Its most characteristic reaction is the blue colour it gives with iodine.

On heating starch with dilute mineral acids dextrose is formed. By the action of diastatic ferments, maltose is the chief end product. In both cases dextrin is an intermediate stage in the process.

Before the formation of dextrin the starch solution loses its opalescence, a substance called *soluble starch* or *amidulin* being formed. This, like native starch, gives a blue colour with iodine. Although the molecular weight of starch is unknown, the formula for soluble starch is probably $(C_6H_{10}O_5)_{200}$. Equations that represent the formation of sugars and dextrins from this are very complex, and are at present hypothetical.

Dextrin is the name given to the intermediate products in the hydration of starch, and two chief varieties are distinguished—*erythro-dextrin*, which gives a reddish-brown colour with iodine; and *achroö-dextrin*, which does not.

It is readily soluble in water, but insoluble in alcohol and ether. It is gummy and amorphous. It does not give Trommer's test, nor does it ferment with yeast. It is dextro-rotatory. By hydrating agencies it is converted into glucose.

Glycogen, or animal starch, is found in liver, muscle, and colourless blood corpuscles. It is also abundant in all embryonic tissues.

Glycogen is a white tasteless powder, soluble in water, but it forms, like starch, an opalescent solution. It is insoluble in alcohol and ether. It is dextro-rotatory. With Trommer's test it gives a blue solution, but no reduction occurs on boiling.

With iodine it gives a reddish or port-wine colour, very similar to that given by erythro-dextrin. Dextrin may be distinguished from glycogen by (1) the fact that it gives a clear, not an opalescent, solution with water; and (2) it is not precipitated by basic lead acetate as glycogen is. It is, however, precipitated by basic lead acetate and ammonia. (3) Glycogen is precipitated by 55 per cent. of alcohol; the dextrins require 85 per cent. or more.

Cellulose.—This is the colourless material of which the cell-walls and woody fibres of plants are composed. By treatment with strong mineral acids, it is, like starch, converted into glucose, but with much greater difficulty. The various digestive ferments have little or no action on cellulose; hence the necessity of boiling starch before it is taken as food. Boiling bursts the cellulose envelope of the starch grains, and so allows the digestive juice to get at the starch proper. Cellulose is found in a few animals, as in the test or outer investments of the Tunicates.

Salting out of the Colloid Carbohydrates.—By saturating solutions of the colloid carbohydrates (starch, soluble starch, glycogen, and some varieties of dextrin) with such neutral salts as magnesium sulphate or ammonium sulphate the carbohydrate is thrown out of solution in the form of a white precipitate. The remaining carbohydrates (sugars and some of the smaller moleculed dextrins like achroö-dextrin) are not precipitated by this means. We shall find in connection with the proteids that this method known as 'salting out' is one largely employed there for precipitating and distinguishing between classes of proteids. The student is therefore warned that a precipitate obtained under such circumstances will not necessarily indicate the presence of proteid.

Further information regarding the carbohydrates is given in Lesson XIII.

LESSON III

THE FATS

Lard and olive oil are given round as examples of fats.

1. They are insoluble in water.

2. They dissolve readily in ether. On pouring some of the ethereal solution on to a piece of blotting paper, a greasy stain is left after the ether has evaporated.

3. *Saponification.*—By boiling with potash fat yields a solution of soap. On adding some sulphuric acid to this and heating, the fatty acid collects in a layer on the surface of the fluid. This experiment may conveniently be performed in the following way:—Melt some lard in an evaporating basin and pour it into a solution of potash in alcohol[1] contained in a small flask and heated carefully *on a water bath* nearly to boiling point. Continue to boil and saponification is soon completed. When the process is completed drop some of the solution into a test-tube containing about 10 c.c. of water; the solution of soap will be clear, and no oil globules should separate out. If there is any separation of oil globules, continue the boiling.

Then drop the soap solution into some 25-per-cent. sulphuric acid contained in a small beaker and heated nearly to boiling; the fatty acid soon separates out and floats on the surface.

4. *Reaction of Fatty Acids.*—(*a*) They produce a greasy stain on paper.

(*b*) Wash the fatty acid obtained in experiment 3 repeatedly with water, until the wash water is no longer acid, and divide it into two portions. Dissolve one portion in ether; this solution reacts acid to phenolphthalein; to show this, place a few drops of phenolphthalein in 5 c.c. of water containing a drop of 20-per-cent. potash. If this red solution is dropped into the solution of fatty acid, the colour is discharged. Place the second portion of fatty acid in some half saturated solution of sodium carbonate and warm; a solution of sodium soap is obtained and carbon dioxide comes off.

5. *Separation of Neutral Fats from Fatty Acids.*—In most fats some free-fatty acid is present. They may be separated by the fact that the latter only dissolves in a solution of sodium carbonate to form soap. The resulting mass is shaken with water and then with ether; the two fluids separate on standing; the ether contains the neutral fat and the water the soap.

6. *Test for Glycerin.*—The most important reaction for glycerin, the other constituent of a fat, is the acrolein test, which is performed in the following way:—Place some lard in a dry test-tube, add a crystal of potassium sulphate and heat. Acrolein is given off, which is recognised by its characteristic unpleasant odour, and by the fact that it blackens a piece of filter paper previously moistened with ammoniacal silver nitrate solution.

7. *Osmic Acid Test.*—Fat, if it contains olein or oleic acid, is blackened by osmic acid. Try this with both the lard and the olive oil.

8. *To determine melting point of a fat or fatty acid*, place a small

[1] 30 grammes of potash are dissolved in 20 c.c. of water, and 200 c.c. of 90 per cent. alcohol added.

quantity in a very narrow test-tube strapped on to a thermometer with an india-rubber band. Place this in a water bath which is gradually heated, and note the temperature at which it melts.

9. *Emulsification.*—(*a*) Take two test-tubes and label them A and B. Place water in A and soap solution in B. To each add a few drops of olive oil and shake. In B an emulsion is formed, but not in A.

(*b*) Shake a few drops of rancid oil with a dilute solution of potash; an emulsion is formed because the potash and free-fatty acid unite to form a soap. Divide this into two parts, and to one of them add a little solution of gum or egg albumin; the emulsion is much more permanent in this specimen. These experiments illustrate the favourable action of soap and of a *suspending medium* like mucilage upon the formation of an emulsion.

Fat is found in small quantities in many animal tissues. It is, however, found in large quantities in three situations, viz. bone



FIG. 5.—A few cells from the margin of a fat lobule: *f. g.*, fat globule distending fat cell; *n*, nucleus; *m*, membranous envelope of the cell; *cr.*, bunch of crystals within a fat cell; *c*, capillary vessel; *v*, venule; *c. t.*, connective tissue cell. The fibres of the connective tissue are not represented.

marrow, adipose tissue, and milk. The consideration of the fat in milk is postponed to Lesson VI.

The contents of the fat cells of adipose tissue are fluid during life, the normal temperature of the body (37° C., or 99° F.) being considerably above the melting-point (25° C.) of the mixture of the fats found there. These fats are three in number, and are called palmitin, stearin, and olein. They differ from one another in chemical composition and in certain physical characters, such as melting-point and solubilities. Olein melts at −5° C., palmitin at 45° C., and stearin at 53–65° C. Thus, it is olein which holds the other two dissolved at

the body temperature. Fats are all soluble in hot alcohol, ether, and chloroform, but insoluble in water.

Chemical Constitution of the Fats.—The fats are compounds of fatty acids with glycerin, and may be termed glycerides or glyceric ethers. The term *hydrocarbon*, applied to them by some authors, is wholly incorrect.

The fatty acids form a series of acids derived from the monatomic alcohols by oxidation. Thus, to take ordinary ethyl alcohol, C_2H_5HO, the first stage in oxidation is the removal of two atoms of hydrogen to form aldehyde, $CH_3.COH$: on further oxidation an atom of oxygen is added to form acetic acid, $CH_3.COOH$ (see also p. 14).

A similar acid can be obtained from all the other alcohols thus :

From	methyl alcohol	$CH_3.HO$,	formic acid	$H.COOH$	is obtained	
,,	ethyl ,,	$C_2H_5.HO$,	acetic ,,	$CH_3.COOH$	,,	
,,	propyl ,,	$C_3H_7.HO$,	propionic ,,	$C_2H_5.COOH$	,,	
,,	butyl ,,	$C_4H_9.HO$,	butyric ,,	$C_3H_7.COOH$	,,	
,,	amyl ,,	$C_5H_{11}.HO$,	valeric ,,	$C_4H_9.COOH$	,,	
,,	hexyl ,,	$C_6H_{13}.HO$,	caproic ,,	$C_5H_{11}.COOH$	,,	

and so on.

Or in general terms :—

From the alcohol with formula $C_nH_{2n+1}.HO$ the acid with formula $C_{n-1}H_{2n-1}.CO.OH$ is obtained. The sixteenth term of this series has the formula $C_{15}H_{31}.CO.OH$, and is called **palmitic acid**; the eighteenth has the formula $C_{17}H_{35}.CO.OH$, and is called **stearic acid**. Each acid, as will be seen, consists of a radicle, $C_{n-1}H_{2n-1}CO$, united to hydroxyl (HO).

Oleic acid, however, is not a member of the fatty acid series proper, but belongs to a somewhat similar series of acids known as the acrylic series, of which the general formula is $C_{n-1}H_{2n-3}COOH$. It is the eighteenth term of the series, and its formula is $C_{17}H_{33}.CO.OH$.

The first member of the group of alcohols from which this acrylic series of acids is obtained is called *allyl alcohol* (CH_2 : $CH.CH_2OH$) ; the aldehyde of this is *acrolein* (CH_2 : CH.CHO), and the formula for the acid (acrylic acid) is CH_2 : CH.COOH. It will be noticed that two of the carbon atoms are united by two valencies, and these bodies are therefore unsaturated; they are unstable and are prone to undergo by uniting with another element a conversion into bodies in which the carbon atoms are united by only one bond. This accounts for their reducing action, and it is owing to this construction that the colour reactions with osmic acid and Sudan III. are due. Fat which contains any member of the acrylic series like oleic acid blackens osmic acid, by reducing it to a lower (black) oxide. Fats like palmitin and stearin do not give this reaction.

Glycerin or **Glycerol** is a triatomic alcohol, $C_3H_5(OH)_3$—*i.e.* three atoms of hydroxyl united to a radicle glyceryl (C_3H_5). The hydrogen in the hydroxyl atoms is replaceable by other organic radicles. As an example take the radicle of acetic acid called acetyl ($CH_3.CO$). The following formulæ represent the derivatives that can be obtained by replacing one, two, or all three hydroxyl hydrogen atoms in this way :—

$$C_3H_5\begin{cases}OH\\OH\\OH\end{cases} \qquad C_3H_5\begin{cases}OH\\OH\\O.CH_3.CO\end{cases} \qquad C_3H_5\begin{cases}OH\\O.CH_3.CO\\O.CH_3.CO\end{cases} \qquad C_3H_5\begin{cases}O.CH_3.CO\\O.CH_3.CO\\O.CH_3.CO\end{cases}$$

[glycerin] [monoacetin] [diacetin] [triacetin]

Triacetin is a type of a neutral fat; stearin, palmitin, and olein ought more properly to be called tristearin, tripalmitin, and triolein respectively. Each consists of glycerin in which the three atoms of hydrogen in the hydroxyls are replaced by radicles of the fatty acid. This is represented in the following formulæ :—

	Acid		*Radicle*		*Fat*
Palmitic acid	$C_{15}H_{31}.COOH$	Palmityl	$C_{15}H_{31}.CO$	Palmitin	$C_3H_5(OC_{15}H_{31}.CO)_3$
Stearic acid	$C_{17}H_{35}.COOH$	Stearyl	$C_{17}H_{35}.CO$	Stearin	$C_3H_5(OC_{17}H_{35}.CO)_3$
Oleic acid	$C_{17}H_{33}.COOH$	Oleyl	$C_{17}H_{33}.CO$	Olein	$C_3H_5(OC_{17}H_{33}.CO)_3$

Decomposition Products of the Fats.—The fats split up into the substances out of which they are built up.

Under the influence of superheated steam, mineral acids, and in the body by means of certain ferments (for instance, the fat-splitting ferment steapsin of the pancreatic juice), a fat combines with water and splits into glycerin and the fatty acid. The following equation represents what occurs in a fat, taking tripalmitin as an example :—

$$\underset{\text{[palmitin—a fat]}}{C_3H_5(O.C_{15}H_{31}CO)_3} + 3H_2O = \underset{\text{[glycerin]}}{C_3H_5(OH)_3} + \underset{\text{[palmitic acid—a fatty acid]}}{3C_{15}H_{31}CO.OH}$$

In the process of saponification, much the same sort of reaction occurs, the final products being glycerin and a compound of the base with the fatty acid, which is called a soap. Suppose, for instance, that potassium hydrate is used ; we get—

$$\underset{\text{[palmitin—a fat]}}{C_3H_5(O.C_{15}H_{31}CO)_3} + 3KHO = \underset{\text{[glycerin]}}{C_3H_5(OH)_3} + \underset{\text{[potassium palmitate—a soap]}}{3C_{15}H_{31}CO.OK}$$

Emulsification.—Another change that fats undergo in the body is very different from saponification. It is a physical rather than a chemical change ; the fat is broken up into very small globules, such as are seen in the natural *emulsion*—milk.

Lecithin ($C_{42}H_{84}NPO_9$).—This is a very complex fat, which

yields on decomposition not only glycerin and a fatty acid, but phosphoric acid, and an alkaloid [$N.(CH_3)_3C_2H_6O_2$] called *choline* in addition. Lecithin is found to a great extent in the nervous system,[1] and to a small extent in bile. Together with **cholesterin**, a crystallisable, monatomic alcohol ($C_{27}H_{45}.HO$) which we shall consider more at length in connection with the bile, it is found in small quantities in the protoplasm of all cells.

[1] See further under Nervous Tissues, Lesson XXII.

LESSON IV

THE PROTEIDS

1. **Tests for Proteids.**—The following tests are to be tried with a mixture of one part of white of egg to ten of water. (Egg-white contains a mixture of albumin and globulin.)

(*a*) *Heat Coagulation.*—Faintly acidulate with a few drops of 2-per-cent. acetic acid and boil. The proteid is rendered insoluble (coagulated proteid).

(*b*) *Precipitation with Nitric Acid.*—The addition of strong nitric acid to the original solution also produces a white precipitate.

(*c*) *Xanthoproteic Reactions.*—On boiling the white precipitate produced by nitric acid it turns yellow; after cooling add ammonia; the yellow becomes orange.

(*d*) *Millon's Test.*—Millon's reagent (which is a mixture of the nitrates of mercury containing excess of nitric acid; see p. 5) gives a white precipitate, which turns brick-red on boiling.

(*e*) After the addition of a few drops of 20-per-cent. acetic acid, potassium ferrocyanide gives a white precipitate.

(*f*) *Piotrowski's Test.*—Add *one drop* of a 1-per-cent. solution of cupric sulphate to the original solution and then caustic potash, and a violet solution is obtained.

Repeat experiment (*f*) with a solution of commercial peptone, and note that a rose-red solution is obtained. This is called the *biuret reaction.*

(*g*) *Adamkiewicz's Reaction* (Hopkins's modification).—Add the albumin solution to dilute glyoxylic acid, and then strong sulphuric acid; an intense violet colour is obtained. Most commercial specimens of glacial acetic acid contain glyoxylic acid and may be used in place of pure glyoxylic acid in this test.

2. **Action of Neutral Salts.**—(*a*) Saturate the solution of egg-white with magnesium sulphate by adding crystals of the salt and grinding it up thoroughly in a mortar. A white precipitate of egg-globulin is produced. Filter. The filtrate contains egg-albumin. The precipitate of the globulin is very small.

(*b*) Half saturate the solution of egg-white with ammonium sulphate. This may be done by adding to the solution an equal volume of a saturated solution of ammonium sulphate. The precipitate produced consists of the globulin; the albumin remains in solution.

(*c*) Completely saturate another portion with ammonium sulphate by adding crystals of the salt and grinding in a mortar—a precipitate is produced of both the globulin and albumin. Filter. The filtrate contains no proteid.

(*d*) Repeat the last experiment (*c*) with a solution of commercial peptone. A precipitate is produced of the albumoses or proteoses it contains. Filter. The filtrate contains the true peptone. This gives the biuret reaction (see above), but large excess of strong potash must be added on account of the presence of ammonium sulphate. *Ammonium sulphate added to saturation precipitates all proteids except peptone.*

LESSON V

THE PROTEIDS (continued)

1. *Action of Acids and Alkalis on Albumin.*—Take three test-tubes and label them A, B, and C.

In each place an equal amount of diluted egg-white, similar to that used in the last lesson.

To A add a few drops of 0·1-per-cent. solution of caustic potash.

To B add the same amount of 0·1-per-cent. solution of caustic potash.

To C add a rather larger amount of 0·1-per-cent. sulphuric acid.

Put all three into the warm bath[1] at about the temperature of the body (36–40° C.).

After five minutes remove test-tube A, and boil. The proteid is no longer coagulated by heat, having been converted into *alkali-albumin*. After

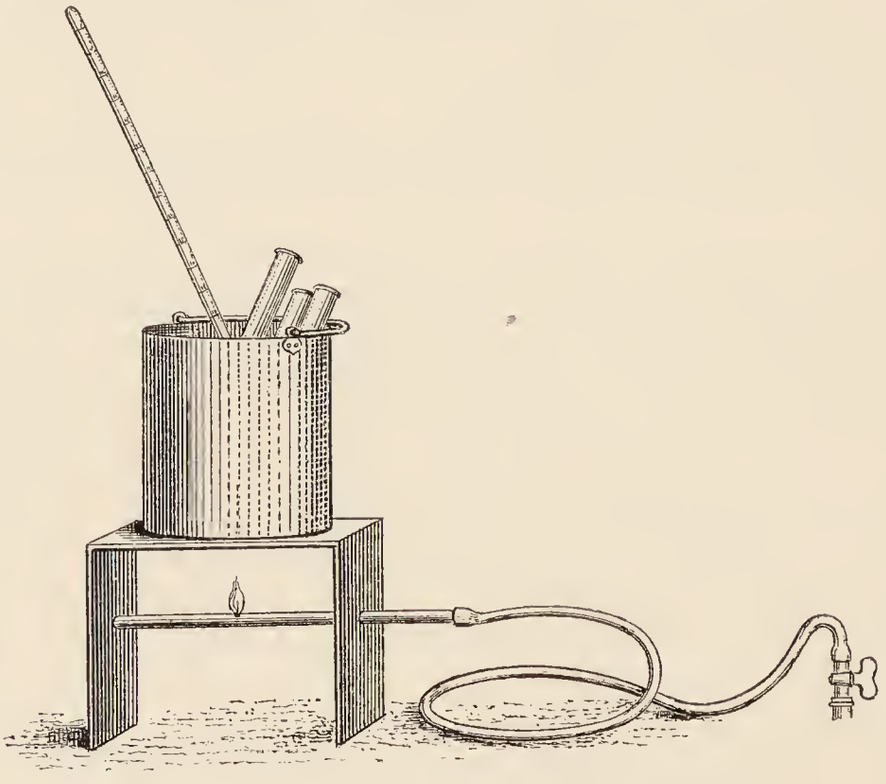


FIG. 6.—Simple warm bath, as described in footnote.

cooling, colour with litmus solution and neutralise with 0·1-per-cent. acid. At the neutral point a precipitate is formed which is soluble in excess of either acid or alkali.

Next remove B. This also now contains alkali-albumin. Add to it a few drops of sodium phosphate, colour with litmus, and neutralise as before.

[1] A convenient form of warm bath suitable for class purposes may be made by placing an ordinary tin pot half full of water over a bent piece of iron which acts as a warm stage as in the figure. The stage is kept warm by a small gas flame. Such a warm bath may be placed between every two or three students.

Note that the alkali-albumin now requires more acid for its precipitation than in A, the acid which is first added converting the sodium phosphate into acid sodium phosphate.

Now remove C from the bath. Boil it. Again there is no coagulation, the proteid having been converted into *acid-albumin*, or *syntonin*. After cooling, colour with litmus and neutralise with 0·1-per-cent. alkali. At the neutral point a precipitate is formed, soluble in excess of acid or alkali. (Acid-albumin is formed more slowly than alkali-albumin, so it is best to leave this experiment to the last.)

2. Take some gelatin and dissolve it in hot water. On cooling, the solution sets into a jelly (gelatinisation).

Take a dilute solution of gelatin, and try all the proteid tests with it enumerated on p. 27. Carefully note down your results.

3. Add a few drops of acetic acid to some saliva. A stringy precipitate of mucin is formed.

4. A tendon has been soaked for a few days in lime water. The fibres are not dissolved, but they are loosened from one another owing to the solution of the interstitial or ground substance by the lime water. Take some of the lime-water extract and add acetic acid. A precipitate of mucin is obtained. The fibres themselves consist of collagen, which yields gelatin on boiling. Vitreous humour or the Whartonian jelly of the umbilical cord is much richer in ground substance than tendon, and, if treated in the same way, a much larger yield of mucin is obtained.

The **Proteids** are the most important substances that occur in animal and vegetable organisms; none of the phenomena of life occur without their presence; and though it is impossible to state positively that they occur as such in living protoplasm, they are invariably obtained by subjecting living structures to analytical processes.

Proteids are highly complex compounds of carbon, hydrogen, oxygen, nitrogen, and sulphur, occurring in a solid viscous condition or in solution in nearly all parts of the body. The different members of the group present differences in chemical and physical properties. They all possess, however, certain common chemical reactions, and are united by a close genetic relationship.

The various proteids differ a good deal in elementary composition. Hoppe-Seyler gives the following percentages:—

	C	H	N	S	O
From	51·5	6·9	15·2	0·3	20·9
To	54·5	7·3	17·0	2·0	23·5

We are, however, not acquainted with the constitutional formula of proteid substances. There have been many theories on the subject, but practically all that is known with certainty is that many different substances may be obtained by the decomposition of proteids. How they are built up into the proteid molecule is unknown. The decompositions that occur in the body are, moreover, different from those which can be made to occur in the laboratory; hence the

conclusion that living protoplasm differs somewhat from the non-living proteid material obtainable from it.

(1) In the body. Carbonic acid, water, and urea[1] are the chief final products. Glycine, leucine, creatine, uric acid, ammonia, &c., are probably intermediate products. Carbohydrate (glycogen) and fats may also originate from proteids.

(2) Outside the body. Various strong reagents break up proteids into ammonia, carbonic acid, amines, fatty acids, amino- acids like leucine, arginine, and glycine, and aromatic compounds like tyrosine.

TESTS FOR PROTEIDS

Solubilities.—All proteids are insoluble in alcohol and ether. Some are soluble in water, others insoluble. Many of the latter are soluble in weak saline solutions. Some are insoluble, others soluble in concentrated saline solutions. It is on these varying solubilities that proteids are classified.

All proteids are soluble with the aid of heat in concentrated mineral acids and alkalis. Such treatment, however, decomposes as well as dissolves the proteid. Proteids are also soluble in gastric and pancreatic juices; but there, again, they undergo a change, being converted into a hydrated variety of proteid of smaller molecular weight called *peptone*. The intermediate substances formed in this process are called *proteoses* or *albumoses*. Commercial peptone contains a mixture of proteoses and true peptone.

Heat Coagulation.—Many of the proteids which are soluble in water or saline solution are rendered insoluble when those solutions are heated. This is true for most of the proteids that occur in nature. The solidifying of white of egg when heated is a familiar instance of this. The temperature of heat coagulation differs in different proteids: thus myosinogen and fibrinogen coagulate at about 56° C.; serum albumin and serum globulin at about 75° C.

The proteids which are coagulated by heating their solution come for the most part into two classes—the *albumins* and the *globulins*. The full distinction between these we shall see immediately. We may, however, state here that the albumins are soluble in distilled water; the true globulins are not, but require salts to hold them in solution.

Indiffusibility.—The proteids (peptones excepted) belong to the class of substances called *colloids* by Thomas Graham; that is, they

[1] Recent research has shown that urea can be also obtained from proteids by analytical methods outside the body (see under URINE).

pass with difficulty, or not at all, through animal membranes. In the construction of dialysers, vegetable parchment is very largely used (see figs. 7 and 8).

Proteids may thus be separated from diffusible (*crystalloid*) substances like salts, but the process is a somewhat tedious one. If some serum or white of egg is placed in a dialyser, and distilled water outside, the greater amount of the salts passes into the water through the membrane; the two proteids, albumin and globulin,

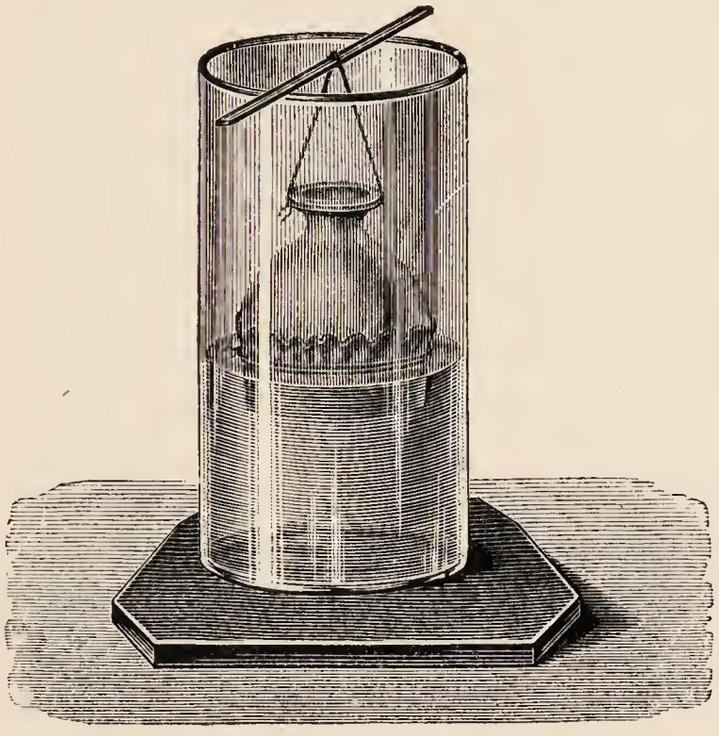


Fig. 7.—Dialyser. The lower opening of the bell jar suspended in water is tightly covered with parchment paper. The fluid to be dialysed is placed within this vessel; the crystalloids pass out into the distilled water outside through the parchment paper.

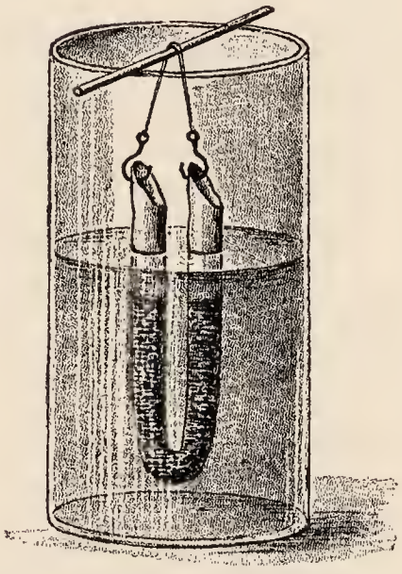


Fig. 8.—In this form of dialyser the substance to be dialysed is placed within the piece of tubing suspended in the larger vessel of water. The tubing is made of parchment paper.

remain inside. The globulin is, however, precipitated, as the salts which previously kept it in solution have been removed.

The terms 'diffusion' and 'dialysis' should be distinguished from each other.

If water is carefully poured on the surface of a solution of any substance, this substance gradually spreads through the water, and the composition of the mixture becomes uniform in time. The time occupied is short for substances like sodium chloride, and long for substances like albumin. The phenomenon is called *diffusion*. If the solutions are separated by a membrane the term 'dialysis' is employed. The word *osmosis* is properly restricted to the passage of water through membranes, and can be best studied when semi-permeable membranes are employed. See fully article Osmosis in Appendix.

Crystallisation.—Hæmoglobin, the red pigment of the blood, is a proteid substance, and is crystallisable (for further details, see The Blood). Like other proteids, it has an enormously large molecule; though crystalline, it is not crystalloid in Graham's sense of

that term. Blood pigment is not the only crystallisable proteid. Long ago crystals of proteid (globulin or vitellin) were observed in the aleurone grains of many seeds, and in the similar proteid occurring in the egg-yolk of some fishes and amphibians. By appropriate methods these have been separated and re-crystallised. Further, egg-albumin itself has been crystallised. If a solution of white of egg is diluted with half its volume of saturated solution of ammonium sulphate, the globulin present is precipitated and is removed by filtration. The filtrate is now allowed to remain some days at the temperature of the air, and as it becomes more concentrated from evaporation, minute spheroidal globules and finally minute needles, either aggregated or separate, make their appearance (Hofmeister). Crystallisation is much more rapid and perfect if a little acetic or sulphuric acid is added (Hopkins). Serum albumin (from horse and rabbit) has also been similarly crystallised (Gürber).

Action on Polarised Light.—All the proteids are levo-rotatory, but the amount of rotation they produce varies with the kind of proteid. See Appendix. Several of the compound proteids (for instance, hæmoglobin and nucleo-proteids) are dextro-rotatory, though their proteid components are levo-rotatory (Gamgee).

Colour Reactions.—The principal colour reactions have been already described in the heading of this lesson. The xanthoproteic and Millon's reactions depend on the presence of an aromatic radicle in the proteid molecule. Peptones behave differently from the native proteids in the copper sulphate test. They give a rose-red colour instead of a violet, if only a trace of copper sulphate is used. The albumoses act in this respect like the peptones. This rose-red colour is also given by the substance called biuret ; [1] hence the test is called the *biuret reaction.* This name does not imply that biuret is present in proteid, but both proteid and biuret give the reaction because they possess a common radicle, probably CONH. The Adamkiewicz reaction is due to the presence in proteid of the tryptophan radicle (skatole-amino-acetic acid). See further under pancreatic digestion.

Precipitants of Proteids.—Proteids are precipitated by a large number of reagents ; the peptones and albumoses are exceptions in many cases, and will be considered separately afterwards (see Lesson VII.).

[1] Biuret is formed by heating solid urea ; ammonia passes off and leaves biuret thus :

$$\underset{\text{[urea]}}{2CON_2H_4} - \underset{\text{[ammonia]}}{NH_3} = \underset{\text{[biuret].}}{C_2O_2N_3H_5}$$

Solutions of the proteids are precipitated by—

1. Strong acids, like nitric acid.
2. Picric acid.
3. Acetic acid and potassium ferrocyanide.
4. Acetic acid and excess of neutral salts like sodium sulphate.
5. Salts of the heavy metals, like copper sulphate, mercuric chloride, lead acetate, silver nitrate, &c.
6. Tannin.
7. Alcohol.
8. Saturation with certain neutral salts, such as ammonium sulphate.

It is necessary that the words *coagulation* and *precipitation* should, in connection with the proteids, be carefully distinguished. The term *coagulation* is used when an insoluble proteid (coagulated proteid) is formed from a soluble one. This may occur—

1. When the proteid is heated—*heat coagulation.*
2. Under the influence of a ferment; for instance, when a curd is formed in milk by rennet or a clot in shed blood by the fibrin ferment —*ferment coagulation.*
3. When an insoluble precipitate is produced by the addition of certain reagents (nitric acid, picric acid, tannin, &c.).

There are, however, other precipitants of proteids in which the precipitate formed is readily soluble in suitable reagents, like saline solution, and the proteid continues to show its typical reactions. This precipitation is not coagulation. Such a precipitate is produced by saturation with ammonium sulphate. Certain proteids, called globulins, are more readily precipitated by such means than others. Thus, serum globulin is precipitated by half-saturation with ammonium sulphate. Full saturation with ammonium sulphate precipitates all proteids but peptone. The globulins are precipitated by certain salts like sodium chloride and magnesium sulphate, which do not precipitate the albumins. The precipitation of proteids by salts in this way is conveniently termed 'salting out.'

The precipitation produced by alcohol is peculiar in that after a time it becomes a coagulation. Proteid freshly precipitated by alcohol is readily soluble in water or saline media; but after it has been allowed to stand some weeks under alcohol it becomes more and more insoluble. Albumins and globulins are most readily rendered insoluble by this method; albumoses and peptones are apparently never rendered insoluble by the action of alcohol. This fact is of value in the separation of these proteids from others.

CLASSIFICATION OF PROTEIDS

Proteids may be of animal or vegetable origin, and both animal and vegetable proteids may be subdivided in the same way. We shall, however, be chiefly concerned with the animal proteids.

If we use the term 'proteid' in the widest sense, the first main subdivision of these substances is into

A. The simple proteids.

B. The compound or conjugated proteids.

C. The albuminoids.

D. The protamines. These substances are regarded by Kossel as the simplest proteids, and though it is doubtful whether this view will be ultimately accepted, it will be convenient to consider them here.

We may now take these four classes one by one.

A. The Simple Proteids

Class I. **Albumins.**—These are soluble in water, in dilute saline solutions, and in saturated solutions of sodium chloride and magnesium sulphate. They are, however, precipitated by saturating their solutions with ammonium sulphate. Their solutions are coagulated by heat, usually at 70–73° C. The following are instances:—

(*a*) Serum albumin. (*b*) Egg albumin. (*c*) Lact-albumin (see Milk).

Class II. **Globulins.**—These are insoluble in water, soluble in dilute saline solutions, and insoluble in concentrated solutions of neutral salts like sodium chloride, magnesium sulphate, and ammonium sulphate. A globulin dissolved in a dilute saline solution may therefore be precipitated—

1. By removing the salt—by dialysis (see p. 31).

2. By increasing the amount of salt. The best salts to employ are ammonium sulphate (half saturation) or magnesium sulphate (complete saturation).

The globulins are coagulated by heat; the temperature of heat coagulation varies considerably. The following are instances:—

(*a*) Fibrinogen
(*b*) Serum globulin (paraglobulin) } in blood plasma.
(*c*) Egg globulin in white of egg.
(*d*) Paramyosinogen in muscle.
(*e*) Crystallin in the crystalline lens.

If we compare together these two important classes of proteids,

we find that they all give the same general tests, that all are coagulated by heat, but that they differ in solubilities. This difference in solubility may be stated in tabular form as follows :—

Reagent	Albumin	Globulin
Water	soluble	insoluble
Dilute saline solution . . .	soluble	soluble
Saturated solution of magnesium sulphate or sodium chloride . .	soluble	insoluble
Half-saturated solution of ammonium sulphate	soluble	insoluble
Saturated solution of ammonium sulphate	insoluble	insoluble

Class III. **Proteoses** } These products of digestion will be studied
Class IV. **Peptones** } in Lessons VII and VIII.

Class V. **Coagulated Proteids**.—There are two principal subdivisions of these :—

(*a*) Proteids in which coagulation has been produced by heat ; they are insoluble in water, saline solutions, weak acids, and weak alkalis ; soluble after prolonged boiling in concentrated mineral acids ; dissolved by gastric and pancreatic juices, they give rise to peptones.

(*b*) Proteids in which coagulation has been produced by ferments :—

i. Fibrin (see Blood). ii. Myosin (see Muscle). iii. Casein (see Milk).

Directions for testing for and separating proteids will be found at the end of this elementary course.

Albuminates are compounds of proteids (albumin or globulin) with mineral substances. Thus, if a solution of copper sulphate is added to a solution of albumin, a precipitate of copper albuminate is obtained. Similarly, by the addition of other salts of the heavy metals, other metallic albuminates are obtainable.

The albuminates which are obtained by the action of dilute acids and alkalis on either albumins or globulins are of considerable physiological interest because they are formed during digestion, and it is to these we shall chiefly confine our attention. The general properties of the *acid-albumin*, or *syntonin*, and the *allkali-abumin* which are thereby respectively formed will be gathered from the practical exercise which stands at the head of this lesson. They are insoluble in pure water, but are soluble in either acid or alkali, and are precipitated by neutralisation unless disturbing influences like the presence of sodium phosphate are present. It may also be added

that, like globulins, they are precipitated by saturation with such neutral salts as sodium chloride and magnesium sulphate. If dissolved in acid or alkali, they are not coagulated by heat.

A variety of alkali albumin (probably a compound containing a large quantity of alkali) may be formed by adding strong potash to undiluted white of egg. The resulting jelly is called Lieberkühn's jelly. A similar jelly is formed by adding strong acetic acid to undiluted egg-white.

The halogens (chlorine, bromine, and iodine) also form albuminates, and may be used for the precipitation of proteids.

B. The Compound or Conjugated Proteids

These complex substances are compounds of albuminous substances with other organic materials, which are as a rule also of complex nature.

They may be divided into the following groups : —

1. **Hæmoglobin** and its allies. These are compounds of proteid with an iron-containing pigment, and will be fully considered under Blood.

2. **Gluco-proteids**. These are compounds of proteids with members of the carbohydrate group. This class includes the mucins, and substances allied to the mucins, called mucoids.

Dr. Pavy has shown that a *small* quantity of a similar carbohydrate can be split off from various other proteids, which we have already classified as simple proteids.

Mucin is a widely distributed substance, occurring in epithelial cells, or shed out by them (mucus, mucous glands, goblet cells) ; and in connective tissue, where it forms the chief constituent of the ground substance or intercellular material.

The mucin obtained from different sources varies in composition and reactions. There are probably several mucins ; they all agree in the following points :—

(*a*) Physical character. Viscid and tenacious.

(*b*) Precipitability from solutions by acetic acid ; they all dissolve in dilute alkalis like lime water.

(*c*) They are all compounds of a proteid with a carbohydrate radicle, which by treatment with dilute mineral acid can be hydrated into a reducing but non-fermentable sugar. It is probable that the carbohydrate radicle may differ in different mucins ; in some it is certainly the case that the so-called sugar is really a nitrogenous derivative of sugar called glucosamine ($C_6H_{11}O_5NH_2$) : that is, glucose in which HO is replaced by NH_2.

The mucoids differ from the mucins either in being non-precipitable from alkaline solutions by acetic acid, or in being readily soluble in excess of acetic acid. One of these, called *ovo-mucoid*, is found in white of egg; and others, called *pseudo-mucin* and *para-mucin*, are sometimes found in the fluid of ovarian cysts and dropsical effusions.

3. **Nucleins and nucleo-proteids.**—These are compounds of proteid with a complex organic acid called nucleic acid, which contains phosphorus.

Nucleo-proteids.—Compounds of proteids with nuclein. They are found in the nuclei and protoplasm of cells. Caseinogen of milk and vitellin of egg-yolk are similar substances. In physical characters they often closely simulate mucin; in fact, the substance called mucin in the bile is in some animals a nucleo-proteid. They may be distinguished from mucin by the fact that they yield on gastric digestion not only peptone but also an insoluble residue of nuclein which is soluble in alkalis, is precipitable by acetic acid from such a solution, and contains a high percentage (10–11) of phosphorus.

Some of the nucleo-proteids also contain iron, and it is probable that the normal supply of iron to the body is contained in the nucleo-proteids, or hæmatogens (Bunge), of plant and animal cells.

The relationship of nucleo-proteids to the coagulation of the blood is described under Blood.

Nucleo-proteids may be prepared from cellular structures like testis, thymus, kidney, &c., by two methods:—

1. *Wooldridge's method.*—The organ is minced and soaked in water for twenty-four hours. Acetic acid added to the aqueous extract precipitates the nucleo-proteid, or, as Wooldridge called it, *tissue fibrinogen.*

2. *Sodium chloride method.*—The minced organ is ground up in a mortar with solid sodium chloride; the resulting viscous mass is poured into excess of distilled water, and the nucleo-proteid rises in strings to the top of the water.

The solvent usually employed for a nucleo-proteid, whichever method it is prepared by, is a 1-per-cent. solution of sodium carbonate.

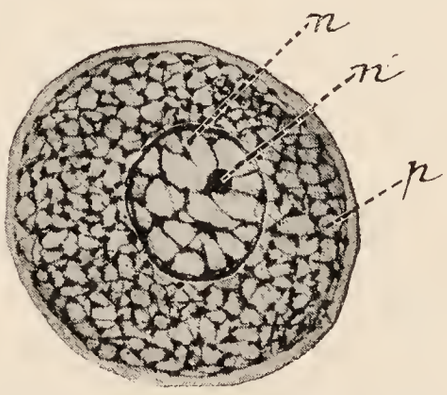


Fig. 9.—Diagram of a cell: *p*, protoplasm composed of spongioplasm and hyaloplasm; *n*, nucleus with intranuclear network of chromatin or nuclein; and *n'*, nucleolus. (Schäfer.)

Nuclein is the chief constituent of cell-nuclei. Its physical characters are something like mucin, but it differs chemically in containing a high percentage of phosphorus. Nuclein is identical with the chromatin of histologists (see fig. 9).

On decomposition nuclein yields a complex organic acid called nucleic acid, together with a variable amount of proteid. Nucleic acid on decomposition yields phosphoric acid and various bases of the xanthine group. Some forms of nuclein, called pseudo-nuclein, such as are obtained from casein and vitellin, differ from the true nucleins in not yielding these xanthine or, as they are sometimes termed, alloxuric or purine bases.

Further particulars concerning the alloxuric bases will be given under Uric Acid, to which they are closely related.

The following diagrammatic way of representing the decomposition of nucleo-proteid will assist the student in remembering the relationships of the substances we have just been considering :—

Nucleo-Proteid

subjected to gastric digestion yields

- Proteid converted into peptone, which goes into solution.
- Nuclein, which remains as an insoluble residue. If this is dissolved in alkali and then hydrochloric acid added it yields
 - Proteid—converted into acid albumin in solution.
 - A precipitate consisting of nucleic acid. If this is heated in a sealed tube with hydrochloric acid, it yields a number of imperfectly known substances like thymic acid and in some cases a reducing sugar. But the best known and constant products of its decomposition are
 - Phosphoric acid.
 - Purine bases, viz. :
 Adenine
 Hypoxanthine
 Guanine
 Xanthine.

The nuclein obtained from the nuclei or heads of the spermatozoa consists of nucleic acid without any proteid admixture. In fishes' spermatozoa, however, the nucleic acid is united to protamine, the chemical properties of which we shall be considering immediately.

C. The Albuminoids

The **albuminoids** form a heterogeneous group of substances which, though similar to the proteids in many particulars, differ from them in certain other points. The principal members of the group are the following :—

1. **Collagen**, the substance of which the white fibres of connective tissue are composed. Some observers regard it as the anhydride of gelatin.

2. **Ossein.**—This is the same substance derived from bone.[1]

3. **Gelatin.**—This substance is produced by boiling collagen with water. It possesses the peculiar property of setting into a jelly when a solution made with hot water cools. It gives most of the proteid colour tests. It contains very little sulphur. On digestion it is like proteid converted into peptone-like substances and is readily absorbed. Though it will replace in diet a certain quantity of proteid, acting as what is called a 'proteid-sparing' food, it cannot altogether take the place of proteid as a food. Animals fed on gelatin instead of proteid waste rapidly. *Chondrin*, a very similar substance obtained from hyaline cartilage, is a mixture of gelatin with mucinoid materials.

4. **Elastin.**—This is the substance of which the yellow or elastic fibres of connective tissue are composed. It is a very insoluble material. The sarcolemma of muscular fibres and certain basement membranes are very similar.

5. **Keratin**, or horny material, is the substance found in the surface layers of the epidermis, in hairs, nails, hoofs, and horns. It is very insoluble, and chiefly differs from proteids in its high percentage of sulphur. A similar substance, called *neurokeratin*, is found in neuroglia and nerve fibres. In this connection it is interesting to note that the epidermis and the nervous system are both formed from the same layer of the embryo—the epiblast.

6. **Chitin** and similar substances found in the exo-skeleton of many invertebrates.

[1] In round numbers the solid matter in *bone* contains two-thirds inorganic or earthy matter, and one-third organic or animal matter. The inorganic constituents are calcium phosphate (84 per cent. of the ash), calcium carbonate (13 per cent.), and smaller quantities of calcium chloride, calcium fluoride, and magnesium phosphate. The organic constituents are ossein (this is the most abundant), elastin from the membranes lining the Haversian canals, lacunæ, and canaliculi, and proteids and nuclein from the bone corpuscles. There is also a small quantity of fat even after removal of all the marrow. *Dentine* is like bone chemically, but the proportion of earthy matter is rather greater. *Enamel* is the hardest tissue in the body; the mineral matter is like that found in bone and dentine; but the organic matter is so small in quantity as to be practically non-existent. (Tomes.) Enamel is epiblastic, not mesoblastic, like bone and dentine.

D. The Protamines

These are basic substances which are combined with nuclein in the heads of the spermatozoa of certain fishes. They resemble proteids in many of their characters—*e.g.* they give Piotrowski's reaction and some of the other tests for proteids. They are regarded by Kossel as the simplest proteids. On hydrolytic decomposition, they first yield substances analogous to the peptones, which are called *protones*, and then they split up into simpler materials which are mainly bases containing six atoms of carbon, and called in consequence the *hexone* bases. These bases have the following names and formulæ :—

Histidine	$C_6H_9N_3O_2$
Arginine	$C_6H_{14}N_4O_2$
Lysine	$C_6H_{14}N_2O_2$

Protamines differ in their composition according to their source, and yield these products in different proportions. Thus *salmine* (the protamine from the salmon) has the empirical formula $C_{30}H_{57}N_{17}O_6$, and *sturine* (from the sturgeon) $C_{36}H_{69}N_{19}O_7$.

A good deal of work still remains to be done at these substances, but the five which have hitherto been most fully worked at are salmine, sturine, scombrine (from the mackerel), cyclopterine (from *Cyclopterus lumpus*), and clupeine (from the herring).

Of these salmine and clupeine appear to be identical; the principal products of the decomposition of salmine, clupeine, and scombrine are arginine, amido-valerianic acid, and a small quantity of an unknown residue.

Sturine yields these same products, with lysine and histidine in addition.

Cyclopterine yields the same products as sturine, with the addition of an aromatic substance.

The more complex proteids and albuminoids yield hexone bases, and so Kossel considers that all these substances contain a protamine nucleus. The more complex proteids, however, yield many other products of decomposition in addition to these bases, of which leucine ($C_6H_{13}NO_2$) and tyrosine ($C_9H_{11}NO_3$), an aromatic substance, may be specially mentioned. Cyclopterine, which also yields an aromatic substance, is thus an important chemical link between the other protamines and the proteids proper. It is interesting to note how many of these decomposition products contain six carbon atoms, and it reminds one that in the sugars obtained from starch there are also six atoms of carbon.

LESSON VI

FOODS

A. **Milk.** 1. Examine a drop of milk with the microscope.

2. Note the specific gravity of fresh milk with the lactometer; compare this with the specific gravity of milk from which the cream has been removed (skimmed milk). The specific gravity of skimmed milk is higher owing to the removal of the lightest constituent—the cream.

3. The reaction of fresh milk is neutral or slightly alkaline.

4. Warm some milk in a test-tube to the temperature of the body, and add a few drops of rennet. After standing, a *curd* is formed from the conversion of *caseinogen*, the chief proteid in milk, into *casein*. The casein entangles the fat globules. The liquid residue is termed *whey*. No curdling is produced if the rennet solution is previously boiled, because heat kills ferments.

5. Take some milk to which 0·2 per cent. of potassium oxalate has been added; warm to 40° C. and add rennet. No curdling takes place because the oxalate has precipitated the calcium salts which are necessary in the coagulation process.

Take a second specimen of oxalated milk and add a few drops of 2-per-cent. solution of calcium chloride, and then rennet; curdling or coagulation takes place if the mixture is kept warm in the usual way.

6. To another portion of warm milk diluted with water add a few drops of 20-per-cent. acetic acid. A lumpy precipitate of caseinogen entangling the fat is formed.

7. Filter off this precipitate, and in the filtrate test for *lactose* or *milk sugar* by Trommer's test (see Lesson II.); for *lact-albumin* by boiling, or by Millon's reagent (see Lesson IV.); and for earthy (that is, calcium and magnesium) *phosphates* by ammonia, which precipitates these phosphates. Phosphates may also be detected by adding nitric acid and ammonium molybdate and boiling; a yellow crystalline precipitate is formed.

8. Fat (*butter*) may be extracted from the precipitate by shaking it with ether; on evaporation of the ethereal extract the fat is left behind, forming a greasy stain on paper. The presence of fat may also be demonstrated by the black colour produced by the addition of osmic acid to the milk.

9. Shake up a little milk with twice its volume of ether; very little fat dissolves and the opacity of the milk is nearly as great as before. Repeat this, but add to the milk a few drops of caustic potash before adding the ether, and then shake. The ether then easily dissolves the fat, and the milk which lies below it becomes translucent.

10. *Caseinogen*, like globulins, is precipitated by saturating milk with sodium chloride or magnesium sulphate, and by half saturation with ammonium sulphate, but differs from the globulins in not being coagulated by heat. The precipitate produced by saturation with salt floats to the

surface with the entangled fat, and the clear salted whey is seen below after an hour or two.

B. **Flour.**—Mix some wheat flour with a little water into a stiff dough. Wrap this up in a piece of muslin and knead it under a tap or in a capsule of water. The starch grains come through the holes in the muslin (identify by iodine test), and an elastic sticky mass remains behind. This is a proteid called *gluten*. Suspend a fragment of gluten in water; add nitric acid and boil; it turns yellow; cool and add ammonia; it turns orange (xanthoproteic reaction). Boil another fragment with Millon's reagent; it turns a brick-red colour.

C. **Bread** contains the same constituents as flour, except that some of the starch has been converted into dextrin and dextrose during baking (most flours, however, contain a small quantity of sugar). Extract breadcrust with cold water, and test the extract for dextrin (iodine test) and for dextrose (Trommer's test). If hot water is used, starch also passes into solution.

D. **Meat.**—This is our main source of proteid food. Cut up some lean meat into fine shreds and grind these up with salt solution. Filter and test for proteids.

THE PRINCIPAL FOOD-STUFFS

We can now proceed to apply the knowledge we have obtained of the proteids, carbohydrates, and fats to the investigation of some important foods. We do not to any great extent use as food the various organic proximate principles in the pure condition; it is necessary that in a suitable diet these should be mixed in certain proportions, and in nature we find them already mixed for us. The chief proximate principles in food are :—

1. Proteids	organic.
2. Carbohydrates	
3. Fats	
4. Water	inorganic.
5. Salts	

In milk and in eggs, which form the exclusive foods of young animals, all varieties of these proximate principles are present in suitable proportions. Hence they are spoken of as perfect foods. Eggs, though a perfect food for the developing bird, contain too little carbohydrate for a mammal. In most vegetable foods carbohydrates are in excess, while in animal food, like meat, the proteids are predominant. In a suitable diet these should be mixed in proper proportions, which must vary for herbivorous and carnivorous animals. We must, however, limit ourselves to the omnivorous animal, man.

A healthy and suitable diet must possess the following characters :—

1. It must contain the proper amount and proportion of the various proximate principles.

2. It must be adapted to the climate, to the age of the individual, and to the amount of work done by him.

3. The food must contain not only the necessary amount of proximate principles, but these must be present in a digestible form. As an instance of this, many vegetables (peas, beans, lentils) contain even more proteid than beef and mutton, but are not so nutritious, as they are less digestible, much passing off in the fæces unused.

The nutritive value of a diet depends mainly on the amount of carbon and nitrogen it contains in a readily digestible form. A man doing a moderate amount of work will eliminate, chiefly from the lungs in the form of carbonic acid, from 250 to 280 grammes of carbon *per diem*. During the same time he will eliminate, chiefly in the form of urea in the urine, about 15 to 18 grammes of nitrogen. These substances are derived from the metabolism of the tissues, and various forms of energy, work and heat being the chief, are simultaneously liberated. During muscular exercise the output of carbon greatly increases; the increased excretion of nitrogen is not nearly so marked. Taking, then, the state of moderate exercise, it is necessary that the waste of the tissues should be replaced by fresh material in the form of food; and the proportion of carbon to nitrogen should be the same as in the excretions: 250 to 15, or 16·6 to 1. The proportion of carbon to nitrogen in proteid is, however, 53 to 15, or 3·5 to 1. The extra supply of carbon must come from non-nitrogenous foods—viz. fat and carbohydrate.

Moleschott gives the following daily diet:—

Proteid	120	grammes.
Fat	90	,,
Carbohydrate	333	,,

Ranke's diet closely resembles Moleschott's; it is—

Proteid	100	grammes.
Fat	100	,,
Carbohydrate	250	,,

In preparing diet tables, such adequate diets as those just given should be borne in mind. The following dietary (from G. N. Stewart) will be seen to be rather more liberal, but may be taken as fairly typical of what is usually consumed by an adult man in the twenty-four hours, doing an ordinary amount of work.

Food-stuff	Quality		Grammes of					
			Nitrogen	Carbon	Proteids	Fats	Carbohydrates	Salts
	Metric system	English weights						
Lean meat	250 grammes	9 oz.	8	33	55	8·5	0	4
Bread .	500 ,,	18 ,,	6	112	40	7·5	245	6·5
Milk .	500 ,,	$\frac{3}{4}$ pint	3	35	20	20	25	3·5
Butter .	30 ,,	1 oz.	0	20	0	27	0	0·5
Fat with meat .	30 ,,	1 ,,	0	22	0	30	0	0
Potatoes .	450 ,,	16 ,,	1·5	47	10	0	95	4·5
Oatmeal .	75 ,,	3 ,,	1·7	30	10	4	48	2
			20·2	299	135	97	413	21

Recent research has, however, shown that, for short periods at any rate, a man will maintain his weight on diets even scantier than those of Ranke and Moleschott.

MILK

Milk is often spoken of as a 'perfect food,' and it is so for infants. For those who are older it is so voluminous that unpleasantly large

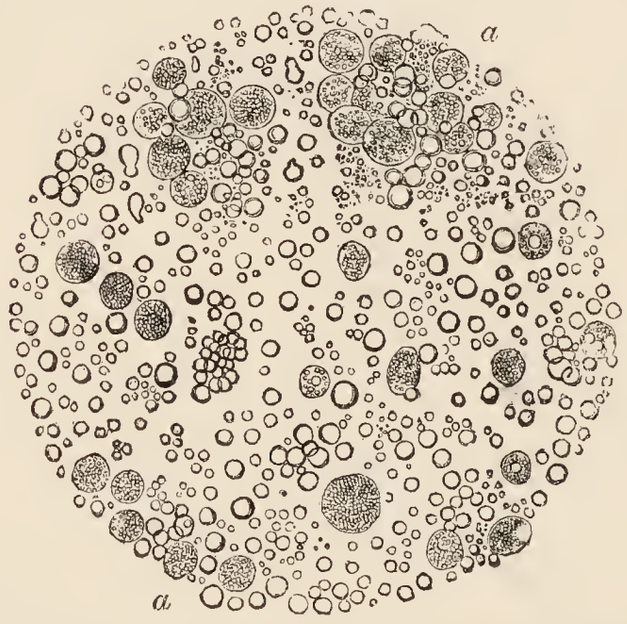


FIG. 10.—Microscopic appearance of milk in the early stage of lactation, showing colostrum corpuscles (*a*) in addition to fat globules. (Yeo.)

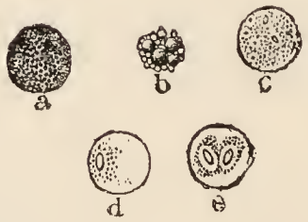


FIG. 11.—*a*, *b*, colostrum corpuscles with fine and coarse fat globules respectively; *c*, *d*, *e*, pale cells devoid of fat. (Heidenhain.)

quantities of it would have to be taken in the course of the day to insure the proper supply of nitrogen and carbon. Moreover, for adults it is relatively too rich in proteid and fat. It also contains too little iron (Bunge); hence children weaned late become anæmic.

The microscope reveals that it consists of two parts : a clear fluid and a number of minute particles that float in it. These consist of minute oil globules, varying in diameter from 0·0015 to 0·005 millimetre.

The milk secreted during the first few days of lactation is called *colostrum*. It contains very little caseinogen, but large quantities of globulin instead. Microscopically, cells from the acini of the mammary gland are seen, which contain fat globules in their interior ; they are called colostrum corpuscles.

Reaction and Specific Gravity.—The reaction of fresh cow's milk and of human milk is amphoteric. This is due to the presence of both acid and alkaline salts ; the latter are usually in excess. Milk readily turns acid or sour as the result of fermentative change, part of its lactose being transformed into lactic acid (see p. 19). The specific gravity of milk is usually ascertained with the hydrometer. That of normal cow's milk varies from 1028 to 1034. When the milk is skimmed the specific gravity rises, owing to the removal of the light constituent, the fat, to 1033 to 1037. In all cases the specific gravity of water, with which other substances are compared, is taken as 1000.

Composition.—Bunge gives the following table, contrasting the milk of woman and cow :—

—	Woman	Cow
	Per cent.	Per cent.
Proteids (chiefly caseinogen) . .	1·7	3·5
Butter (fat)	3·4	3·7
Lactose	6·2	4·9
Salts	0·2	0·7

Hence, in feeding infants on cow's milk, it will be necessary to dilute it, and add sugar to make it approximately equal to natural human milk.

The Proteids of Milk.—The principal proteid in milk is called *caseinogen* ; this is the one which is coagulated by rennin or rennet to form *casein*. Cheese consists of casein with the entangled fat. The other proteid in milk is an albumin. It is present in small quantities only ; it differs in some of its properties (specific rotation, coagulation temperature, and solubilities) from serum-albumin ; it is called *lact-albumin*.

The Coagulation of Milk.—Rennet is the agent usually employed for this purpose ; it is a ferment secreted by the stomach, especially by sucking animals, and is generally obtained from the calf.

The *curd* consists of the casein and entangled fat : the liquid

residue called *whey* contains the sugar, salts, and albumin of the milk. There is also a small quantity of a new proteid called *whey-proteid*, which differs from caseinogen by not being convertible into casein. It is produced by the decomposition of the caseinogen molecule during the process of curdling.

The curd formed in human milk is more finely divided than that in cow's milk : hence it is more digestible. In feeding children and invalids on cow's milk, the lumpy condition of the curd may be obviated by the addition of lime water or barley water to the milk.

Considerable discussion has taken place as to whether the caseinogen of human milk may not be a different proteid from that of cow's milk, especially in relation to the amount and manner of combination of its phosphorus. The differences, however, appear to be explicable on the hypothesis that they are due to variations in the amounts of calcium salts and of citric acid which are present.

Caseinogen itself may be precipitated by acids such as acetic acid, or by saturation with neutral salts like sodium chloride. This, however, is not coagulation, but precipitation. The precipitate may be collected and dissolved in lime water; the addition of rennet then produces coagulation in this solution, provided that a sufficient amount of calcium salts is present.

The addition of rennet produces coagulation in milk, provided that a sufficient amount of calcium salts is present. If the calcium salts are precipitated by the addition of potassium oxalate, rennet causes no formation of casein. The process of curdling in milk is a double one ; the first action due to rennet is to produce a change in caseinogen ; the second action is that of the calcium salt, which precipitates the altered caseinogen as casein. In blood also, calcium salts are necessary for coagulation, but there they act in a different way, namely, in the production of fibrin-ferment (see Coagulation of Blood).

Caseinogen is not a globulin, though it is, like globulins, precipitated by neutral salts. It differs from a globulin in not being coagulated by heat. It is a nucleo-proteid : that is, a compound of a proteid with the proteid-like but phosphorus-rich material called *pseudo-nuclein* (see p. 38).

Caseinogen, as was originally pointed out by Hammarsten, is a proteid with acid properties; it is quite insoluble in water, but it forms soluble salts with such metallic bases as potassium, sodium, and calcium. The caseinogen as it exists in milk is combined with calcium as calcium caseinogenate. When acetic acid is added to milk, we therefore get calcium acetate, and a precipitate of free caseinogen. On 'dissolving' this caseinogen in an alkali like soda or potash, we have the formation of sodium caseinogen-

ate or potassium caseinogenate as the case may be. The precipitate obtained in milk by the addition of alcohol, or by 'salting out,' is not free caseinogen, but calcium caseinogenate. When we add potassium oxalate to milk, we get the reaction represented in the following equation:—Calcium caseinogenate + potassium oxalate = calcium oxalate + potassium caseinogenate. When we add calcium chloride to oxalated milk, the following equation represents what occurs:—Potassium caseinogenate + calcium chloride = calcium caseinogenate + potassium chloride.

Calcium caseinogenate forms an opalescent solution in water, and reacts with the rennin ferment. The caseinogenates of magnesium, barium, and strontium have similar characters. The caseinogenates of potassium, sodium, and ammonium differ from the above by forming a nearly clear solution in water, and they do not react with the rennin ferment. (W. A. Osborne.)

The Fats of Milk.—The chemical composition of the fat of milk (butter) is very like that of adipose tissue. It consists chiefly of palmitin, stearin, and olein. There are, however, smaller quantities of fats derived from fatty acids lower in the series, especially butyrin and caproin. The relation between these varies somewhat, but the proportion is roughly as follows:—Olein, $\frac{3}{7}$; palmitin, $\frac{1}{3}$; stearin, $\frac{1}{6}$; butyrin, caproin, and caprylin, $\frac{1}{14}$. The old statement that each fat globule is surrounded by a membrane of caseinogen is, according to Ramsden's recent work, correct. Milk also contains small quantities of lecithin, a phosphorised fat; of cholesterin, an alcohol which resembles fat in its solubilities (see BILE); and a yellow fatty pigment or lipochrome.

Milk Sugar or Lactose.—This is a saccharose ($C_{12}H_{22}O_{11}$). Its properties have already been described in Lesson II., p. 19.

Souring of Milk.—When milk is allowed to stand, the chief change which it is apt to undergo is a conversion of a part of its lactose into lactic acid. This is due to the action of micro-organisms, and would not occur if the milk were contained in closed sterilised vessels. Equations showing the change produced are given on p. 19. When souring occurs, the acid which is formed precipitates a portion of the caseinogen. This must not be confounded with the formation of casein from caseinogen which is produced by rennet. There are, however, some bacterial growths which produce true coagulation like rennet.

Alcoholic Fermentation in Milk.—When yeast is added to milk, the sugar does not readily undergo the alcoholic fermentation. Other somewhat similar fungoid growths are, however, able to produce the change, as in the preparation of koumiss; the milk sugar is first inverted, that is dextrose and galactose are formed from it (see p. 19), and it is from these sugars that alcohol and carbonic acid originate.

The Salts of Milk.—The chief salt present is calcium phosphate; a small quantity of magnesium phosphate is also present. The other salts are chiefly chlorides of sodium and potassium.

EGGS

In this country the eggs of hens and ducks are those particularly selected as foods. The *shell* is made of calcareous matter, especially calcium carbonate. The *white* is composed of a richly albuminous fluid enclosed in a network of firmer and more fibrous material. The amount of solids is 13·3 per cent.; of this 12·2 is proteid in nature. The proteids are albumin, with smaller quantities of egg-globulin (see p. 34) and ovo-mucoid (p. 37). The remainder is made up of sugar (0·5 per cent.), traces of fats, lecithin and cholesterin, and 0·6 per cent. of inorganic salts. The *yolk* is rich in food materials for the development of the future embryo. In it there are two varieties of yolk-spherules, one kind yellow and opaque (due to admixture with fat and a yellow lipochrome), and the other smaller, transparent and almost colourless: these are proteid in nature, consisting of the nucleo-proteid called *vitellin* (p. 38). Small quantities of sugar, lecithin, cholesterin, and inorganic salts are also present.

The nutritive value of eggs is high, as they are so readily digestible; but the more an egg is cooked the more insoluble do its proteid constituents become.

MEAT

This is composed of the muscular and connective (including adipose) tissues of certain animals. The flesh of some animals is not eaten; in some cases this is a matter of fashion; some flesh, like that of the carnivora, is stated to have an unpleasant taste; and in other cases (*e.g.* the horse) it is more lucrative to use the animal as a beast of burden.

Meat is the most concentrated and most easily assimilable of nitrogenous foods. It is our chief source of nitrogen. Its chief solid constituent is proteid, and the principal proteid is myosin. In addition to the extractives and salts contained in muscle, there is always a certain percentage of fat, even though all visible adipose tissue is dissected off. The fat-cells are placed between the muscular fibres, and the amount of fat so situated varies in different animals. It is particularly abundant in pork; hence the indigestibility of this form of flesh: the fat prevents the gastric juice from obtaining ready access to the muscular fibres.

The following table gives the chief substances in some of the principal meats used as food :—

Constituents	Ox	Calf	Pig	Horse	Fowl	Pike
Water . .	76·7	75·6	72·6	74·3	70·8	79·3
Solids . .	23·3	24·4	27·4	25·7	29·2	20·7
Proteids and gelatin . .	20·0	19·4	19·9	21·6	22·7	18·3
Fat . . .	1·5	2·9	6·2	2·5	4·1	0·7
Carbohydrate .	0·6	0·8	0·6	0·6	1·3	0·9
Salts . . .	1·2	1·3	1·1	1·0	1·1	0·8

The large percentage of water in meat should be particularly noted ; if a man wished to take his daily minimum of 100 grammes of proteid entirely in the form of meat, it would be necessary for him to consume about 500 grammes (*i.e.* a little more than 1 lb.) of meat *per diem.*

FLOUR

The best wheat flour is made from the interior of wheat grains, and contains the greater proportion of the starch of the grain and most of the proteid. Whole flour is made from the whole grain *minus* the husk, and thus contains not only the white interior but also the harder and browner outer portion of the grain. This outer region contains a somewhat larger proportion of the proteids of the grain. Whole flour contains 1 to 2 per cent. more proteid than the best white flour, but it has the disadvantage of being less readily digested. Brown flour contains a certain amount of bran in addition ; it is still less digestible, but is useful as a mild laxative, the insoluble cellulose mechanically irritating the intestinal canal as it passes along.

The best flour contains very little sugar. The presence of sugar indicates that germination has commenced in the grains. In the manufacture of malt from barley this is purposely allowed to go on.

When mixed with water, wheat flour forms a sticky adhesive mass called dough. This is due to the formation of gluten, and the forms of grain poor in gluten cannot be made into dough (oats, rice, &c.). Gluten does not exist in the flour as such, but is formed on the addition of water from the pre-existing soluble proteids (*e.g.* globulins) in the flour.

The following table contrasts the composition of some of the more important vegetable foods :—

Constituents	Wheat	Barley	Oats	Rice	Lentils	Peas	Potatoes
Water . . .	13·6	13·8	12·4	13·1	12·5	14·8	76·0
Proteid . . .	12·4	11·1	10·4	7·9	24·8	23·7	2·0
Fat	1·4	2·2	5·2	0·9	1·9	1·6	0·2
Starch . . .	67·9	64·9	57·8	76·5	54·8	49·3	20·6
Cellulose . . .	2·5	5·3	11·2	0·6	3·6	7·5	0·7
Mineral salts . .	1·8	2·7	3·0	1·0	2·4	3·1	1·0

We see from this table—

1. The great quantity of starch always present.

2. The small quantity of fat; that bread is generally eaten with butter is a popular recognition of this fact.

3. Proteid, except in potatoes, is pretty abundant, and especially so in the pulses (lentils, peas, &c.). The proteid in the pulses is not gluten, but consists of vitellin and globulin-like substances.

In the mineral matters in vegetables, salts of potassium and magnesium are, as a rule, more abundant than those of sodium and calcium.

BREAD

Bread is made by cooking the dough of wheat flour mixed with yeast, salt, and flavouring materials. A ferment in the flour acts at the commencement of the process when the temperature is kept a little over that of the body, and forms dextrin and sugar from the starch, and then the alcoholic fermentation, due to the action of the yeast, begins. The bubbles of carbonic acid, burrowing passages through the bread, make it light and spongy. This enables the digestive juices subsequently to soak into it readily and affect all parts of it. During baking the gas and alcohol are expelled from the bread, the yeast is killed, and a crust forms from the drying of the outer portions of the dough.

White bread contains, in 100 parts, 7 to 10 of proteid, 55 of carbohydrates, 1 of fat, 2 of salts, and the rest water.

COOKING OF FOOD

The cooking of foods is a development of civilisation, and much relating to this subject is a matter of education and taste rather than of physiological necessity. Cooking, however, serves many useful ends :—

1. It destroys all parasites and danger of infection. This relates not only to bacterial growths, but also to larger parasites, such as tapeworms and trichinæ.

2. In the case of vegetable foods it breaks up the starch grains, bursting the cellulose and allowing the digestive juices to come into contact with granulose.

3. In the case of animal foods it converts the insoluble collagen of the universally distributed connective tissues into the soluble gelatin. The loosening of the fibres is assisted by the formation of steam between them. By thus loosening the binding material, the more important elements of the food, such as muscular fibres, are rendered accessible to the gastric and other juices. Meat before it is cooked is generally kept a certain length of time to allow *rigor mortis* to pass off.

Of the two chief methods of cooking, roasting and boiling, the former is the more economical, as by its means the meat is first surrounded with a coat of coagulated proteid on its exterior, which keeps in the juices to a great extent, letting little else escape than the dripping (fat). Whereas in boiling, unless bouillon and bouilli are used, there is considerable waste. Cooking, especially boiling, renders the proteids more insoluble than they are in the raw state, but this is counterbalanced by the other advantages that cooking possesses.

Beef Tea.—In making beef tea and similar extracts of meat, it is necessary that the meat should be placed in cold water, and this is gradually and carefully warmed. In cooking a joint it is usual to put the meat into boiling water at once, so that the outer part is coagulated, and the loss of material minimised.

An extremely important point in this connection is that beef tea and similar meat extracts should not be regarded as foods. They are valuable as pleasant stimulating drinks for invalids, but they contain very little of the nutritive material of the meat, their chief constituents, next to water, being the salts and extractives (creatine, creatinine, lactic acid, &c.) of flesh.

Many invalids restricted to a liquid diet get tired of milk, and imagine that they get sufficient nutriment by taking beef tea instead. It is very important that this erroneous idea should be corrected. One of the greatest difficulties that a physician has to deal with in these cases is the distaste which many adults evince for milk. It is essential that this should be obviated as far as possible by preparing the milk in different ways to avoid monotony. Some can take koumiss; but a less expensive variation may be introduced in the shape of junkets, which, although well known in the West of England, are comparatively unknown in other parts. The preparation of a junket consists of adding to warm milk in a bowl or dish a small quantity of rennet (Clark's essence is very good for this purpose)

and flavouring material according to taste. The mixture is then put aside, and in a short time the milk sets into a jelly (coagulation of casein), which may then be served with or without cream.

Soup contains the extractives of meat, a small proportion of the proteids, and the principal part of the gelatin. The gelatin is usually increased by adding bones and fibrous tissue to the stock. It is the presence of this substance which causes the soup when cold to gelatinise.

ACCESSORIES TO FOOD

Among these must be placed *alcohol*, the value of which within moderate limits is not as a food but as a stimulant; *condiments* (mustard, pepper, ginger, curry powder, &c.), which are stomachic stimulants the abuse of which is followed by dyspeptic troubles; and *tea*, *coffee*, *cocoa*, and similar drinks. These are stimulants chiefly to the nervous system; tea, coffee, maté (Paraguay), guarana (Brazil), cola nut (Central Africa), bush tea (South Africa), and a few other plants used in various countries all owe their chief property to an alkaloid called *theine* or *caffeine* ($C_8H_{10}N_4O_2$); cocoa to the closely related alkaloid, *theobromine* ($C_7H_8N_4O_2$); coca to *cocaine*. These alkaloids are all poisonous, and used in excess, even in the form of infusions of tea and coffee, produce over-excitement, loss of digestive power, and other disorders well known to physicians. Coffee differs from tea in being rich in aromatic matters; tea contains a bitter principle, tannin. To avoid the injurious solution of too much tannin, tea should only be allowed to infuse (draw) for a few minutes. Cocoa is not only a stimulant but a food as well; it contains about 50 per cent. of fat, and 12 per cent. of proteid. In cocoa as manufactured for the market, the amount of fat is reduced to 30 per cent., and the amount of proteid rises proportionately to about 20 per cent. But cocoa as usually taken in the form of an infusion does not contain much of these food substances. The percentage of proteid in solution (mainly albumose) in a cup of cocoa prepared in the ordinary way is about 0·4.

Green vegetables are taken as a palatable adjunct to other foods, rather than for their nutritive properties. Their potassium salts are, however, abundant. Cabbage, turnips, and asparagus contain 80 to 92 water, 1 to 2 proteid, 2 to 4 carbohydrates, and 1 to 1·5 cellulose per cent. The small amount of nutriment in most green foods accounts for the large meals made by, and the vast capacity of the alimentary canal of, herbivorous animals.

LESSON VII

THE DIGESTIVE JUICES

SALIVA

1. To a little saliva in a test-tube add acetic acid. Mucin is precipitated in stringy flakes.

2. Filter some fresh saliva to separate cells and mucus, and apply the xanthoproteic or Millon's test to the filtrate; the presence of proteid is shown.

3. Put some 0·5-per-cent. starch solution into two test-tubes. Add some filtered saliva to one of them, and put both in the water-bath at 40° C. After five minutes remove them and test both fluids with iodine and Trommer's test. The saliva will be found to have converted the starch into dextrin and sugar (maltose).

4. The presence of potassium sulphocyanide (KCNS) in saliva may be shown by the red colour given by a drop of ferric chloride. This colour is discharged by mercuric chloride.

5. The reaction of saliva is alkaline to litmus paper.

GASTRIC DIGESTION

1. Half fill four test-tubes—

A with water. B with 0·2-per-cent. hydrochloric acid. C with 0·2-per-cent. hydrochloric acid. D with solution of white of egg (1 to 10 of water).

2. To A add a few drops of glycerin extract of stomach[1] (this contains pepsin) and a piece of a solid proteid like fibrin.

To B also add pepsin solution and a piece of fibrin.

To C add only a piece of fibrin.

To D add a few drops of pepsin solution and fill up the tube with 0·2-per-cent. hydrochloric acid.

3. Put the tubes into the water-bath at 40° C., and observe them carefully.

In A the fibrin remains unaltered.

In B it becomes swollen, and gradually dissolves.

In C it becomes swollen, but does not dissolve.

4. After half an hour examine the solution in test-tube B.

(*a*) Colour some of the liquid with litmus and neutralise with dilute alkali. Acid-albumin, syntonin, or parapeptone is precipitated.

(*b*) Take another test-tube, and put into it a drop of 1-per-cent. solution of copper sulphate; empty it out so that the merest trace of copper sulphate remains adherent to the wall of the tube; then add the solution from test-tube B and a few drops of strong caustic potash. A pink colour (biuret re-

[1] Benger's liquor pepticus may be used instead of the glycerin extract of stomach.

action) is produced. This should be carefully compared with the violet tint given by unaltered albumin.

(*c*) To a third portion of the fluid in test-tube B add a drop of nitric acid; albumoses or propeptones are precipitated. This precipitate dissolves on heating and reappears on cooling.

5. Repeat these three tests with the digested white of egg in test-tube D.

6. Examine an artificial gastric digestion which has been kept a week. Note the absence of putrefactive odour; in this it contrasts very forcibly with an artificial pancreatic digestion under the same circumstances.

FERMENTS

The word **fermentation** was first applied to the change of sugar into alcohol and carbonic acid by means of yeast. The evolution of carbonic acid causes frothing and bubbling; hence the term 'fermentation.' The agent yeast which produces this is called the ferment. Microscopic investigation shows that yeast is composed of minute rapidly growing unicellular organisms (torulæ) belonging to the fungus group of plants.



FIG. 12.—Cells of the yeast plant in process of budding, between which are some bacteria. (Yeo's 'Physiology.')

The souring of milk, the transformation of urea into ammonium carbonate in decomposing urine, and the formation of vinegar (acetic acid) from alcohol are produced by the growth of very similar organisms. The complex series of changes known as putrefaction, which are accompanied by the formation of malodorous gases and which are produced by the growth of various forms of bacteria, also come into the same category.

That the change or fermentation is produced by these organisms is shown by the fact that it occurs only when the organisms are present, and stops when they are removed or killed by a high temperature or by certain substances (carbolic acid, mercuric chloride, &c.) called antiseptics. The organisms produce fermentative effects by shedding out soluble ferments or enzymes.

The 'germ theory' of disease explains the infectious diseases by considering that the change in the system is of the nature of fermentation, and, like the others we have mentioned, produced by microbes; the transference of the bacteria or their spores from one person to another constitutes infection. The poisons produced by the growing bacteria appear to be either alkaloidal (ptomaines) or proteid in nature. The existence of poisonous proteids is a very remarkable thing, as no chemical differences can be shown to exist between them and those which are not poisonous, but which are useful as foods.

Snake venom is an instance of a very virulent poison of proteid nature.

There is another class of chemical transformations which at first sight differ very considerably from all of these. They, however, resemble these fermentations in the fact that they occur independently of any apparent change in the agents that produce them. The agents that produce them are not living organisms, but chemical

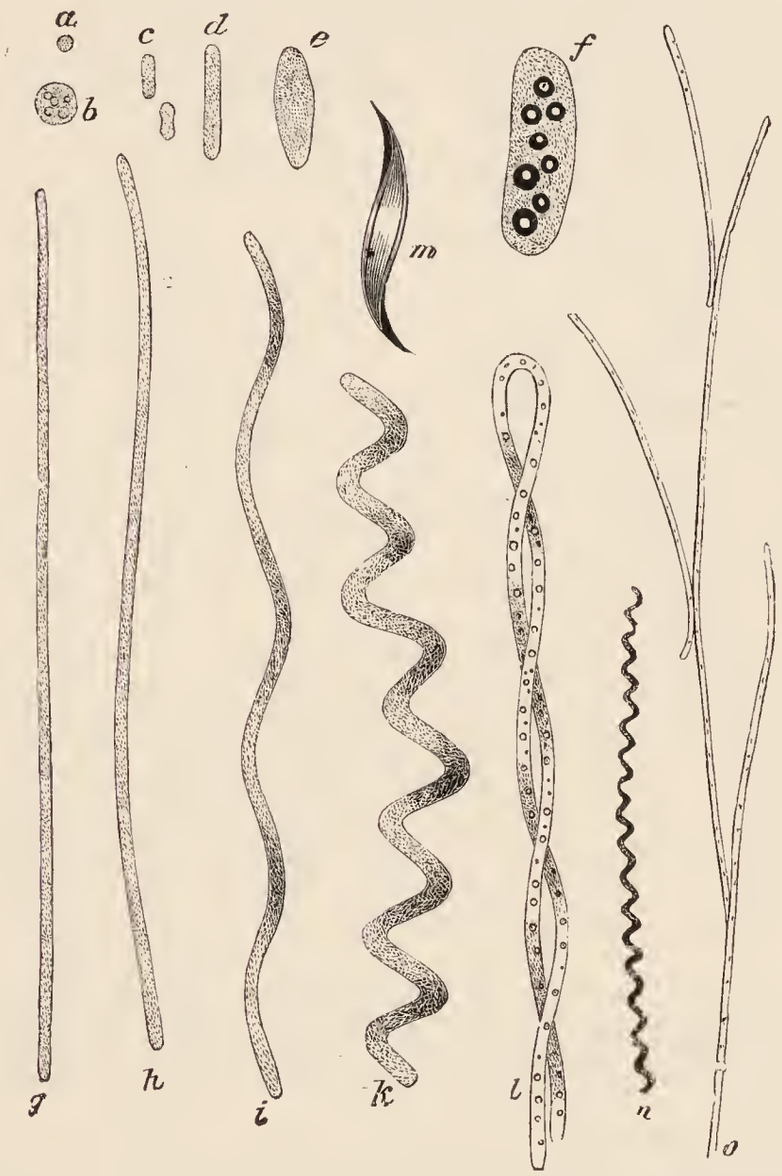


FIG. 13.—Typical forms of Schizomycetes (after Zopf); *a*, micrococcus; *b*, macrococcus; *c*, bacterium; *d*, bacillus; *e*, clostridium; *f*, monas Okenii; *g*, leptothrix; *h*, *i*, vibrio; *k*, spirillum; *l*, spirulina; *m*, spiromonas; *n*, spirochæte; *o*, cladothrix.

substances, the result of the activity of living cells. The change of starch into sugar by the ptyalin of the saliva is an instance.

Ferments may therefore be divided into two classes:—

1. The organised ferments—torulæ, bacteria, &c.
2. The unorganised ferments or enzymes—like ptyalin.

Each may be again subdivided according to the nature of the chemical change produced.

In digestion, the study of which we are just commencing, it is the

unorganised ferments with the action of which we have chiefly to deal. The unorganised ferments may be classified as follows :—

(*a*) Amylolytic—those which change amyloses (starch, glycogen) into sugars. Examples : ptyalin, diastase, amylopsin.

(*b*) Proteolytic—those which change proteids into proteoses and peptones. Examples : pepsin, trypsin.

(*c*) Steatolytic or lipolytic—those which split fats into fatty acids and glycerin. An example, steapsin, is found in pancreatic juice.

(*d*) Inversive—those which convert saccharoses (cane sugar, maltose, lactose) into glucose. Examples : invertin of intestinal juice and of yeast cells.



FIG. 14.—Bacillus anthracis, the agent that produces anthrax or splenic fever (Koch) ; A, bacilli mingled with blood corpuscles from the blood of guinea-pig, some of the bacilli dividing ; B, the same after three hours' culture in a drop of aqueous humour. They grow out into long leptothrix-like filaments, which subsequently divide up, and spores are developed in the segments.

(*e*) Coagulative—those which convert soluble into insoluble proteids. Examples : rennet, fibrin ferment.

Most ferment actions are hydrolytic—*i.e.* water is added to the material acted on, which then splits into new materials. This is seen by the following examples :—

1. Conversion of cellulose into carbonic acid and marsh gas (methane) by putrefactive organisms.

$$\underset{\text{[cellulose]}}{(C_6H_{10}O_5)n} + \underset{\text{[water]}}{nH_2O} = \underset{\text{[carbonic acid]}}{3nCO_2} + \underset{\text{[methane]}}{3nCH_4}$$

2. Inversion of cane sugar by the unorganised ferment invertin :—

$$\underset{\text{[cane sugar]}}{C_{12}H_{22}O_{11}} + \underset{\text{[water]}}{H_2O} = \underset{\text{[dextrose]}}{C_6H_{12}O_6} + \underset{\text{[levulose]}}{C_6H_{12}O_6}$$

It appears also that some enzymes are oxygen carriers and produce oxidation. They are termed **oxidases.** Very little is known of these at present.

A remarkable fact concerning the ferments is that the substances they produce in time put a stop to their activity; thus in the case of the organised ferments the alcohol produced by yeast, the phenol, cresol, &c., produced by putrefactive organisms from proteids, first stop the growth of, and ultimately kill, the organisms which produce them. In the case of the unorganised ferments, the products of their activity hinder and finally stop their action, but on the removal of these products the ferments resume work.

This fact suggested to Croft Hill the question whether ferments will act in the reverse manner to their usual action; and in the case of one ferment, at any rate, he found this to be the case. Inverting ferments, as we have just seen, usually convert a disaccharide into monosaccharides. One of these inverting ferments, called *maltase,* converts maltose into dextrose. If, however, the ferment is allowed to act on strong solutions of dextrose, it converts a small proportion of that sugar back into maltose again. 'Reversible action' has since this been shown to occur in the case of other enzymes.

Ferments act best at a temperature of about 40° C. Their activity is stopped, but the ferments are not destroyed, by cold; it is stopped and the ferments killed by too great heat. A certain amount of moisture and oxygen is also necessary; there are, however, certain micro-organisms that act without free oxygen; these are called **anaërobic,** in contradistinction to those that require oxygen, and which are therefore called **aërobic.**

The chemical nature of the enzymes is very difficult to investigate; they are substances that to a great extent elude the grasp of the chemist. So far research has taught us that they are either proteid in nature or are substances closely allied to the proteids.

The distinction between organised ferments and enzymes is, however, more apparent than real; for the micro-organisms exert their action by enzymes that they secrete. This has long been known in connection with the invertin of yeast and with the enzyme secreted by the micrococcus ureæ which converts urea into ammonium carbonate. In recent years Buchner by crushing yeast cells succeeded in obtaining from them an enzyme which produces the alcoholic fermentation, and there is now no doubt that what is true for yeast is true for all the organised ferments; in several cases this has already been proved experimentally.

THE SALIVA

The **saliva** is the first digestive juice to come in contact with the food; it is secreted by three pairs of salivary glands, the parotid, submaxillary, and sublingual. The secretions from these differ somewhat in composition, but they are mixed in the mouth, the secretion of the minute mucous glands of the mouth and a certain number of epithelial cells and *débris* being added to it. The so-called 'salivary corpuscles' are derived either from the glands themselves or from the tonsils.

The secretion of saliva is a reflex action; the taste or smell of food excites the nerve-endings of the afferent nerves (glossopharyngeal and olfactory); the efferent or secretory nerves are contained in the chorda tympani (a branch of the seventh cranial nerve) which supplies the submaxillary and sublingual, and in a branch of the glossopharyngeal which supplies the parotid. The sympathetic branches which supply the blood-vessels with constrictor nerves contain in some animals secretory fibres also.

The parotid gland is called a serous or albuminous gland; before secretion the cells of the acini are swollen out with granules; after secretion has occurred the cells shrink, owing to the granules having been shed out to contribute to the secretion (see fig. 15).

The submaxillary and sublingual glands are called mucous glands: their secretion contains mucin. Mucin is absent from parotid saliva. The granules in the cells are larger than those of the parotid gland; they are composed of mucinogen, the precursor of mucin (see fig. 16).

In a section of a mucous gland prepared in the ordinary way the mucinogen granules are swollen out, and give a highly refracting appearance to the mucous acini (see fig. 17).

COMPOSITION OF SALIVA

On microscopic examination of mixed saliva a few epithelial scales from the mouth and salivary corpuscles from the salivary glands are seen. The liquid is transparent, slightly opalescent, of slimy consistency, and may contain lumps of nearly pure mucin. On standing it becomes cloudy owing to the precipitation of calcium carbonate, the carbonic acid which held it in solution as bicarbonate escaping.

Of the three forms of saliva which contribute to the mixture found in the mouth, the sublingual is richest in solids (2·75 per cent.). The submaxillary saliva comes next (2·1 to 2·5 per cent.). When

artificially obtained by stimulation of nerves in the dog the saliva obtained by stimulation of the sympathetic is richer in solids than that obtained by stimulation of the chorda tympani. The parotid saliva is poorest in total solids (0·3 to 0·5 per cent.), and contains no mucin. Mixed saliva contains in man an average of about 0·5 per cent. of solids : it is alkaline in reaction, due to the salts in it ; and has a specific gravity of 1002 to 1006.

The solid constituents dissolved in saliva may be classified thus :—

Organic
- *a.* Mucin : this may be precipitated by acetic acid.
- *b.* Ptyalin : an amylolytic ferment.
- *c.* Proteid : of the nature of a globulin.
- *d.* Potassium sulphocyanide.

Inorganic
- *e.* Sodium chloride : the most abundant salt.
- *f.* Other salts : sodium carbonate ; calcium phosphate and carbonate ; magnesium phosphate ; potassium chloride.

THE ACTION OF SALIVA

The action of saliva is twofold, physical and chemical.

The physical use of saliva consists in moistening the mucous membrane of the mouth, assisting the solution of soluble substances in the food, and in virtue of its mucin lubricating the bolus of food to facilitate swallowing.

The chemical action of saliva is due to its active principle, ptyalin. This substance belongs to the class of *unorganised ferments* or *enzymes*, and to that special class of unorganised ferments which are called *amylolytic* (starch-splitting) or *diastatic* (resembling diastase, the similar ferment in germinating barley and other grains).

The starch is first split into dextrin and maltose ; the dextrin is subsequently converted into maltose also ; this occurs more quickly with erythro-dextrin, which gives a red colour with iodine, than with the other variety of dextrin called achroö-dextrin, which gives no colour with iodine. Brown and Morris give the following equation :—

$$\underset{\text{[starch]}}{10(C_6H_{10}O_5)_n} + \underset{\text{[water]}}{4nH_2O}$$
$$= \underset{\text{[maltose]}}{4nC_{12}H_{22}O_{11}} + \underset{\text{[achroö-dextrin]}}{(C_6H_{10}O_5)_n} + \underset{\text{[erythro-dextrin]}}{(C_6H_{10}O_5)_n}$$

Ptyalin acts in a similar way, but more slowly on glycogen ; it has no action on cellulose ; hence it is inoperative on uncooked starch grains, for in these the cellulose layers are intact.

Ptyalin acts best at about the temperature of the body (35–40°), and in a neutral medium; a small amount of alkali makes but

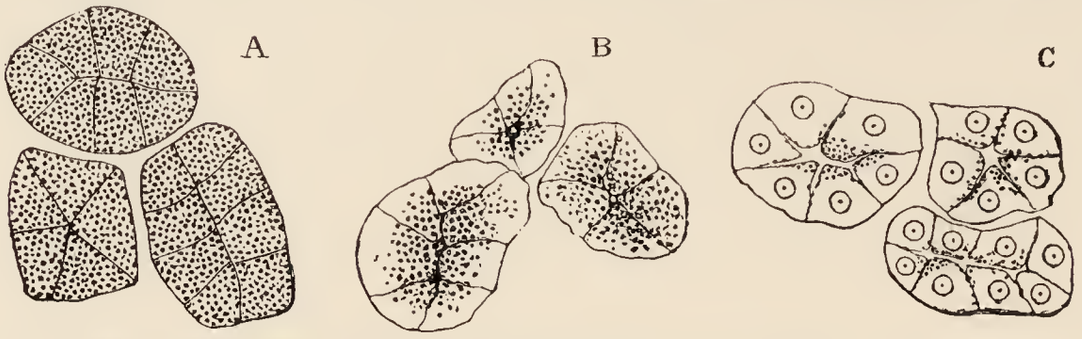


FIG. 15.—Alveoli of serous gland: A, loaded before secretion; B, after a short period of active secretion; C, after a prolonged period. (Langley.)

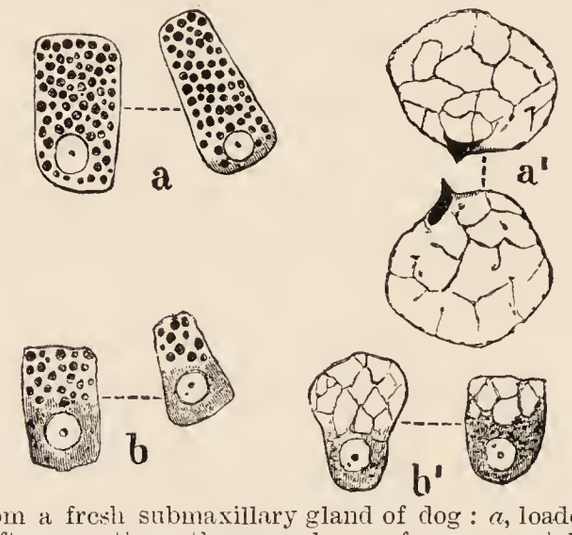


FIG. 16.—Mucous cells from a fresh submaxillary gland of dog: *a*, loaded with mucinogen granules before secretion; *b*, after secretion: the granules are fewer, especially at the attached border of the cell; *a'* and *b'* represent cells in a loaded and discharged condition respectively which have been irrigated with water or dilute acid. The mucous granules are swollen into a transparent mass of mucin traversed by a network of protoplasmic cell-substance. (Foster, after Langley.)

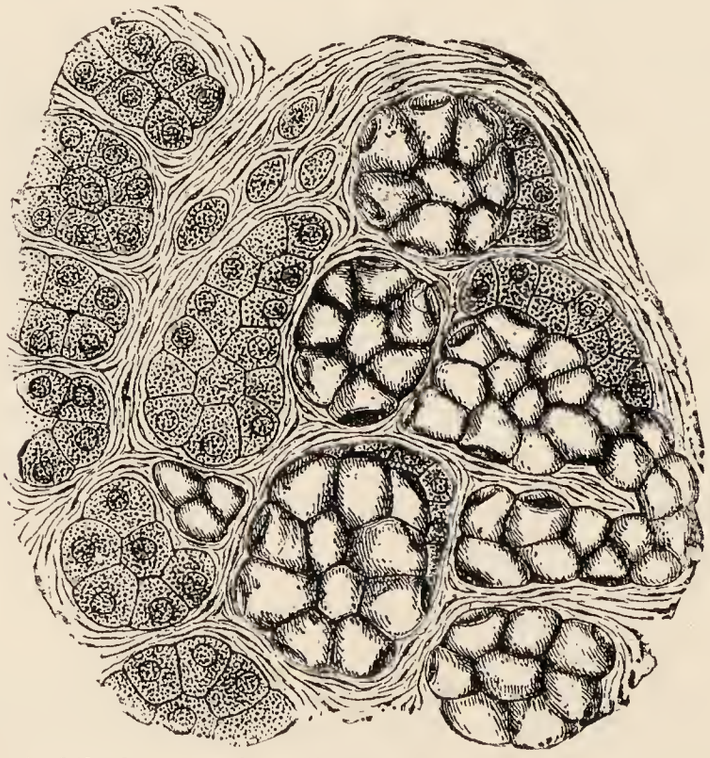


FIG. 17.—Section of part of the human submaxillary gland. (Heidenhain.) To the right is a group of mucous alveoli, to the left a group of serous alveoli.

little difference; a very small amount of acid stops its activity. The conversion of starch into sugar by saliva in the stomach con-

tinues for a considerable time, for the swallowed masses which fall into the fundus of the stomach are not subjected to peristalsis and admixture with gastric juice until a later stage in digestion; the hydrochloric acid which is poured out by the gastric glands first neutralises the saliva and combines with the proteids in the food; but immediately free hydrochloric acid appears the ptyalin is destroyed, so that it does not resume work even when the semi-digested food once more becomes alkaline in the duodenum.

THE SECRETION OF GASTRIC JUICE

The juice secreted by the glands in the mucous membrane of the stomach varies in composition in the different regions, but the mixed gastric juice, as it may be termed, is a solution of a proteolytic ferment called pepsin in a saline solution, which also contains a little free hydrochloric acid.

The gastric juice can be obtained during the life of an animal by means of a gastric fistula. Gastric fistulæ have also been made in human beings, either by accidental injury or by surgical operations. The most celebrated case is that of Alexis St. Martin, a young Canadian who received a musket wound in the abdomen in 1822. Observations made on him by Dr. Beaumont formed the starting-point for our correct knowledge of the physiology of the stomach and its secretion.

We now make artificial gastric juice by mixing weak hydrochloric acid (0·2 to 0·4 per cent.) with a glycerin or aqueous extract of the stomach of a recently killed animal. This artificial juice acts like the normal juice.

Two kinds of glands are distinguished in the stomach, which differ from each other in their position, in the character of their epithelium, and in their secretion. The *cardiac glands* are those situated in the cardiac half of the stomach: their ducts are short, their tubules long in proportion. The latter are filled with polyhedral cells, only a small lumen being left; they are more closely granular than the corresponding cells in the pyloric glands. They are called *principal* or *central* cells. Between them and the basement membrane of the tubule are other cells which stain readily with aniline dyes. They are called *parietal* or *oxyntic* (*i.e.* acid-forming) cells. The *pyloric glands*, in the pyloric half of the stomach, have long ducts and short tubules lined with cubical cells. There are no parietal cells.

The **central cells** of the cardiac glands and the cells of the pyloric

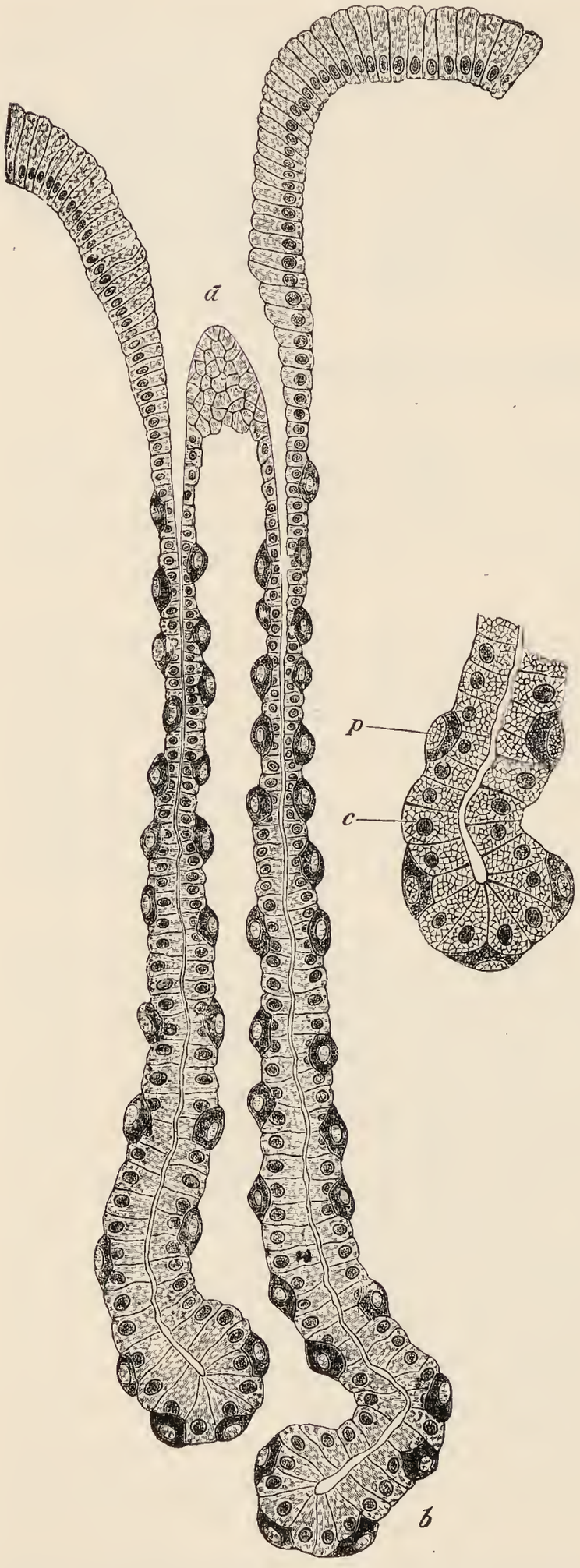


FIG. 18.—A cardiac gland from the dog's stomach (Klein); *d*, duct or mouth of the gland; *b*, base or fundus of one of its tubules; on the right the base of a tubule is more highly magnified; *c*, central cell; *p*, parietal cell.

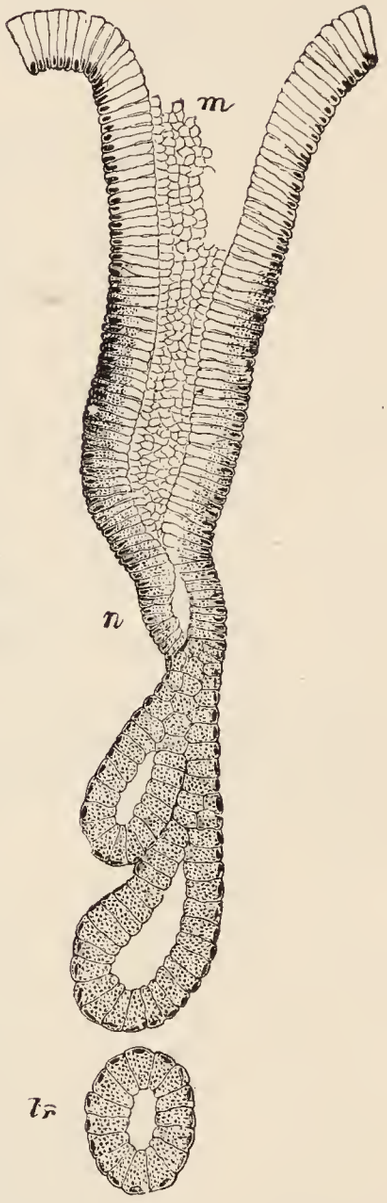


FIG. 19.—A pyloric gland from a section of the dog's stomach (Ebstein): *m*, mouth; *n*, neck: *tr*, a deep portion of tubule cut transversely.

glands are loaded with granules. During secretion they discharge their granules, those that remain being chiefly situated near the lumen, leaving in each cell a clear outer zone (see fig. 20). These are the cells that secrete the pepsin. Like secreting cells generally, they select certain materials from the lymph that bathes them; these materials are worked up by the protoplasmic activity of the cells into the secretion, which is then discharged into the lumen of the gland. The most important substance in a digestive secretion is the ferment. In the case of a gastric juice this is pepsin. We can trace an intermediate step in this process by the presence of the granules. The granules are not, however, composed of pepsin, but of a mother-substance which is readily converted into pepsin. We shall find a similar ferment precursor in the cells of the pancreas, and the term *zymogen* is applied to these ferment percursors. The zymogen in the gastric cells is called *pepsinogen*. The rennet-ferment or rennin that causes the curdling of milk is distinct from pepsin, and is preceded by another zymogen; it is, however, formed by the same cells.

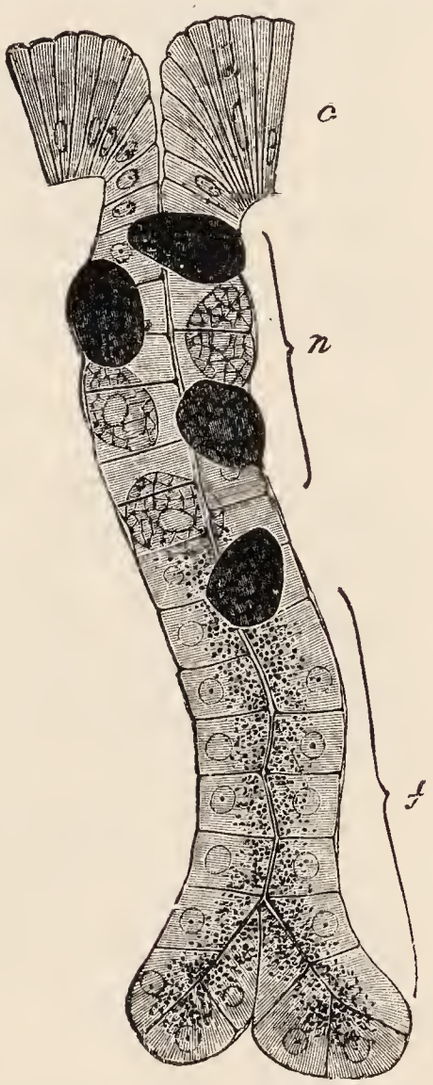


FIG. 20.—A cardiac gland of simple form from the bat's stomach. Osmic acid preparation (Langley): *c*, columnar epithelium of the surface; *n*, neck of the gland, with central and parietal cells; *f*, base or fundus, occupied only by principal or central cells, which exhibit the granules accumulated towards the lumen of the gland.

The **parietal cells** undergo merely a change of size during secretion; at first they are somewhat enlarged, and after secretion they shrink. They are also called oxyntic cells, because it is believed that they secrete the hydrochloric acid of the juice. Heidenhain succeeded in making in one dog a *cul-de-sac* of the fundus, in another of the pyloric region of the stomach; the former secreted a juice containing both acid and pepsin; the latter, parietal cells being absent, secreted a viscid alkaline juice containing pepsin. The formation of a free acid from the alkaline blood and lymph is an important but puzzling problem. There is no doubt that it is formed from the chlorides of the blood and lymph, and of the many theories advanced as to how this is done, Maly's is, on the whole, the most satisfactory. He considers that the acid originates by the interaction of the

calcium chloride with the disodium hydrogen phosphate of the blood, thus :—

$$2Na_2HPO_4 + 3CaCl_2 = Ca_3(PO_4)_2 + 4NaCl + 2HCl$$

[disodium hydrogen phosphate] [calcium chloride] [calcium phosphate] [sodium chloride] [hydrochloric acid]

or more simply by the interaction of sodium chloride and sodium dihydrogen phosphate, as is shown in the following equation :—

$$NaH_2PO_4 + NaCl = Na_2HPO_4 + HCl$$

[sodium dihydrogen phosphate] [sodium chloride] [disodium hydrogen phosphate] [hydrochloric acid]

The sodium dihydrogen phosphate in the above equation is probably derived from the interaction of the disodium hydrogen phosphate and the carbonic acid of the blood, thus :—

$$Na_2HPO_4 + CO_2 + H_2O = NaHCO_3 + NaH_2PO_4.$$

But, as Professor Gamgee has pointed out, these reactions can hardly be considered to occur in the blood generally but rather in the oxyntic cells, which possess the necessary selective powers in reference to the saline constituents of the blood, and the hydrochloric acid, as soon as it is formed, passes into the secretion of the gland in consequence of its high power of diffusion.

COMPOSITION OF GASTRIC JUICE

The following table gives the percentage composition of the gastric juice of man and the dog :—

Constituents	Human	Dog
Water	99·44	97·30
Organic substances (chiefly pepsin)	0·32	1·71
HCl	0·20	0·50
$CaCl_2$	0·006	0·06
NaCl	0·14	0·25
KCl	0·05	0·11
NH_4Cl	—	0·05
$Ca_3(PO_4)_2$	0·01	0·17
$Mg_3(PO_4)_2$		0·02
$FePO_4$		0·008

One sees from this how much richer in all constituents the gastric juice of the dog is than that of man. Carnivorous animals have always a more powerful gastric juice than other animals : they have more work for it to do ; but the great contrast seen in the table is,

no doubt, partly due to the fact that the persons from whom it has been possible to collect gastric juice have been invalids. In the foregoing table one also sees the great preponderance of chlorides over other salts: apportioning the total chlorine to the various metals present, that which remains over must be combined with hydrogen to form the free hydrochloric acid of the juice.

Pepsin stands apart from nearly all other ferments by requiring an acid medium in order that it may act. A compound of the two substances called *pepsin-hydrochloric acid* is the really active agent. Other acids may take the place of hydrochloric acid, but none act so well. Lactic acid is often found in gastric juice: this appears to be derived by fermentative processes from the food.

Pawlow has shown that in dogs the secretory fibres for the gastric glands are contained in the vagus nerves.

By an ingenious surgical operation he succeeded in separating off from the stomach a diverticulum which pours its secretion through an opening in the abdominal wall. This small stomach was found to act in every way like the main stomach of the animal. The pure juice so obtained is clear and colourless; it has a specific gravity of 1003 to 1006. It is feebly dextro-rotatory, gives no biuret reaction, but gives the ordinary proteid reactions. It contains from 0·4 to 0·6 per cent. of hydrochloric acid. It is strongly proteolytic, and inverts cane sugar. When cooled to 0° C. it deposits a fine precipitate of pepsin; this settles in layers, and the layers first deposited contain most of the acid, which is loosely combined with and carried down by the pepsin. Pepsin is also precipitable by saturation with ammonium sulphate (Kühne). Elementary analysis gave the following results:—

Pepsin precipitated by cold—	Precipitated by Am_2SO_4
Carbon . . . 50·73 per cent.	50·37
Hydrogen . . 7·23 ,,	6·88
Chlorine . . 1·01 to 1·17 ,,	0·89
Sulphur . . . 0·98 ,,	1·34
Nitrogen . . . not estimated	14·55 to 15·0
Oxygen . . . the remainder	the remainder.

The juice is most abundant in the early periods of digestion, but it continues to be secreted in declining quantity as long as any food remains to be dealt with. When there is no food given there is no juice. But sham feeding with meat will cause it to flow.

The larger the proportion of proteid in the diet, the more abundant and active is the juice secreted, provided the animal is hungry; the psychical element is of great importance.

THE ACTION OF GASTRIC JUICE

The principal actions of the gastric juice have been already practically studied: the action of pepsin in converting the proteids of the food into the diffusible peptones is its chief action. The curdling of milk by rennet will be found described in Lesson VI.

There is still further action—that is, the gastric juice is antiseptic; putrefactive processes do not normally occur in the stomach, and the organisms that produce such processes, many of which are swallowed with the food, are in great measure destroyed, and thus the body is protected from them. The acid is the agent in the juice that possesses this power.

The formation of peptones is a process of hydrolysis; peptones may be formed by other hydrating agencies like superheated steam and heating with dilute mineral acids. There are certain intermediate steps in this process; the intermediate substances are called propeptones or proteoses. The word 'proteose' is the best to employ: it includes the albumoses (from albumin), globuloses (from globulin), vitelloses (from vitellin), &c. Similar substances are also formed from gelatin (gelatoses) and elastin (elastoses).

Another intermediate step in gastric digestion is called parapeptone: this is acid-albumin or syntonin. In classifying the products of digestion it will be convenient to take albumin as our example, but we must remember that globulin, myosin, and all the other proteids form corresponding products. The products of digestion may be classified according to the order in which they are formed as follows:—

1. Parapeptone or acid-albumin.
2. Propeptone
 - (*a*) Proto-albumose
 - (*b*) Hetero-albumose
 - (*c*) Deutero-albumose

 } The primary albumoses, *i.e.* those which are formed first.
3. Peptone.

It is very doubtful whether all the albumin passes through the acid-albumin stage.

Peptones.—These are the final products of the action of gastric juice on native proteids.

They are soluble in water, are not coagulated by heat, and are not precipitated by nitric acid, copper sulphate, ammonium sulphate, and a number of other precipitants of proteids. They are precipitated but not coagulated by alcohol. They are also precipitated by

tannin, picric acid, potassio-mercuric iodide, phospho-molybdic acid and phospho-tungstic acid.

They give the biuret reaction (rose-red, with a trace of copper sulphate and caustic potash or soda).

Peptone is readily diffusible through animal membranes. The utility of the formation of diffusible substances during digestion is obvious.

Proteoses.—These are the intermediate products in the hydrolysis of native proteids into peptones.

They are not coagulated by heat; they are precipitated but not coagulated by alcohol; like peptone they give the biuret reaction. They are precipitated by nitric acid, the precipitate being soluble on heating, and reappearing when the liquid cools. The last is a distinctive property of proteoses. They are slightly diffusible.

The following table will give us at a glance the chief characters of peptones and proteoses in contrast with those of native proteids like albumin and globulin:—

Variety of proteid	Action of heat	Action of alcohol	Action of nitric acid	Action of ammonium sulphate	Action of copper sulphate and caustic potash	Diffusibility
Albumin	Coagulated	Precipitated, then coagulated	Precipitated in the cold; not readily soluble on heating	Precipitated by complete saturation	Violet colour	*Nil*
Globulin	Ditto	Ditto	Ditto	Precipitated by half saturation; also precipitated by saturation with $MgSO_4$	Ditto	Ditto
Proteoses (albumoses)	Not coagulated	Precipitated, but not coagulated	Precipitated in the cold; readily soluble on heating; the precipitate reappears on cooling [1]	Precipitated by saturation	Rose-red colour (biuret reaction)	Slight
Peptones	Not coagulated	Precipitated, but not coagulated	Not precipitated	Not precipitated	Rose-red colour (biuret reaction)	Great

The difference between the different proteoses is mainly one of solubility. The primary proteoses (proto- and hetero-) are precipitated

[1] In the case of deutero-albumose this reaction only occurs in the presence of excess of salt.

by saturation with magnesium sulphate or sodium chloride. Deutero-proteose is not; it is, however, precipitated by saturation with ammonium sulphate. Proto- and deutero- proteoses are soluble in water: hetero-proteose is not; it requires a salt to hold it in solution.

To sum up: the main action of the gastric juice is upon the proteids of the food, converting them into more soluble and diffusible products. The fats are not chemically altered in the stomach; their proteid envelopes are, however, dissolved, and the solid fats are melted. Starch is unaffected; but cane sugar is inverted. The inversion of cane sugar is due to the hydrochloric acid of the juice, and is frequently assisted by inverting ferments contained in the vegetable food swallowed. The stomach does not digest itself, because it forms an antipepsin; similarly in the intestine an antipepsin is formed. The formation of anti- bodies will be treated under the heading Immunity. (See Blood.)

LESSON VIII

THE DIGESTIVE JUICES (*continued*)

Pancreatic Digestion

1. A 1-per-cent. solution of sodium carbonate, to which a little glycerin extract of pancreas [1] has been added, forms a good artificial pancreatic fluid.

2. Half fill three test-tubes with this solution.

A. To this add half its bulk of diluted egg-white (1 in 10).

B. To this add a piece of fibrin.

C. Boil this; cool; then add fibrin.

3. Put all into the water-bath at 40° C. After half an hour, test A and B for alkali-albumin by neutralisation, for albumoses by nitric acid, and for albumoses and peptone by the biuret reaction.

4. Note in B that the fibrin does not swell up and dissolve, as in gastric digestion, but that it is eaten away from the edges to the interior.

5. In C no digestion occurs, as the ferments have been destroyed by boiling.

6. Take a solution of starch, equal quantities in three test-tubes.

D. To this add a few drops of glycerin extract of pancreas (without the sodium carbonate).

E. To this add a few drops of bile.

F. To this add both bile and pancreatic extract.

7. Put these into the water-bath, and test small portions of each every half-minute by the iodine reaction. It disappears first in F; then in D; while E undergoes no change. Test D and F for maltose by Fehling's solution.

8. Shake up a few drops of olive oil with artificial pancreatic juice (extract of pancreas and sodium carbonate). A milky fluid (emulsion) is formed, from which the oil does not readily separate on standing.

9. The foregoing experiments illustrate the action that pancreatic juice has on all three classes of organic food.

i. *On Proteids.*—Fibrin, albumin, &c. are converted into proteoses and peptone by the ferment *trypsin* in an alkaline medium.

ii. *On Carbohydrates.*—Starch is converted into sugar (maltose) by the ferment *amylopsin*, especially in presence of bile.

iii. *On Fats.*—These are emulsified. In the body they are also split into fatty acid and glycerin by the ferment *steapsin*; but this cannot be shown with the artificial juice, as steapsin is not soluble in glycerin.

Bile

1. Ox bile is given round. Observe its colour, taste, smell, and reaction to litmus paper.

2. Acidulate a little bile with 20-per-cent. acetic acid. A stringy precipitate

[1] Benger's liquor pancreaticus diluted with two or three times its volume of 1-per-cent. sodium carbonate may be used instead.

of a mucinoid substance is obtained. Filter this off and boil the filtrate; no proteid coagulable by heat is present.

3. Add a few drops of bile to (*a*) acid-albumin prepared as described in Lesson V, and (*b*) solution of proteoses to which half its volume of 0·2-per-cent. hydrochloric acid has been added. A precipitate occurs in each case. Bile salts precipitate the unpeptonised proteid which leaves the stomach.

4. *Pettenkofer's Test for Bile Salts.*—To a thin film of bile in a capsule add a drop of solution of cane sugar and a drop of concentrated sulphuric acid. A purple colour is produced. This occurs more quickly on the application of heat. The test may also be performed as follows:—Shake up some bile and cane sugar solution in a test-tube until a froth is formed. Pour concentrated sulphuric acid gently down the side of the tube; it produces a purple colour in the froth.

5. *Gmelin's Test for Bile Pigments.*—On to a little fuming nitric acid (*i.e.* nitric acid containing nitrous acid in solution) in a test-tube pour gently a little bile. Notice the succession of colours—green, blue, red, and yellow—at the junction of the two liquids. This test may also be performed in a capsule. Place a drop of fuming nitric acid in the middle of a thin film of bile; it becomes surrounded by rings of the above-mentioned colours.

6. *Hay's Test for Bile Salts.*—Take two small beakers half full of water; to one add a few drops of bile, or solution of bile salts. Sprinkle a little flowers of sulphur on the surface of each. It remains floating on the pure water, but where bile is present the surface tension of the water is reduced, and the sulphur consequently rapidly sinks. This test is very sensitive.

7. *Preparation of Glycocholic Acid.*—The preparation of the bile acids is usually a task of some difficulty; the following exercise is a simple one, though unfortunately, for reasons which are not explicable, it does not always succeed. Take a stoppered cylinder, place in it 200 c.c. of ox bile, 10 c.c. of hydrochloric acid, and 25 c.c. of ether and shake vigorously. Add a crystal of glycocholic acid, and allow the mixture to stand in a cool place. In a time varying from a few minutes to some hours, a mass of crystals of glycocholic acid separates out. This may be filtered off, washed, and dissolved in a little boiling water, and filtered hot. On cooling, needle-like crystals of the acid again separate out.

8. *Cholesterin.*—(*a*) Examine crystals of cholesterin with the microscope. Heat these on the slide with a drop of sulphuric acid and water (5′·1); the edges of the crystals turn red. (*b*) Salkowski's reaction. Dissolve some cholesterin in chloroform, and gently shake with an equal amount of concentrated sulphuric acid; the solution turns red, and the subjacent acid acquires a green fluorescence.

THE PANCREAS

The Pancreas is a compound tubulo-racemose gland; between the secreting acini are situated little masses of epithelial cells without ducts, called 'islets of Langerhans.' Examination of the secreting cells in different stages of activity reveals changes comparable to those already described in the case of salivary and gastric cells. Granules indicating the presence of a zymogen which is called *trypsinogen* (that is, the precursor of trypsin, the most important ferment of the pancreatic juice) crowd the cells before secretion: these are discharged during secretion, so that in an animal whose

pancreas has been powerfully stimulated to secrete, as by the administration of pilocarpine, the granules are seen only at the free border of the cells (see fig. 21).

As in the case of gastric juice, experiments on the pancreatic secretion are usually performed with an artificial juice, made by mixing a weak alkaline solution (1-per-cent. sodium carbonate) with an extract of pancreas. The pancreas should be kept some time before the extract is made, so as to ensure that the transformation of trypsinogen into trypsin has taken place.

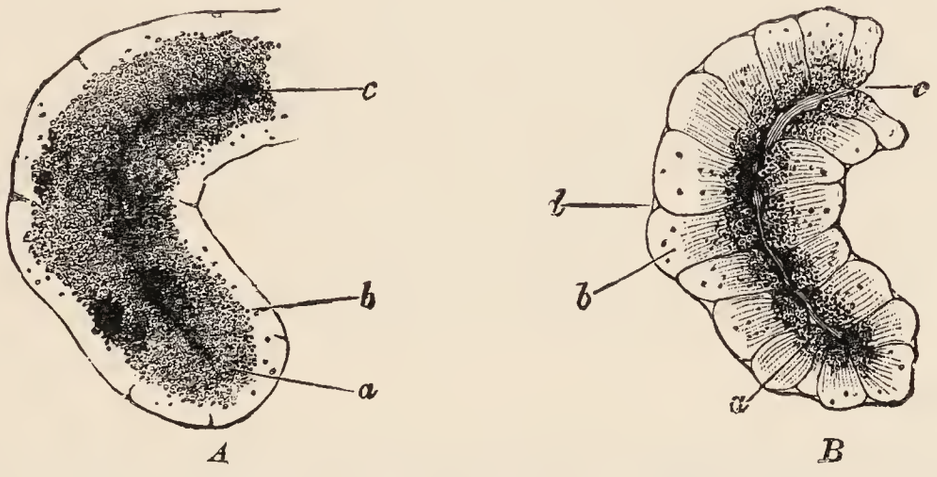


FIG. 21.—Part of an alveolus of the rabbit's pancreas : A, before discharge ; B, after. (From Foster, after Kühne and Lea.)

Quantitative analysis of human pancreatic juice gives the following results :—

Water	97·6 per cent.
Organic solids . . .	1·8 ,,
Inorganic salts . . .	0·6 ,,

Dog's pancreatic juice is considerably richer in solids.

The organic substances in pancreatic juice are—

(*a*) Ferments. These are the most important both quantitatively and functionally. They are four in number :—

i. Trypsin, a proteolytic ferment.

ii. Amylopsin or pancreatic diastase, an amylolytic ferment.

iii. Steapsin, a fat-splitting ferment.

iv. A milk-curdling ferment.

(*b*) A small amount of proteid matter, coagulable by heat.

(*c*) Traces of leucine, tyrosine, xanthine, and soaps.

The inorganic substances in pancreatic juice are—

Sodium chloride, which is the most abundant, and smaller quantities of potassium chloride, and phosphates of sodium, calcium, and

magnesium. The alkalinity of the juice is due to phosphates and carbonates, especially of sodium.

ACTION OF PANCREATIC JUICE

The action of pancreatic juice, which is the most powerful and important of all the digestive juices, may be described under the headings of its four ferments.

1. **Action of Trypsin.**—Trypsin acts like pepsin, but with certain differences, which are as follows:—

(*a*) It acts in an alkaline, pepsin in an acid medium.

(*b*) It acts more rapidly than pepsin; deutero-proteoses can be detected as intermediate products in the formation of peptone. Primary (*i.e.* proto- and hetero-) proteoses have not been found; the action is apparently too rapid to admit of their detection.

(*c*) An albuminate of the nature of alkali-albumin is formed in place of the acid-albumin of gastric digestion.

(*d*) It acts more powerfully on certain albuminoids (such as elastin) which are difficult of digestion in gastric juice. Collagen, however, is not digested.

(*e*) Acting on solid proteids like fibrin, it eats them away from the surface to the interior; there is no preliminary swelling as in gastric digestion.

(*f*) Trypsin acts further than pepsin, on prolonged action partly decomposing the peptone which has left the stomach into simpler products, of which the most important are leucine and tyrosine.

Besides leucine and tyrosine, smaller quantities of aspartic acid [amino-succinic acid, $C_2H_3(NH_2)(COOH)_2$], glutamic acid [amino-pyrotartaric acid $C_3H_5(NH_2)(COOH)_2$], lysine, arginine, ammonia, and a substance called tryptophan are also formed; tryptophan gives a reddish-violet colour with chlorine or bromine water. Tryptophan is skatole-amino-acetic acid (Hopkins).

Kühne considered that the peptones which leave the stomach may be arranged into two classes: (1) hemipeptone, which is split by the prolonged action of pancreatic juice into the substances (leucine, tyrosine, &c.) just enumerated; and (2) antipeptone, which resists this further action.

This theory held the field for many years, but recent researches have shown that it has little or no foundation in fact.

We know that the action of proteolytic enzymes is, by the process of hydrolysis, to split the heavy proteid molecule into smaller and smaller molecules; first we get proteoses, then peptones, and finally after prolonged action simple substances like leucine and tyrosine.

The substance formerly called antipeptone is really not a peptone at all. Siegfried was the first to state that it is a substance of low molecular weight, but further work by Kutscher has shown that it is not a single substance, but a mixture of several of which he has separated out the hexone bases arginine and histidine (see p. 40) and aspartic acid.

These facts can all be easily accounted for on the supposition that a variable fraction of the proteid molecule is broken off with comparative ease, and appears as leucine, tyrosine, and other amino- acids. This is more easily performed by the more powerful tryptic enzyme than by the comparatively feeble agent pepsin. Pepsin, however, is not entirely inactive in this direction, for although leucine and tyrosine cannot as a rule be found in a gastric digestion, yet there are analogous substances of low molecular weight (aspartic acid and the hexone bases) which were incorrectly grouped together by the earlier workers as a peptone (antipeptone). By the prolonged action of pepsin-hydrochloric acid small quantities of leucine and tyrosine are also formed.

2. **Action of Amylopsin.**—The conversion of starch into maltose is the most powerful and rapid of all the actions of the pancreatic juice. It is much more powerful than saliva, and will act even on unboiled starch. The absence of this ferment in the pancreatic juice of infants is an indication that milk, and not starch, is their natural diet.

3. **Action on Fats.**—The action of pancreatic juice on fats is a double one: it forms an *emulsion,* and it decomposes the fats into fatty acids and glycerin by means of its fat-splitting ferment steapsin. The fatty acids unite with the alkaline bases to form soaps (*saponification*). The chemistry of this is described on p. 25. The fat-splitting power of pancreatic juice cannot be studied with a glycerin extract, as steapsin is not soluble in glycerin: either the fresh juice or a watery extract of pancreas must be used.

The formation of an emulsion has been already practically studied in Lesson III. Shake up olive oil and water together, and allow the mixture to stand; the finely divided oil globules soon separate from and float on the surface of the water; but if the oil is shaken up with a solution of soap, the conditions of surface tension are such that the oil globules remain as such in the mixture, and a milky fluid or emulsion is the result. This emulsion is rendered still more permanent in the presence of colloid or viscous materials, especially when a small amount of free fatty acid is being continually liberated. The acid combines with the alkali to form a soap; the soap probably

forms a thin layer on the outside of each oil globule, which prevents them running together again. Pancreatic fluid possesses all the necessary qualifications for forming an emulsion. It is alkaline, and so liberates fatty acid from the fat; these acids form soap with the alkali present; moreover, it is viscous from the presence of proteid.

4. **Milk-curdling Ferment.**—The addition of pancreatic extracts to milk causes clotting, which differs in some of its details from the curdling produced by rennin; but this action can hardly ever be called into play, as the milk upon which the juice has to act has been already curdled by the rennin of the stomach.

THE SECRETION OF PANCREATIC JUICE

Pawlow found the secretory fibres for the pancreas mainly in the vagus, but partly in the sympathetic nerves. The vagus also contains secreto-inhibitory fibres. The secretion is beautifully adjusted to the work it has to do. It is most abundant when most wanted—that is, about three hours after a meal; and its three principal ferments are present in a proportion suitable to the kind of food to be digested.

One of the most effective ways of producing a flow of the juice is to introduce acid into the duodenum, and no doubt the acid of the gastric juice is the normal stimulus for the pancreatic flow. This flow still occurs when all the nerves supplying the duodenum and pancreas are cut, and it was held by Popielski and by Wertheimer and Le Page that it must be due to a local reflex, the centres being situated in the scattered ganglia of the pancreas, and of the solar plexus. Starling and Bayliss, however, pointed out that it cannot be a nervous reflex, since it occurs after extirpation of the solar plexus, and destruction of all nerves passing to an isolated loop of jejunum. Moreover, atropine does not paralyse the secretory action. It must therefore be due to direct excitation of the pancreatic cells by a substance or substances conveyed to the gland from the bowel by the blood-stream.

The exciting substance is not acid; injection of 0·4 per cent. of hydrochloric acid into the blood-stream has no influence on the pancreas. The substance in question must be produced in the intestinal mucous membrane under the influence of the acid. This conclusion was confirmed by experiment. If the mucous membrane of the jejunum or duodenum is exposed to the action of 0·4 per cent. hydrochloric acid, a body is produced which, when injected into the blood-stream in minimal doses, produces a copious secretion of pancreatic juice. This substance is termed *secretin*. It is associated with

another substance which lowers arterial blood-pressure. The two substances are not identical, since acid extracts of the lower end of the ileum produce a lowering of blood-pressure, but have no excitatory influence on the pancreas.

Secretin is split off from a precursor, *pro-secretin*, which is present in relatively large amounts in the duodenal mucous membrane, and gradually diminishes as we descend the intestine. Pro-secretin can be dissolved out of the mucous membrane by normal saline solution. It has no influence on the pancreatic secretion. Secretin can be split off from it by boiling or by treatment with acid.

What secretin is chemically we do not yet know. It is soluble in alcohol and ether. It is not a proteid, but probably is an organic substance of low molecular weight. It is, moreover, the same substance in all animals, and not specific to different kinds of animals.

Whether this discovery is an isolated instance of chemical sympathy between different organs, or whether it is only one of a class of similar mechanisms, must be left to the future.

INTESTINAL DIGESTION

The pancreatic juice does not act alone on the food in the intestines. There are, in addition, the bile, the succus entericus (secreted by the crypts of Lieberkühn), and bacterial action to be considered.

The **succus entericus** or intestinal juice has no action on fats or native proteids, but it appears to have to some extent the power of converting starch into sugar; its best known action is due to a ferment it contains called *invertin*, which inverts saccharoses—that is, converts cane sugar and maltose into glucose. The original use of the term 'inversion' has been explained on p. 17. It may be extended to include the similar hydrolysis of other saccharoses, although there may be no formation of levorotatory substances. There are probably several inverting ferments in the succus entericus, one of which acts on cane sugar, one on maltose, and one on milk sugar.

A few years ago, however, Pawlow showed that succus entericus has a still more important action, which is to intensify the proteolytic power of the pancreatic juice. Fresh pancreatic juice has practically no power on proteids, for what it contains is not trypsin, but its precursor, trypsinogen.

If fresh pancreatic and intestinal juices are mixed together, the result is a very powerful proteolytic mixture, though neither juice by itself has any proteolytic activity. The substance in the intestinal

juice that liberates trypsin from trypsinogen has been called by Pawlow a 'ferment of the ferments' or *entero-kinase*.

Intestinal juice contains another ferment called *erepsin* (Otto Cohnheim) which is capable of breaking up proteoses and peptone into simple substances (leucine, tyrosine, hexone bases, ammonia, &c.). Whether this ferment is of real importance is a question upon which there are differences of opinion at present.

Bacterial Action.—The gastric juice is an antiseptic; the pancreatic juice is not. An alkaline fluid like pancreatic juice is just the most suitable medium for bacteria to flourish in. Even in an artificial digestion the fluid is very soon putrid, unless special precautions to exclude or kill bacteria are taken. It is often difficult to say where pancreatic action ends and bacterial action begins, as many of the bacteria that grow in the intestinal contents, having reached that situation in spite of the gastric juice, act in the same way as the pancreatic juice. Some form sugar from starch, others peptone, leucine, and tyrosine from proteids, while others again break up fats. There are, however, certain actions that are entirely due to these putrefactive organisms.

i. On carbohydrates. The most frequent fermentation they set up is the lactic acid fermentation: this may go further and result in the formation of carbonic acid, hydrogen, and butyric acid (see p. 19). Cellulose is broken up into carbonic acid and methane. This is the chief cause of the gases in the intestine, the amount of which is increased by vegetable food.

ii. On fats. In addition to acting like steapsin, lower acids (valeric, butyric, &c.) are produced. The formation of acid products from fats and carbohydrates gives to the intestinal contents an acid reaction. Recent researches show that the contents become acid much higher up in the small intestine than was formerly considered to be the case. These organic acids do not hinder pancreatic digestion to any appreciable extent.

iii. On proteids. Fatty acids and amino- acids, especially leucine and tyrosine, are produced; but these putrefactive organisms have a special action in addition, producing substances having an evil odour like indole, skatole, and phenol. There are also gaseous products in some cases.

If excessive, putrefactive processes are harmful; if within normal limits, they are useful, helping the pancreatic juice and, further, preventing the entrance into the body of poisonous products. It is possible that, in digestion, poisonous alkaloids are formed. Certainly this is so in one well-known case. Lecithin, a material contained in

small quantities in many foods, and in large quantities in egg-yolk and brain, is broken up by the pancreatic juice into glycerin, phosphoric acid, stearic acid, and an alkaloid called choline. We are, however, protected from the poisonous action of choline by the bacteria, which break it up into carbonic acid, methane, and ammonia.

LEUCINE AND TYROSINE

These two substances have been frequently mentioned in the preceding pages. As types of the decomposition products of proteids they are important, though probably only small quantities are normally formed during digestion.

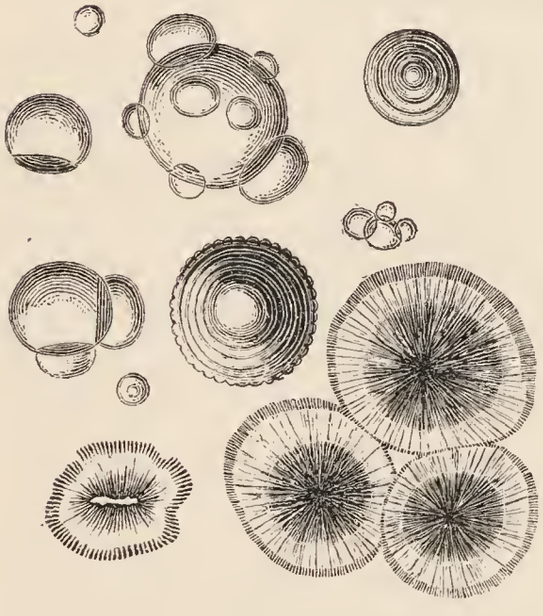


FIG. 22.—Leucine crystals.

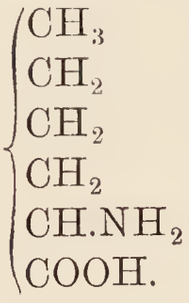


FIG. 23.—Tyrosine crystals.

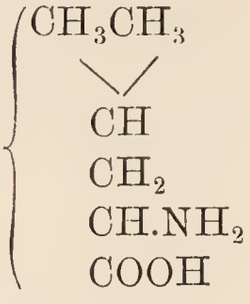
They belong to the group of amino- acids. On p. 24 we have given a list of the fatty acids; if we replace one of the hydrogen atoms in a fatty acid by the amino- group (NH_2), we obtain what is called an amino- acid. Take acetic acid: its formula is $CH_3.COOH$; replace one H by NH_2, and we get $CH_2.NH_2.COOH$, which is *amino-acetic acid* or *glycine*. If we take caproic acid—a term a little higher in the series—its formula is $C_5H_{11}.COOH$; amino-caproic acid is $C_5H_{10}.NH_2.COOH$, which is also called *leucine*. There are, however, several isomeric amino-caproic acids. It was thought until quite recently that leucine was the amino- acid of normal caproic acid, but it has been shown to be *a*-amino-iso-butyl-

acetic acid. The difference in the structure of these two compounds may be represented by the following graphic formulæ :

Normal α-amino-caproic acid.



Iso-butyl α-amino-acetic acid



Tyrosine is a little more complicated, as it is not only an amino- acid, but also contains an aromatic radicle. Propionic acid has the formula $C_2H_5.COOH$; amino-propionic acid is $C_2H_4.NH_2.COOH$, and is called *alanine*. If another H in this is replaced by oxyphenyl ($C_6H_4.OH$), we get $C_2H_3.NH_2.C_6H_4OH.COOH$, which is oxyphenyl-amino-propionic acid, or tyrosine. Figs. 22 and 23 represent the crystalline forms of leucine and tyrosine.

EXTIRPATION OF THE PANCREAS

Complete removal of the pancreas in animals and diseases of the pancreas in man produce a condition of diabetes, in addition to the loss of pancreatic action in the intestines. Grafting the pancreas from another animal into the abdomen of the animal from which the pancreas has been removed relieves the diabetic condition.

How the pancreas acts otherwise than in producing the pancreatic juice is not known. It must, however, have other functions related to the general metabolic phenomena of the body, which are disturbed by removal or disease of the gland. This is an illustration of a universal truth—viz. that each part of the body does not merely do its own special work, but is concerned in the great cycle of changes which is called general metabolism. Interference with any organ upsets not only its specific function, but causes disturbances through the body generally. The interdependence of the circulatory and respiratory systems is a well-known instance. Removal of the thyroid gland upsets the whole body, producing widespread changes known as myxœdema. Removal of the testis produces not only a loss of the spermatic secretion, but changes the whole growth and appearance of the animal. Removal of the greater part of the kidneys produces rapid wasting, and the breaking down of the tissues to form an increased quantity of urea. The precise way in which these glands are related to the general body processes is, however, a subject of which we know as yet very little. The theory at present most in favour is that certain glands produce an internal secretion,

which leaves the gland *viâ* the lymph, and is then distributed to minister to parts elsewhere. Removal of such glands as the thyroid or suprarenal produces disease or death because this internal secretion can no longer be formed. In the case of the pancreas, the external secretion of the pancreas (that is, pancreatic juice) is formed by the cells lining the acini, and the internal secretion, stoppage of which in some way leads to diabetes, is produced in the islets of Langerhans.

In diabetes the oxidative powers of the body cells are lessened, and the capability of these cells to prepare sugar for oxidation is impaired. In this process the sugar or its derivative glycuronic acid is split into smaller molecules, and ultimately into water and carbon dioxide. The close relationship of sugar and glycuronic acid is shown by the following formulæ:—

COH	COH
$(CHOH)_4$	$(CHOH)_4$
CH_2OH	$COOH$
[dextrose]	[glycuronic acid]

That is, two hydrogen atoms in the CH_2OH group are replaced by one of oxygen. This oxidation is readily brought about in the body, and glycuronic acid is usually found in diabetic urine; but the further oxidation into water and carbon dioxide is a more difficult task, because it involves the disruption of the linkage of the carbon atoms. Perhaps it is here that the internal secretion of the pancreas is effective. This, however, is at present a mere theory, and certainly Lépine's idea that the ferment of the pancreatic internal secretion is one which initiates glycolysis or sugar-splitting *in the blood*, has been abundantly disproved. It may be that the active principle of the pancreatic internal secretion stimulates the glycolytic action of the tissue-cells. It is conceivable that in the other great cause of experimental glycosuria, namely injury to nervous structures, as in Bernard's puncture experiment, the derangement of the nervous system exerts some unknown influence on the pancreas as well as on the liver.

Many poisons produce temporary glycosuria, but the most interesting and powerful of these is phloridzin. The diabetes produced is very intense. Phloridzin is a glucoside, but the sugar passed in the urine is too great to be accounted for by the small amount of sugar derivable from the drug. Besides that, phloretin, a derivative of phloridzin, free from sugar, produces the same results.

Phloridzin produces diabetes in starved animals, or in those in which any carbohydrate store must have been got rid of by the previous administration of the same drug. Phloridzin-diabetes is therefore analogous to those intense forms of diabetes in man in which the sugar must be derived from protoplasmic metabolism.

A puzzling feature is the absence of an increase of sugar in the blood; if the phloridzin is directly injected into one renal artery, sugar rapidly appears in the secretion of that kidney; the sugar is formed within the kidney cells from some substance in the blood, but whether that substance is proteid or not is uncertain. The action of the kidney cells in forming sugar has been compared to that of the mammary cells in forming lactose.

Death in diabetic patients is usually preceded by deep coma, or unconsciousness. Some poison must be produced that acts soporifically upon the brain. The breath and urine of these patients smell strongly of acetone; hence the term *acetonæmia*. This apple-like smell should always suggest

the possible onset of coma and death, but it is exceedingly doubtful whether acetone (which can certainly be detected in the urine) is the true poison; ethyl-diacetic acid, which accompanies and is the source of the acetone, was regarded by some as the actual poison, but these substances, when introduced into the circulation artificially, do not cause serious symptoms. The principal poison is amino-hydroxybutyric acid. The alkalinity and carbonic acid of the blood are decreased, and the ammonia of the urine is increased; this indicates an attempt of the body to neutralise the poisonous acids.

THE BILE

Bile is the secretion of the liver which is poured into the duodenum; it has been collected in living animals by means of a biliary fistula; the same operation has occasionally been performed in human beings. At death the gall bladder yields a good supply of bile which is more concentrated than that obtained from a fistula.

Bile is being continuously poured into the intestine, but there is an increased discharge immediately on the arrival of food in the duodenum; there is a second increase in secretion a few hours later.

Though the chief blood supply of the liver is by a vein (the portal vein), the amount of blood in the liver varies with its needs, being increased during the periods of digestion. This is due to the fact that in the area from which the portal vein collects blood—stomach, intestine, spleen, and pancreas—the arterioles are all dilated, and the capillaries are thus gorged with blood. Further, the active peristalsis of the intestine and the pumping action of the spleen are additional factors in driving more blood onwards to the liver.



Fig. 24.—Hæmatoidin crystals.

The bile is secreted from the portal blood at a much lower pressure than one finds in glands, such as the salivary glands, the blood supply of which is arterial. Heidenhain found that the pressure in the bile duct of a dog averaged 15 mm. of mercury, which is about double that in the portal vein.

The second increase in the flow of bile—that which occurs some hours after the arrival of the semi-digested food (chyme) in the intestine—appears to be due to the effect of the digestive products carried by the blood to the liver stimulating the hepatic cells to activity; this is supported by the fact that proteid food increases the quantity of bile secreted, whereas fatty food which is absorbed, not by the portal vein, but by the lacteals, has no such effect.

The chemical processes by which the constituents of the bile are formed are obscure. We, however, know that the biliary pigment is produced by the decomposition of hæmoglobin. Bilirubin is, in fact, identical with the iron-free derivative of hæmoglobin called hæmatoidin, which is found in the form of crystals in old blood-clots such as occur in the brain after cerebral hæmorrhage (see fig. 24).

An injection of hæmoglobin into the portal vein, or of substances like water which liberate hæmoglobin from the red blood corpuscles, produces an increase of bile pigment. If the spleen takes any part in the elaboration of bile pigment, it does not proceed so far as to liberate hæmoglobin from the corpuscles. No free hæmoglobin is discoverable in the blood plasma in the splenic vein.

The amount of bile secreted is differently estimated by different observers; the amount secreted daily in man appears to vary from 500 c.c. to 1 litre (1,000 c.c.).

THE CONSTITUENTS OF BILE

The constituents of the bile are the bile salts proper (taurocholate and glycocholate of soda), the bile pigments (bilirubin, biliverdin), a mucinoid substance, small quantities of fats, soaps, cholesterin, lecithin, urea, and mineral salts, of which sodium chloride and the phosphates of iron, calcium, and magnesium are the most important.

Bile is a yellowish, reddish-brown, or green fluid, according to the relative preponderance of its two chief pigments. It has a musk-like odour, a bitter-sweet taste, and a neutral or faintly alkaline reaction.

The specific gravity of human bile from the gall bladder is 1026 to 1032; that from a fistula, 1010 to 1011. The greater concentration of gall-bladder bile is partly but not wholly explained by the addition to it from the walls of that cavity of the mucinoid material.

The amount of solids in bladder bile varies from 9 to 14 per cent., in fistula bile from 1·5 to 3 per cent. The following table shows that this low percentage of solids is almost entirely due to want of bile salts. This can be accounted for in the way first suggested by Schiff—that there is normally a bile circulation going on in the body; a large quantity of the bile salts that passes into the intestine is first split up, then reabsorbed and again secreted. Such a circulation would obviously be impossible in cases where all the bile is discharged to the exterior.

The following table gives some important analyses of human bile:—

Constituents	Fistula bile (healthy woman. Copeman and Winston)	Fistula bile (case of cancer. Yeo and Herroun)	Normal bile (Frerichs)
Sodium glycocholate .	0·6280	0·165	9·14
Sodium taurocholate .		0·055	
Cholesterin, lecithin, fat .	0·0990	0·038	1·18
Mucinoid material . .	0·1725	0·148	2·98
Pigment	0·0725		
Inorganic salts . .	0·4510	0·878	0·78
Total solids . . .	1·4230	1·284	14·08
Water (by difference) .	98·5570	98·716	85·92
	100·0000	100·000	100·00

Bile Mucin.—There has been considerable diversity of opinion as to whether bile mucin is really mucin. The most recent work in Hammarsten's laboratory shows that differences occur in different animals. Thus in the ox there is very little true mucin, but a great amount of nucleo-proteid; in human bile, on the other hand, there is very little if any nucleo-proteid: the mucinoid material present there is really mucin. (On the general characters of Mucin and Nucleo-Proteids see pp. 36 to 38.)

The Bile Salts.—The bile contains the sodium salts of complex amino- acids called the bile acids. The two acids most frequently found are glycocholic and taurocholic acids. The former is the more abundant in the bile of man and herbivora; the latter in carnivorous animals, like the dog. The most important difference between the two acids is that taurocholic acid contains sulphur, and glycocholic acid does not.

Glycocholic acid ($C_{26}H_{43}NO_6$) is by the action of dilute acids and alkalis, and also in the intestine, hydrolysed and split into glycine or amino-acetic acid and cholalic acid.

$$\underset{\text{[glycocholic acid]}}{C_{26}H_{43}NO_6} + H_2O = \underset{\text{[glycocine]}}{CH_2.NH_2.COOH} + \underset{\text{[cholalic acid]}}{C_{24}H_{40}O_5}$$

The glycocholate of soda has the formula $C_{26}H_{42}NaNO_6$.

Taurocholic acid ($C_{26}H_{45}NO_7S$) similarly splits into taurine or amino-ethyl-sulphonic acid and cholalic acid.

$$\underset{\text{[taurocholic acid]}}{C_{26}H_{45}NO_7S} + H_2O = \underset{\text{[taurine]}}{C_2H_4.NH_2.HSO_3} + \underset{\text{[cholalic acid]}}{C_{24}H_{40}O_5}$$

The taurocholate of soda has the formula $C_{26}H_{44}NaNO_7S$.

The colour reaction called **Pettenkofer's reaction**, described in the practical exercises at the head of this lesson, is due to the presence

of cholalic acid. The sulphuric acid acting on sugar forms a small quantity of a substance called furfuraldehyde, in addition to other products. The furfuraldehyde gives the purple colour with cholalic acid.

The Bile Pigments.—The two chief bile pigments are bilirubin and biliverdin. Bile which contains chiefly the former (such as dog's bile) is of a golden or orange-yellow colour, while the bile of many herbivora, which contains chiefly biliverdin, is either green or bluish green. Human bile is generally described as containing chiefly bilirubin, but there have been some cases described in which biliverdin was in excess. The bile pigments show no absorption bands with the spectroscope; their origin from the blood pigment has already been stated (p. 81).

Bilirubin has the formula $C_{16}H_{18}N_2O_3$: it is thus an iron-free derivative of hæmoglobin. The iron is apparently stored up in the liver cells, perhaps for future use in the manufacture of new hæmoglobin. The bile contains only a trace of iron.

Biliverdin has the formula $C_{16}H_{18}N_2O_4$ (*i.e.* one atom of oxygen more than in bilirubin): it may occur as such in bile; it may be formed by simply exposing red bile to the oxidising action of the atmosphere; or it may be formed as in Gmelin's test by the more vigorous oxidation produced by fuming nitric acid.

Gmelin's test consists of a play of colours—green, blue, red, and finally yellow—produced by the oxidising action of fuming nitric acid (that is, nitric acid containing nitrous acid in solution). The end or yellow product is called *cholctelin*, $C_{16}H_{18}N_2O_6$.

Hydrobilirubin.—If a solution of bilirubin or biliverdin in dilute alkali is treated with sodium amalgam or allowed to putrefy, a brownish pigment is formed called hydrobilirubin, $C_{32}H_{44}N_4O_7$. With the spectroscope it shows a dark absorption band between *b* and F, and a fainter band in the region of the D line.

Urobilin.—Hydrobilirubin is interesting because a similar substance is formed from the bile pigment by reduction processes in the intestine, and constitutes *stercobilin*, the pigment of the fæces. Some of this is absorbed and ultimately leaves the body in the urine as one of its pigments called *urobilin*. A small quantity of urobilin is sometimes found preformed in the bile. The identity of urobilin and stercobilin has been frequently disputed, but the recent work of Garrod and Hopkins has confirmed the old statement that they are the same substance with different names. Urobilin has a well-marked absorption band in the region of the F line, and when partially precipitated from an alkaline solution by acidification, it also shows an absorption band

in the region of the E line. Hydrobilirubin differs from urobilin in containing much more nitrogen in its molecule (9·2 instead of 4·1 per cent.), and is probably a product of less complete reduction than urobilin. (See further Lesson XXVI.)

Cholesterin.—This substance is contained not only in bile, but very largely in nervous tissues. Like lecithin, it is an abundant constituent of the white substance of Schwann. It is found also in blood corpuscles. In bile it is normally present in small quantities only, but it may occur in excess, and so form the concretions known as gallstones, which are generally more or less tinged with bilirubin.

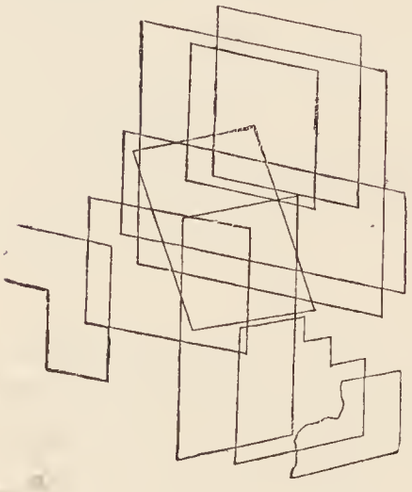


FIG. 25.—Cholesterin crystals.

Though its solubilities remind one of a fat, cholesterin is not a fat. It is, in fact, chemically speaking, a monatomic alcohol. Its formula is $C_{27}H_{45}.HO$.

From alcohol or ether containing water it crystallises in the form of rhombic tables, which contain a molecule of water of crystallisation: these are easily recognised under the microscope (fig. 25). It gives the tests described under the practical exercise at the head of this lesson.

'The mode of origin of cholesterin in the body has not been clearly made out. Whether it is formed in the tissues generally, in the blood, or in the liver, is not known; nor has it been determined conclusively that it is derived from albuminous or nervous matter. It is also doubtful if we are to regard it as a waste substance of no use to the body, as its presence in the blood corpuscles, in nervous matter, in the egg, and in vegetable grains, points to a possible function of a histogenetic or tissue-forming character.' (McKendrick.)

A substance called iso-cholesterin, isomeric with ordinary cholesterin, is found in the fatty secretion of the skin (sebum); it is largely contained in the preparation called lanoline made from sheep's-wool fat. It does not give Salkowski's reaction.

THE USES OF BILE

One of the most remarkable facts concerning the bile is its apparently small use in the digestion of food. It is doubtless, to a large extent, excretory. In some animals it has a slight action on fats and starch, but it appears to be rather a coadjutor to the pancreatic juice (especially in the digestion of fat) than to have any

independent digestive activity. Its auxiliary action in starch digestion has been shown in one of our practical exercises (p. 69).

Bile is said to be a natural antiseptic, lessening the putrefactive processes in the intestine. This is also very doubtful. Though the bile salts are weak antiseptics, the bile itself is readily putrescible, and the power it has of diminishing putrescence in the intestine is due chiefly to the fact that by increasing absorption it lessens the amount of putrescible matter in the bowel.

When the bile meets the chyme the turbidity of the latter is increased, owing to the precipitation of unpeptonised proteid. This is an action due to the bile salts, and it has been surmised that this conversion of the chyme into a more viscid mass is to hinder somewhat its progress through the intestines ; it clings to the intestinal wall, thus allowing absorption to take place. The neutralisation of the acid gastric juice by the bile also allows the alkalinity of the pancreatic juice to have full play. Bile is a solvent of fatty acids, and assists the absorption of fat (see p. 91).

THE FATE OF THE BILIARY CONSTITUENTS

We have seen that fistula bile is poor in solids as compared with normal bile, and that this is explained on the supposition that the normal bile circulation is not occurring—the liver cannot excrete what it does not receive back from the intestine. Schiff was the first to show that if the bile is led back into the duodenum, or even if the animal is fed on bile, the percentage of solids in the bile excreted is at once raised. It is on these experiments that the theory of a bile circulation is mainly founded. The bile circulation relates, however, chiefly, if not entirely, to the bile salts : they are found but sparingly in the fæces ; they are only represented to a slight extent in the urine ; hence it is calculated that seven-eighths of them are reabsorbed from the intestine. Small quantities of cholalic acid, taurine, and glycine are found in the fæces ; the greater part of these products of the decomposition of the bile salts is taken by the portal vein to the liver, where they are once more synthesised into the bile salts. Some of the taurine is absorbed and excreted as tauro-carbamic acid ($C_2H_4NHCO.NH_2HSO_3$) in the urine. Some of the absorbed glycine may be excreted as urea or uric acid. The cholesterin and mucus are found in the fæces ; the pigment is changed into stercobilin, a substance like hydrobilirubin. Some of the stercobilin is absorbed, and leaves the body as the urinary pigment, urobilin.

THE FÆCES

The fæces are alkaline in reaction, and contain the following substances:—

1. Water: in health from 68 to 82 per cent.; in diarrhœa it is more abundant still.

2. Undigested food; that is, if food is taken in excess, some escapes the action of the digestive juices. On a moderate diet unaltered proteid is never found.

3. Indigestible constituents of the food: cellulose, keratin, mucin, chlorophyll, gums, resins, cholesterin.

4. Constituents digestible with difficulty: uncooked starch, tendons, elastin, various phosphates, and other salts of the alkaline earths.

5. Products of decomposition of the food: indole, skatole, phenol, acids such as fatty acids, lactic acid, &c.; hæmatin from hæmoglobin; insoluble soaps like those of calcium and magnesium.

6. Bacteria of all sorts and *débris* from the intestinal wall; cells, nuclei, mucus, &c.

7. Bile residues: mucus, cholesterin, traces of bile acids and their products of decomposition, stercobilin from the bile pigment.[1]

MECONIUM

Meconium is the name given to the greenish-black contents of the intestine of new-born children. It is chiefly concentrated bile, with *débris* from the intestinal wall. The pigment is a mixture of bilirubin and biliverdin; it is not stercobilin.

ABSORPTION

Food is digested in order that it may be absorbed. It is absorbed in order that it may be assimilated—that is, become an integral part of the living material of the body.

Having now considered the action of digestive juices, we can study the absorption which follows. In the mouth and œsophagus the thickness of the epithelium and the quick passage of the food through these parts reduce absorption to a minimum. Absorption takes place more rapidly in the stomach: the small intestine, with its folds and villi to increase its surface, is, however, the great place for absorption; and although the villi are absent from the large intestine, absorption occurs there also, but to a less extent.

Foods such as water and soluble salts like sodium chloride are

[1] Stercobilin may originate also from the hæmatin of the food. (MacMunn.)

absorbed unchanged. The organic foods, however, are considerably changed, colloid materials like starch and proteid being converted respectively into the diffusible materials sugar and peptone.

There are two channels of absorption, the blood vessels (portal capillaries) and the lymphatic vessels or lacteals.

Absorption, however, is no mere physical process of diffusion and filtration. We must also take into account the fact that the cells through which the absorbed substances pass are living, and in virtue of their vital activity not only select materials for absorption, but also change those substances while in contact with them. These cells are of two kinds—(1) the columnar epithelium that covers the surface; and (2) the lymph cells in the lymphoid tissue beneath. It is now generally accepted that of the two the former, the columnar epithelium, is the more important. When these cells are removed or rendered inactive by sodium fluoride, absorption practically ceases, though the opportunities for simple filtration or diffusion would be by such means increased.

Absorption of Carbohydrates.—Though the sugar formed from starch by ptyalin and amylopsin is maltose, that found in the blood is glucose. Under normal circumstances little if any is absorbed by the lacteals. The glucose is formed from the maltose by the succus entericus, and perhaps also by the vital action of the epithelial cells. Cane sugar and milk sugar are also converted into glucose before absorption.

The carbohydrate food which enters the blood as glucose is taken to the liver, and there stored up in the form of glycogen—a reserve store of carbohydrate material for the future needs of the body. Glycogen, however, is found in animals which take no carbohydrate food. It must, then, be formed by the protoplasmic activity of the liver cells from their proteid constituents. The carbohydrate store leaves the liver in the blood of the hepatic vein as glucose (dextrose) once more.

The above is a brief statement of the glycogenic functions of the liver as taught by Claude Bernard, and accepted by the majority of physiologists. It has always been strongly contested by Pavy, who holds that the glycogen formed in the liver from the sugar of the portal blood is never during life reconverted into sugar, but is used in the formation of other substances like fat and proteid; in support of this he has shown that proteids contain a carbohydrate radicle. He denies that the *post-mortem* formation of sugar from glycogen that occurs in an excised liver is a true picture of what occurs during life.

Absorption of Proteids.—A certain amount of soluble proteid is absorbed unchanged. Thus, after taking a large number of eggs, egg albumin is found in the urine. Patients fed *per rectum* derive nourishment from proteid food, though proteolytic ferments are not present in this part of the intestine.

Most proteid, however, is normally absorbed as peptone and proteose (albumose). Peptones and proteoses are absent from the blood under all circumstances, even from the portal blood during the most active digestion. In other words, during absorption the epithelial cells change the products of proteolysis (peptones and

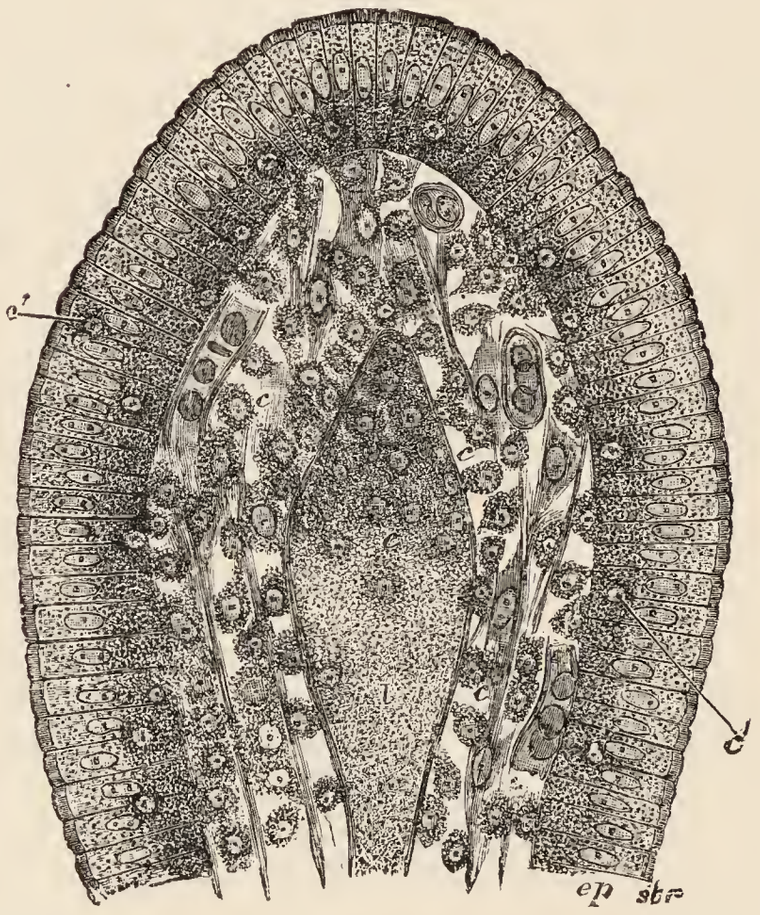


FIG. 26.—Section of the villus of a rat killed during fat-absorption (E. A. Schäfer): *ep*, epithelium; *str*, striated border; *c*, lymph cells; *c'*, lymph cells in the epithelium; *l*, central lacteal containing disintegrating lymph cells.

proteoses) back once more into native proteids (albumin and globulin).

The greater part of the proteid absorbed passes into the blood; a little into the lymph vessels also; but this undergoes the same change.

When peptone (using the word to include the proteoses also) is injected into the blood stream, poisonous effects are produced, the coagulability of the blood is lessened, the blood pressure falls, secretion ceases, and in the dog 0·3 gramme of 'peptone' per kilogramme of body weight is sufficient to kill the animal.

The epithelial cells of the alimentary canal thus protect us from those poisonous effects by converting the harmful peptone into the useful albumin and globulin.

The question of proteid absorption is in a rather unsettled state. Some recent observations lend support to the view that the peptones are very largely broken up into simpler substances, but the absence of these in the blood stream shows that the absorptive epithelium is capable of resynthesising them into proteids. The small amount of peptones obtainable from the intestinal contents may be due to the fact that they are so rapidly absorbed, or it may be due to their having been broken up into crystalline products (leucine, tyrosine, &c.) by trypsin and erepsin.

Absorption of Fats.—The fats undergo in the intestine two changes : one a physical change (emulsification), the other a chemical change (saponification). The lymphatic vessels are the great channels for fat absorption, and their name, lacteals, is derived from the milk-like appearance of their contents (chyle) during the absorption of fat.

The way in which the minute fat globules pass from the intestine into the lacteals has been the subject of much controversy. The course they take may be studied by killing animals at varying periods after a meal of fat, and making osmic acid microscopic preparations of the villi. Figs. 26 and 27 illustrate the appearances observed by Schäfer.

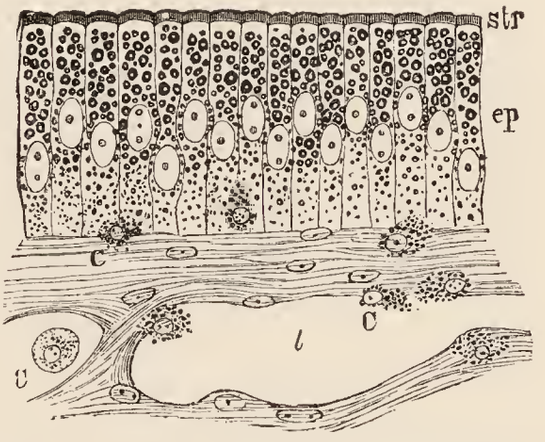


FIG. 27.—Mucous membrane of frog's intestine during fat absorption (E. A. Schäfer) : *ep*, epithelium ; *str*, striated border ; c, lymph cells ; *l*, lacteal.

The columnar epithelium cells become first filled with fatty globules of varying size, which are generally larger near the free border. The globules pass down the cells, the larger ones breaking up into smaller ones during the journey ; they are then transferred to the amœboid cells of the lymphoid tissue beneath ; these ultimately penetrate into the central lacteal, where they either disintegrate or discharge their cargo into the lymph stream. The globules are by this time divided into immeasurably small ones, the molecular basis of chyle. The chyle enters the blood stream by the thoracic duct, and after an abundant fatty meal the blood plasma is quite milky ; the fat droplets are so small that they circulate without hindrance through the capillaries. The fat in the blood after a meal is eventually stored up in connective tissue cells of adipose tissue. It must, however, be borne in mind that the fat of the body is not exclusively derived

from the fat of the food, but it may originate also both from proteid and from carbohydrates.

As the fat globules were never seen penetrating the striated border of the epithelial cells, there was a great difficulty in understanding how they reached the interior of these cells; the cells will not take up other particles, and it appears certain that they do not in the higher animals protrude pseudopodia from their borders (this, however, does occur in the endoderm of some of the lower invertebrates).

Recent research has in a great measure solved this difficulty. In the first place particles may be present in the epithelium and lymphoid cells while no fat is being absorbed. These particles are protoplasmic in nature, as they stain with reagents that stain protoplasmic granules; but as they also stain darkly with osmic acid, they are apt to be mistaken for fat. There is, however, no doubt that the particles found during fat absorption are composed of fat. There is also no doubt that the epithelial cells have the power of again forming fat out of the fatty acids and glycerin into which it has been broken up in the intestine. Munk, who performed a large number of experiments on the subject, showed that the splitting of fats into glycerin and fatty acids occurs to a much greater extent than was formerly supposed; these substances, being soluble, pass readily into the epithelium cells, and these cells perform the synthetic act of building them into fat once more; the fat so formed appears in the form of small globules, surrounding or becoming mixed with the protoplasmic granules that are ordinarily present. Another remarkable fact which he made out is that after feeding an animal on fatty acids the chyle contains fat. The necessary glycerin must have been formed by protoplasmic activity during absorption. The more recent work of Moore and Rockwood has shown that fat is absorbed entirely as fatty acid or soap; and that preliminary emulsification, though advantageous for the formation of these substances, is not essential.

We thus see how with increase of knowledge, due to improved methods of research, a complete change has come over the ideas which physiologists have held regarding this subject. It is not so many years ago that the physical change—emulsification—which fats undergo in the intestine, was considered to be more important than the chemical changes—fat-splitting and saponification. In fact the small amount of chemical change which was supposed to occur was regarded as quite subordinate, and of value merely in assisting the process of emulsification. We now know that the exact converse is

the truth ; the chemical change is the important process, and emulsification the subordinate one.

Bile aids the digestion of fat, in virtue of its being a solvent of fatty acids, and it probably assists fat absorption by reducing the surface tension of the intestinal contents ; membranes moistened with bile allow fatty materials to pass through them more readily than would otherwise be the case. In cases of disease in which bile is absent from the intestines, a large proportion of the fat in the food passes into the fæces.

LESSON IX

THE BLOOD

Blood Plasma

1. The coagulation of the blood has been prevented in specimen A by the addition of neutral salt (an equal volume of saturated sodium-sulphate solution, or a quarter of its volume of saturated magnesium-sulphate solution). The corpuscles have settled, and the supernatant salted plasma has been siphoned off.

2. The coagulation of the blood in specimen B has been prevented by the addition of an equal volume of a 0·4-per-cent. solution of potassium oxalate in normal saline solution.

3. Put a small quantity of A into three test-tubes and dilute each with about ten times its volume of liquid:

A 1. With distilled water.

A 2. With solution of fibrin ferment containing a little calcium chloride.[1]

A 3. With the same.

4. Put A 1 and A 2 into the water-bath at 40° C.; leave A 3 at the temperature of the air. A 1 coagulates slowly or not at all; A 2 coagulates rapidly; A 3 coagulates less rapidly than A 2.

5. Add to some of B a few drops of dilute (2 per cent.) calcium chloride solution: it coagulates, and more quickly, if the temperature is 40° C.

Blood Serum

Blood serum is the fluid residue of the blood after the separation of the clot; it is blood plasma *minus* fibrin. The general appearance of fibrin obtained by whipping fresh blood will already be familiar to the student, as he has used it in experiments on digestion.

Serum has a yellowish tinge due to serum lutein, but as generally obtained it is often contaminated with a small amount of oxyhæmoglobin, and so looks reddish. It contains proteids (giving the general tests already studied in Lesson IV.), extractives, and salts in solution. The proteids are serum albumin and serum globulin. The fibrin ferment is also a proteid-like substance. It is present in only small quantities, and in the following experiments is precipitated with serum globulin.

Separation of the serum proteids.—(*a*) Dilute serum with fifteen times its volume of water. It becomes cloudy owing to the partial precipitation of

[1] An easy way of preparing an impure but efficient solution of fibrin ferment is to take 5 c.c. of blood serum and dilute it with a litre of distilled water. A partial precipitation of globulin takes place, and carries down the ferment with it. After a few hours pour off the supernatant fluid and dissolve the precipitate in half a litre of tap water to which a few drops of 2-per-cent. solution of calcium chloride have been added. This solution can be then given round to the class as fibrin ferment.

serum globulin. Add a few drops of 2-per-cent. acetic acid; the precipitate becomes more abundant, but dissolves in excess of the acid.

(*b*) Pass a stream of carbonic acid through serum diluted with twenty times its bulk of water. A partial precipitation of serum globulin occurs.

(*c*) Saturate some serum with magnesium sulphate by adding crystals of the salt and grinding in a mortar. A precipitate of serum globulin is produced.

(*d*) Half saturate the serum with ammonium sulphate by adding to it an equal volume of a saturated solution of the salt. Serum globulin is precipitated.

(*e*) Completely saturate the serum with ammonium sulphate by adding crystals of the salt and grinding in a mortar; a precipitate is produced of both the globulin and the albumin. Filter through a dry filter paper; the filtrate contains no proteid.

Hæmoglobin

6. Direct the spectroscope to the window and carefully focus Fraunhofer's lines. Note especially D in the yellow, and E, the next well-marked line, in the green.

7. Direct the spectroscope to a luminous gas flame: these lines are absent. Place a little sodium chloride in the flame. Notice the bright yellow line in the position of the D line.

8. Take a series of six test-tubes of about equal size. Fill the first with diluted defibrinated ox-blood (1 part of blood to 31 of water); then fill the second tube with the same mixture diluted with an equal bulk of water (1 in 64); half fill the third tube with this and fill up the tube with an equal bulk of water (1 in 128), and so on. The sixth tube will contain 1 part of blood to 1,024 of water, and will be nearly colourless.

9. Into another series of six test-tubes put a few drops of ammonium sulphide; then pour in some of the contents of each of the first series and warm *very* gently.

10. Examine the tubes with the spectroscope and map out on a chart the typical absorption bands of oxyhæmoglobin in the first series, and of (reduced) hæmoglobin in the second series. Notice that in the more dilute specimens of hæmoglobin the bands are no longer seen, whereas those of oxyhæmoglobin in specimens similarly diluted are still visible.

11. Take a tube which shows the single band of reduced hæmoglobin and shake it with the air; the bright red colour returns to it and it shows spectroscopically the two bands of oxyhæmoglobin for a short time. Continue watching the two bands, and note that they fade and are replaced by a single band as reduction again occurs.

12. Mix a drop of defibrinated rat's blood on a slide with a drop of water, or mount it in a drop of Canada balsam. Examine the crystals of oxyhæmoglobin as they form.

13. Smear a little blood, obtained by pricking the finger, on a slide and allow it to dry; cover, and run glacial acetic acid under the cover glass, and boil. Examine microscopically for the dark brown crystals of hæmin.

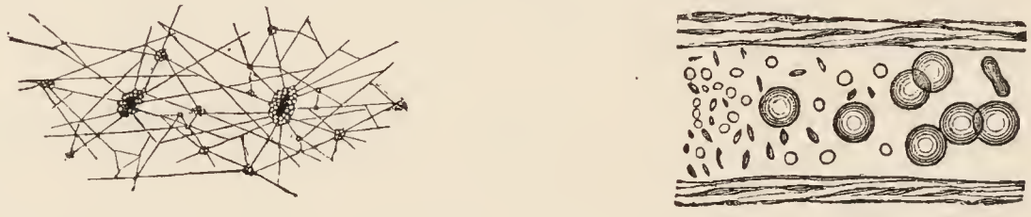
COAGULATION OF BLOOD

Microscopic investigation of vertebrate blood shows that it consists of a fluid which holds in suspension large numbers of solid bodies—the red and the white corpuscles and the blood platelets.

After blood is shed it rapidly becomes viscous and then sets into

a firm red jelly. The jelly soon contracts and squeezes out a straw-coloured fluid called the serum, in which the shrunken clot ultimately floats.

With the microscope, filaments of fibrin are seen forming a network throughout the fluid, many radiating from small clumps of blood platelets. It is the formation of fibrin which is the essential act of coagulation : this entangles the corpuscles and forms the clot. Fibrin is formed from the plasma, and may be obtained free from corpuscles when blood plasma is allowed to clot, the corpuscles having previously been removed. It may also be obtained from blood by whipping it with a bunch of twigs ; the fibrin adheres to the twigs and entangles but few corpuscles. These may be removed by



A B

FIG. 28.—Fibrin filaments and blood platelets : A, network of fibrin shown after washing away the corpuscles from a preparation of blood that has been allowed to clot. Many of the filaments radiate from small clumps of blood platelets. B (from Osler), blood corpuscles and blood platelets within a small vein.

washing with water. Serum is plasma *minus* fibrin. The relation of plasma, serum, and clot can be seen at a glance in the following scheme of the constituents of the blood :—

$$\text{Blood}\begin{cases}\text{Plasma}\begin{cases}\text{Serum}\\ \text{Fibrin}\end{cases}\\ \text{Corpuscles}\end{cases}\Bigg\}\text{Clot}$$

It may be roughly stated that in 100 parts by weight of blood 60–65 parts consist of plasma and 35–40 of corpuscles.

The *buffy coat* is seen when blood coagulates slowly, as in horse's blood. The red corpuscles sink more rapidly than the white, and the upper stratum of the clot (buffy coat) consists mainly of fibrin and white corpuscles.

Coagulation is hastened by—

1. A temperature a little over that of the body.
2. Contact with foreign matter.
3. Injury to the vessel walls.
4. Agitation.
5. Addition of calcium salts.
6. Injection of nucleo-proteid produces intravascular clotting

(positive phase). Very minute doses, however, produce the opposite effect—namely delay of coagulation (negative phase).

Coagulation is hindered or prevented by—

1. A low temperature. In a vessel cooled by ice, coagulation may be prevented for an hour or more.

2. The addition of a large quantity of neutral salts like sodium sulphate or magnesium sulphate.

3. Contact with the living vascular walls.

4. Contact with oil.

5. Addition of a soluble oxalate (*e.g.* potassium oxalate) : this precipitates the calcium necessary for coagulation as insoluble calcium oxalate.

6. Injection of commercial peptone (which consists chiefly of proteoses) into the circulation of the living animal.

7. Addition of leech extract. This acts in virtue of a proteose it contains.

The theory generally received which accounts best for the coagulation of the blood is that of Hammarsten, and it may be briefly stated as follows :

When blood is within the vessels one of the constituents of the plasma, a proteid of the globulin class called fibrinogen, exists in a soluble form.

When the blood is shed the fibrinogen molecule is split into two parts : one part is a globulin, which remains in solution, the other is the insoluble material fibrin.

This change is brought about by the activity of a special unorganised ferment called the fibrin ferment.

This ferment does not exist in healthy blood contained in healthy blood vessels, but is one of the products of the disintegration of the white corpuscles and blood platelets that occurs when the blood leaves the vessels or comes in contact with foreign matter.

To this may be added, as the result of recent research, that a soluble calcium salt is essential for the formation of the ferment ; that the fibrin ferment belongs to the class of nucleo-proteids ; that other nucleo-proteids (Wooldridge's tissue-fibrinogens, see p. 37) obtained from most of the cellular organs of the body produce intravascular clotting when injected into the circulation of a living animal.

The fibrin ferment may very conveniently be called *thrombin*. Like other ferments it is preceded by a mother subtance or zymogen which may be called *prothrombin* ; the action of calcium salts is to convert prothrombin into thrombin. We may therefore represent the process of clotting in the following tabular way :

In the plasma a proteid substance exists, called

FIBRINOGEN.

From the colourless corpuscles a nucleo-proteid is shed out, called

PROTHROMBIN.

By the action of calcium salts prothrombin is converted into fibrin ferment, or

THROMBIN.

Thrombin acts on fibrinogen in such a way that two new substances are formed.

One of these is unimportant, viz. a globulin (fibrino-globulin) which remains in solution. Its amount is very small.

The other is important, viz. FIBRIN, which entangles the corpuscles and so forms the CLOT.

THE PLASMA AND SERUM

The liquid in which the corpuscles float may be obtained by employing one or other of the methods already described for preventing the blood from coagulating. The corpuscles, being heavy, sink, and the supernatant plasma can then be removed by a pipette or siphon; the separation can be effected more thoroughly by the use of a centrifugal machine (see fig. 60, Lesson XXI.).

On counteracting the influence which has prevented the blood from coagulating, the plasma then itself coagulates. Thus plasma obtained by the use of cold clots on warming gently; plasma which has been decalcified by the action of a soluble oxalate clots on the addition of a calcium salt; plasma obtained by the use of a strong solution of salt coagulates when this is diluted by the addition of water, the addition of fibrin ferment being necessary in most cases; where coagulation occurs without the addition of fibrin ferment, no doubt some is present from the partial disintegration of the corpuscles which has already occurred. Pericardial and hydrocele fluid resemble pure plasma very closely in composition. As a rule, however, they contain few or no white corpuscles, and do not clot spontaneously, but after the addition of fibrin ferment or liquids like serum that contain fibrin ferment they always yield fibrin.

Pure plasma may be obtained from horse's veins by what is known as the 'living test-tube' experiment. If the jugular vein is ligatured in two places, so as to include a quantity of blood within it, then removed from the animal and hung in a cool place, the blood will not

coagulate for many hours. The corpuscles settle, and the supernatant plasma can be removed with a pipette.

The plasma is alkaline, yellowish in tint, and its specific gravity is about 1026 to 1029.

Its chief constituents may be enumerated as follows :—

1,000 parts of plasma contain—

Water	902·90
Solids	97·10
Proteids : 1, yield of fibrin . . .	4·05
2, other proteids . . .	78·84
Extractives (including fat) . . .	5·66
Inorganic salts	8·55

In round numbers plasma contains 10 per cent. of solids, of which 8 per cent. are proteid in nature. The small quantity of fibrinogen as indicated by the amount of fibrin formed should be carefully noted.

Serum contains the same three classes of constituents—proteids, extractives, and salts. The extractives and salts are the same in the two liquids. The proteids differ, as is shown in the following table :—

Proteids of Plasma	*Proteids of Serum*
Fibrinogen	Serum globulin
Serum globulin	Serum albumin
Serum albumin	Fibrin ferment (nucleo-proteid)
	Fibrino-globulin

The gases of the plasma and serum are small quantities of oxygen, nitrogen, and carbonic acid. The greater part of the oxygen of the blood is combined in the red corpuscles with hæmoglobin ; the carbonic acid is chiefly combined as carbonates (see RESPIRATION).

We may now consider one by one the various constituents of the plasma and serum.

A. **Proteids.**—*Fibrinogen.*—This is the substance acted on by fibrin ferment. It yields, under this action, an insoluble product called fibrin, and a soluble proteid of the globulin class.

Fibrinogen is a globulin. It differs from serum globulin, and may be separated from it by the fact that half saturation with sodium chloride precipitates it. It coagulates by heat at the low temperature of 56° C. As judged from the yield of fibrin, it is the least abundant of the proteids of the plasma (see table on upper part of this page).

Serum globulin and serum albumin.—These substances are considered in the practical exercises at the head of this lesson : see also

Lesson IV. Both serum globulin and serum albumin probably consist of more than one proteid substance (see Lesson XX.).

Fibrin ferment.—Schmidt's method of preparing it is to take serum and add excess of alcohol. This precipitates all the proteids, fibrin ferment included. After some weeks the alcohol is poured off; the serum globulin and serum albumin have been by this means rendered insoluble in water; an aqueous extract is, however, found to contain fibrin ferment, which is not so easily coagulated by alcohol as the other proteids are. A simpler method of preparing fibrin ferment in an impure but efficient form is given in the footnote on p. 92.

B. **Extractives.**—These are non-nitrogenous and nitrogenous. The non-nitrogenous are sugar (0·12 per cent.), fats, soaps, cholesterin; and the nitrogenous are urea (0·02 to 0·04 per cent.), and still smaller quantities of uric acid, creatine, creatinine, xanthine, and hypoxanthine.

C. **Salts.**—The most abundant salt is sodium chloride: it constitutes between 60 and 90 per cent. of the total mineral matter. Potassium chloride is present in much smaller amount. It constitutes about 4 per cent. of the total ash. The other salts are phosphates and sulphates.

Schmidt gives the following table:—

1,000 parts of plasma yield—

Mineral matter	8·550
Chlorine	3·640
SO_3	0·115
P_2O_5	0·191
Potassium	0·323
Sodium	3·341
Calcium phosphate	0·311
Magnesium phosphate	0·222

THE WHITE BLOOD CORPUSCLES

These corpuscles are typical animal cells. Their nucleus consists of nuclein; their cell-protoplasm yields proteids belonging to the nucleo-proteid and globulin groups. The nucleo-proteid obtained from them is the zymogen of the fibrin ferment, the addition of a calcium salt converting it into the ferment. The protoplasm of these cells often contains small quantities of fat and glycogen.

THE RED BLOOD CORPUSCLES

The red blood corpuscles are much more numerous than the white, averaging in man 5,000,000 per cubic millimetre, or 400 to 500 red to each white corpuscle. The method of enumeration of the corpuscles is described in the Appendix.

They vary in size and structure in different groups of vertebrates. In mammals they are biconcave (except in the camel tribe, where they are biconvex) non-nucleated discs, in man averaging $\frac{1}{3200}$ inch in diameter ; during fœtal life nucleated red corpuscles are, however, found. In birds, reptiles, amphibians, and fishes they are biconvex oval discs with a nucleus : they are largest in the amphibia.

Water causes the corpuscles to swell up, and dissolves out the red pigment (oxyhæmoglobin), leaving a globular colourless stroma. *Salt solution* causes the corpuscles to shrink : they become crenated or wrinkled. The action of water and salt solution suggests the existence of a membrane on the surface of the corpuscles through which osmosis takes place, but the existence of such a membrane is still a matter of discussion. If there is no actual membrane, the outer denser portion of the stroma plays the *rôle* of one during osmotic phenomena. *Dilute alkalis* (0·2 per cent. potash) dissolve the corpuscles. *Dilute acids* (1 per cent. acetic acid) act like water, and in nucleated corpuscles render the nucleus distinct. *Tannic acid* causes a discharge of hæmoglobin from the stroma, but this is immediately altered and precipitated. It remains adherent to the stroma as a brown globule, consisting probably of hæmatin. *Boric acid* acts similarly, but in nucleated red corpuscles the pigment collects chiefly round the nucleus, which may then be extruded from the corpuscles.



FIG. 29.—*a–e*, successive effects of water on a red blood corpuscle ; *f*, a red corpuscle crenated by salt solution ; *g*, action of tannin on a red corpuscle.

Composition.—1,000 parts of red corpuscles contain—

Water		688 parts
Solids	organic	303·88 „
	inorganic	8·12 „

100 parts of dried corpuscles contain—

Proteid	5 to 12 parts
Hæmoglobin	86 „ 94 „
Lecithin	1·8 „
Cholesterin	0·1 part

The proteid present appears to be identical with the nucleo-proteid of white corpuscles. The mineral matter consists chiefly of chlorides of potassium and sodium, and phosphates of calcium and magnesium. In man potassium chloride is more abundant than sodium chloride ; this, however, does not hold good for all animals.

Oxygen is contained in combination with the hæmoglobin to form oxyhæmoglobin. The corpuscles also contain a certain amount of carbonic acid (see RESPIRATION, at the end of this lesson).

The pigment of the red corpuscles.—The pigment is by far the most abundant and important of the constituents of the red corpuscles. It is a substance which gives the reactions of a proteid, but differs from most other proteids in containing the element iron; it is also readily crystallisable.

It exists in the blood in two conditions: in arterial blood it is combined loosely with oxygen, is of a bright red colour, and is called oxyhæmoglobin; the other condition is the deoxygenated or reduced hæmoglobin (better called simply hæmoglobin). This is found in the blood after asphyxia. It also occurs in all venous blood—that is, blood which is returning to the heart after it has supplied the tissues with oxygen. Venous blood, however, always contains a considerable quantity of oxyhæmoglobin also. Hæmoglobin is the oxygen-carrier of the body, and it may be called a respiratory pigment.

Crystals of oxyhæmoglobin may be obtained with readiness from the blood of such animals as the rat, guinea-pig, or dog; with difficulty from other animals, such as man, ape, and most of the common mammals. The following methods are the best:—

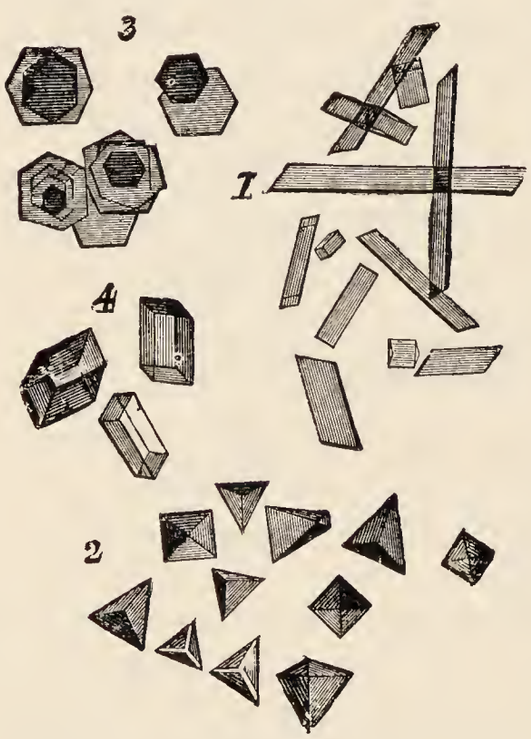


FIG. 30.—Oxyhæmoglobin crystals magnified: 1, from human blood; 2, from the guinea-pig; 3, squirrel; 4, hamster.

1. Mix a drop of defibrinated blood of the rat on a slide with a drop of water; put on a cover glass; in a few minutes the corpuscles are rendered colourless, and then the oxyhæmoglobin crystallises out from the solution so formed.

2. Microscopical preparations may also be made by Stein's method, which consists in using Canada balsam instead of water in the above experiment.

3. On a larger scale the crystals may be obtained by mixing the blood with one-sixteenth of its volume of ether; the corpuscles dissolve and the blood assumes a *laky* appearance. After a period, varying from a few minutes to days, abundant crystals are deposited.

The accompanying figures represent the form of the crystals so obtained.

In nearly all animals the crystals are rhombic prisms; but in the guinea-pig they are rhombic tetrahedra (four-sided pyramids); in the squirrel, hexagonal plates; and in the hamster, rhombohedra and hexagonal plates.

The crystals also contain a varying amount of water of crystallisation: this may in part explain their different crystalline forms and solubilities.

Different observers have analysed hæmoglobin. They find carbon, hydrogen, nitrogen, oxygen, sulphur, and iron. The percentage of iron is 0·4. The amounts of the other elements are variously given, but, roughly, they are the same as in the proteids. We know at present as little of the chemical structure of hæmoglobin as of the proteids generally.

Oxyhæmoglobin may be estimated in the blood (1) by the amount of iron in the ash, or (2) by certain colorimetric methods which are described in the Appendix.

On adding an acid or alkali to hæmoglobin, it is broken up into two parts, a proteid called *globin*, and a brown pigment called *hæmatin*, which contains all the iron of the original substance.

Globin is a somewhat unique proteid. It is coagulable by heat, soluble in dilute acids, and precipitable from such solutions by ammonia. It closely resembles a substance previously separated from red corpuscles by Kossel and termed by him *histone* (Schulz).

Hæmatin is not crystallisable; according to Hoppe-Seyler its formula is $C_{34}H_{35}N_4FeO_5$; other observers give different formulæ. It presents different spectroscopic appearances in acid and alkaline solutions, and yields several products under the influence of certain reagents, which we shall consider in the advanced course. For the present, we will mention only two of these, hæmin and hæmatoporphyrin.

Hæmin is of great importance, as the obtaining of this substance in a crystalline form is the best chemical test for blood. Hæmin crystals, sometimes called Teichmann's crystals, are composed of the hydrochloride of hæmatin. They may be prepared for microscopic examination by boiling a fragment of dried blood with a drop of glacial acetic acid on a slide; on cooling, dark-brown plates and prisms belonging to the triclinic system, often in star-shaped clusters and with rounded angles (fig. 31), separate out.

In the case of an old blood-stain it is necessary to add a crystal of sodium chloride. Fresh blood contains sufficient sodium chloride in itself. The action of the acetic acid is (1) to split the hæmoglobin into hæmatin and globin; and (2) to evolve hydro-

chloric acid from the sodium chloride. The hæmatin unites with the hydrochloric acid, and thus hæmin is formed. The formula for hæmin is $C_{35}H_{35}N_4FeClO_4$ (Mörner).

Hæmatoporphyrin is iron-free hæmatin; it may be prepared by mixing blood with strong sulphuric acid: the iron is taken out as ferrous sulphate. This substance is also found sometimes in nature: it occurs in certain invertebrate pigments, and may also be found in certain forms of pathological urine. It shows well-marked spectroscopic bands, and so is not identical with the iron-free derivative of hæmoglobin called hæmatoidin which is formed in extravasations of blood in the body (see p. 81). The two substances are possibly isomeric.



Fig. 31.—Hæmin crystals magnified. (Preyer.)

COMPOUNDS OF HÆMOGLOBIN WITH GASES

Hæmoglobin forms at least four compounds with gases:—

With oxygen	1. Oxyhæmoglobin.
	2. Methæmoglobin.
With carbonic oxide	3. Carbonic oxide hæmoglobin.
With nitric oxide	4. Nitric oxide hæmoglobin.

These compounds have similar crystalline forms: each probably consists of a molecule of hæmoglobin combined with one of the gas. They part with the combined gas somewhat readily, and are arranged in order of stability in the above list, the least stable first.

Oxyhæmoglobin is the compound that exists in arterial blood. Many of its properties have been already mentioned. The oxygen linked to the hæmoglobin, which is removed by the tissues through which the blood circulates, may be called the *respiratory oxygen of hæmoglobin*. The processes that occur in the lungs and tissues, resulting in the oxygenation and deoxygenation respectively of the hæmoglobin, may be imitated outside the body, using either blood or pure solutions of hæmoglobin. The respiratory oxygen can be removed, for example, in the Torricellian vacuum of a mercurial air-pump, or by passing a neutral gas like hydrogen through the blood, or by the use of reducing agents like ammonium sulphide or Stokes's

reagent.[1] One gramme of hæmoglobin will combine with 1·34 c.c. of oxygen.

If any of these methods for reducing oxyhæmoglobin is used, the bright red (arterial) colour of oxyhæmoglobin changes to the purplish (venous) tint of hæmoglobin. On once more allowing oxygen to come into contact with the hæmoglobin, as by shaking the solution with the air, the bright arterial colours return.

These colour-changes may be more accurately studied with the spectroscope, and the constant position of the absorption bands seen constitutes an important test for blood pigment. It will be first necessary to describe briefly the instrument used.

The Spectroscope.—When a ray of white light is passed through a prism, it is refracted or bent at each surface of the prism; the whole ray is, however, not equally bent, but it is split into its constituent colours, which may be allowed to fall on a screen. The band of colours beginning with the red, passing through orange, yellow, green, blue, and ending with violet, is called a *spectrum*: this is seen in nature in the rainbow. It may be obtained artificially by the glass prism or prisms of a spectroscope.

The spectrum of sunlight is interrupted by numerous dark lines crossing it vertically, called Fraunhofer's lines. These are perfectly constant in position, and serve as landmarks in the spectrum. The more prominent are A, B, and C, in the red; D, in the yellow; E, *b*, and F, in the green; G and H, in the violet. These lines are due to certain volatile substances in the solar atmosphere. If the light from burning sodium or its compounds is examined spectroscopically, it will be found to give a bright yellow line, or rather two bright yellow lines very close together. Potassium gives two bright red lines and one violet line; and the other elements, when incandescent, give characteristic lines, but none so simple as sodium. If now the flame of a lamp be examined, it will be found to give a continuous spectrum like that of sunlight in the arrangement of its colours, but unlike it in the absence of dark lines; but if the light from the lamp be made to pass through sodium vapour before it reaches the spectroscope, the bright yellow light will be found absent, and in its place a dark line, or rather two dark lines very close together, occupying the same position as the two bright lines of the sodium spectrum. The sodium vapour thus absorbs the same rays as those which it itself produces at a higher temperature. Thus

[1] Stokes's reagent must always be freshly prepared: it is a solution of ferrous sulphate to which a little tartaric acid has been added, and then ammonia till the reaction is alkaline.

the D line, as we term it, in the solar spectrum is due to the presence of sodium vapour in the solar atmosphere. The other dark lines are similarly accounted for by other elements.

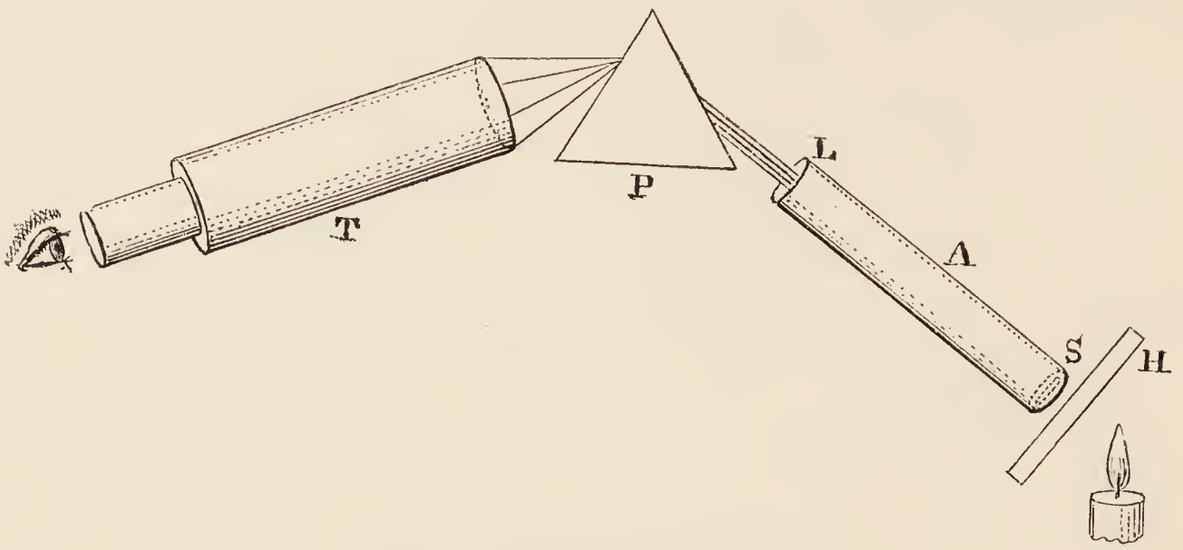


Fig. 32.—Diagram of spectroscope.

The large form of spectroscope (fig. 32) consists of a tube A, called the collimator, with a slit at the end S, and a convex lens at the end L. The latter makes the rays of light passing through the slit from the

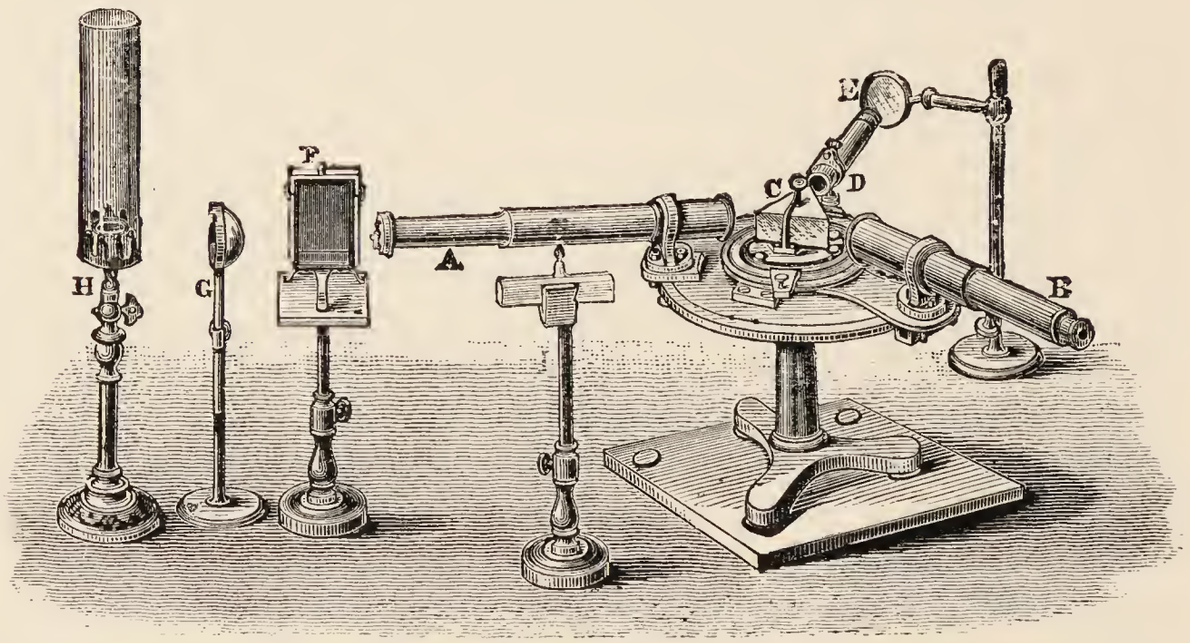


Fig. 33.—Spectroscope: A, collimator with adjustable slit at one (left) end and collimating lens at the other (right) end; B, telescope moving on graduated arc divided into degrees; C, prism or combination of prisms; D, tube for scale; E, mirror for illuminating scale; F, vessel with parallel glass sides for holding fluid, shown with the flat side towards the reader; I, long spectroscope bottle for examining a deep layer of fluid; H, Argand burner; G, condenser for concentrating the light from H on the slit. (From a photograph taken by Dr. MacMunn for McKendrick's 'Physiology.')

source of light parallel: they fall on the prism P, and then the spectrum so formed is focussed by the telescope T.

The third tube, D, seen in the next figure (fig. 33), carries a small transparent scale of wave-lengths, as in accurate observations the

position of any point in the spectrum is given in the terms of the corresponding wave-lengths.

If we now interpose between the source of light and the slit S a piece of coloured glass (H in fig. 32), or a solution of a coloured substance contained in a vessel with parallel sides (the hæmatoscope of Hermann, F in fig. 33), the spectrum is found to be no longer continuous, but is interrupted by a number of dark shadows, or *absorption bands*, corresponding to the light absorbed by the coloured medium. Thus a solution of oxyhæmoglobin of a certain strength gives two bands between the D and E lines; hæmoglobin gives only one; and other red solutions, though to the naked eye similar to oxyhæmoglobin, will give characteristic bands in other positions.

A convenient form of small spectroscope is the *direct-vision spectroscope*, in which, by an arrangement of alternating prisms of crown and flint glass, placed as in fig. 34, the spectrum is

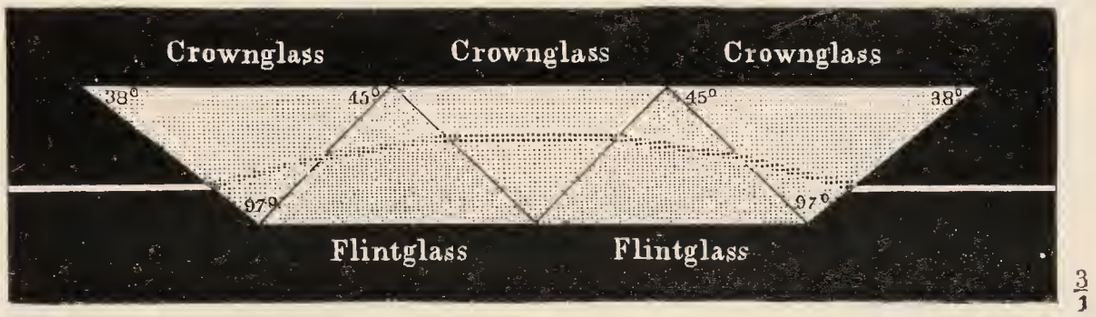


Fig. 34.—Arrangement of prisms in direct-vision spectroscope.

observed by the eye in the same line as the tube furnished with the slit.

Such small spectroscopes may be used for class purposes, and may for convenience be mounted on a stand provided with a gas-burner and a receptacle for the test-tube (see fig. 35).

In the examination of the spectrum of small coloured objects, a combination of the microscope and direct-vision spectroscope, called the *micro-spectroscope*, is used.

Fig. 36 illustrates a method of representing absorption spectra diagrammatically. The solution was examined in a layer 1 centimetre thick. The base line has on it at the proper distances the chief Fraunhofer lines, and along the right-hand edges are percentages of the amount of oxyhæmoglobin present in I, of hæmoglobin in II. The width of the shadings at each level represents the position and amount of absorption corresponding to the percentages.

The characteristic spectrum of oxyhæmoglobin, as it actually

appears through the spectroscope, is seen in the next figure (fig. 37, spectrum 2). There are two distinct absorption bands between the

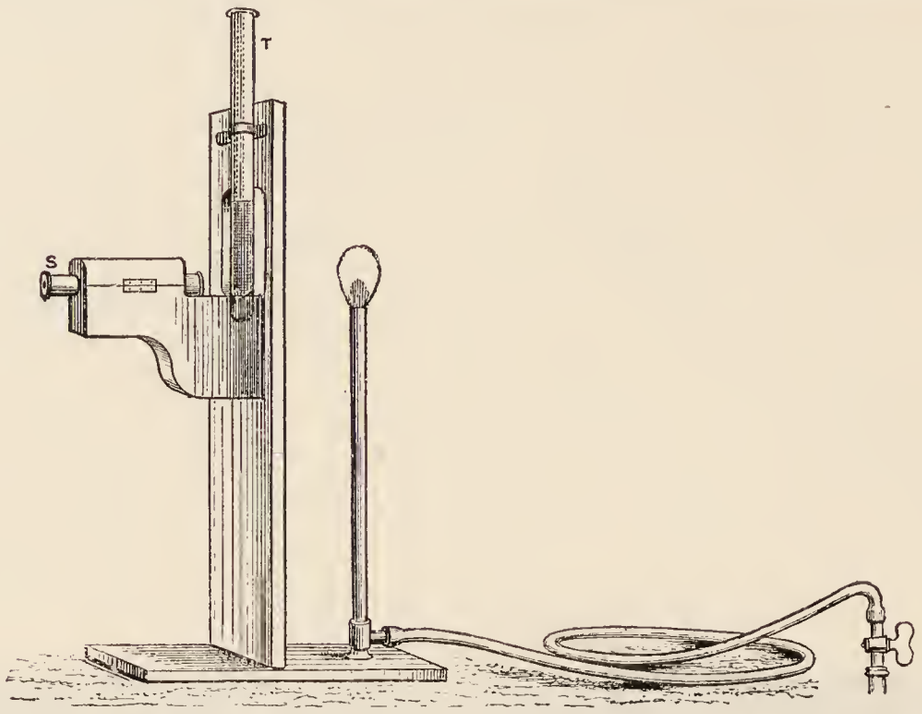


FIG. 35.—Stand for direct-vision spectroscope: S, spectroscope; T, test-tube for coloured substance under investigation.

D and E lines; the one nearest to D (the α band) is narrower, darker, and has better defined edges than the other (the β band).

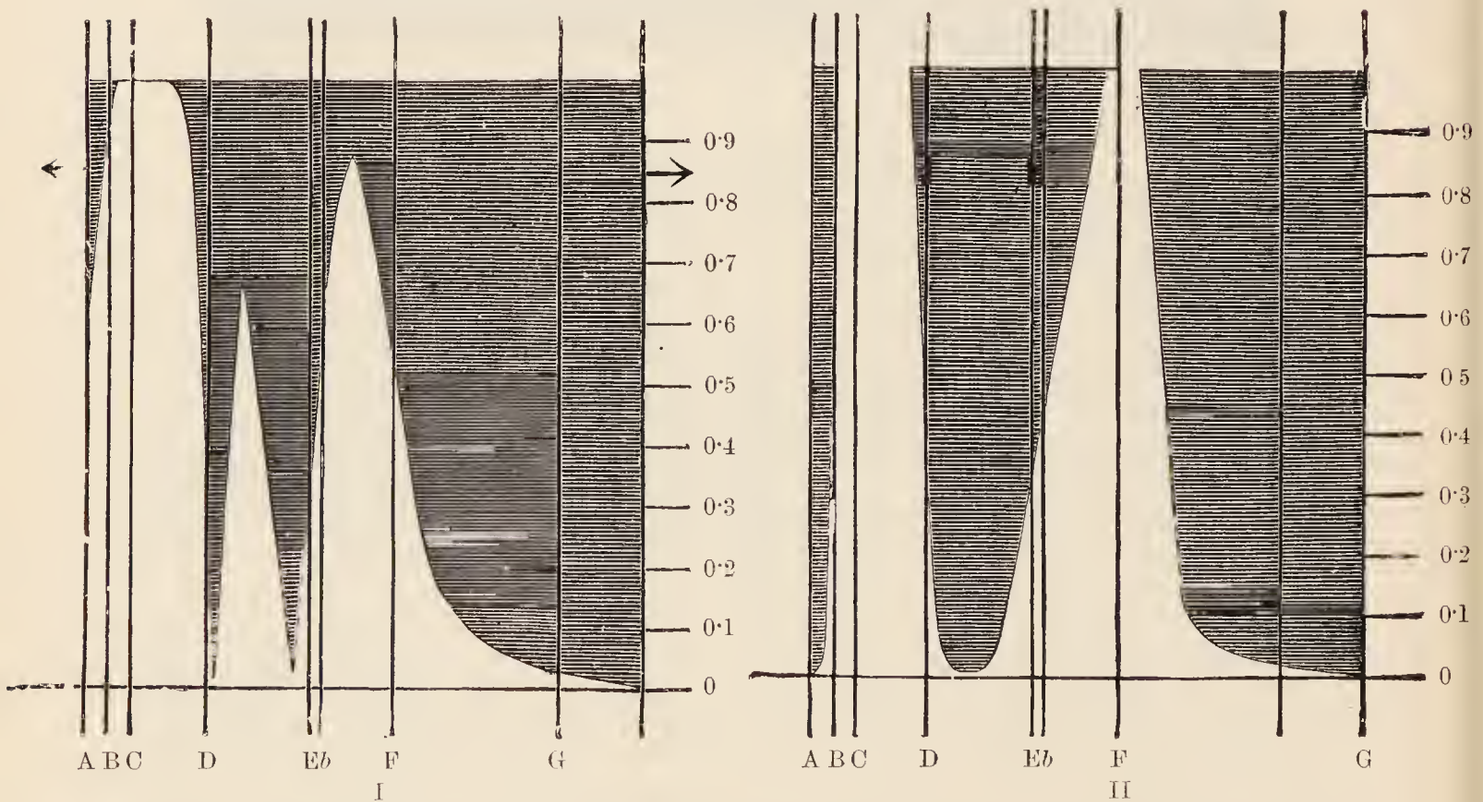


FIG. 36.—Graphic representations of the amount of absorption of light by solution (I) of oxyhæmoglobin, (II) of hæmoglobin, of different strengths. The shading indicates the amount of absorption of the spectrum; the figures on the right border express percentages. (Rollett.)

As will be seen on looking at fig. 36, a solution of oxyhæmoglobin of concentration greater than 0·65 per cent. and less than 0·85 per cent.

(examined in a cell of the usual thickness of 1 centimetre) gives one thick band overlapping both D and E, and a stronger solution only lets the red light through between C and D. A solution which gives the two characteristic bands must therefore be a very dilute one. The one band (γ band) of hæmoglobin (fig. 37, spectrum 3) is not so well defined as the α or β bands. On dilution it fades rapidly, so that in a solution of such strength that both bands of oxyhæmoglobin would be quite distinct the single band of hæmoglobin has disappeared from view. The oxyhæmoglobin bands can be distinguished in a solution which contains only one part of the pigment to 10,000

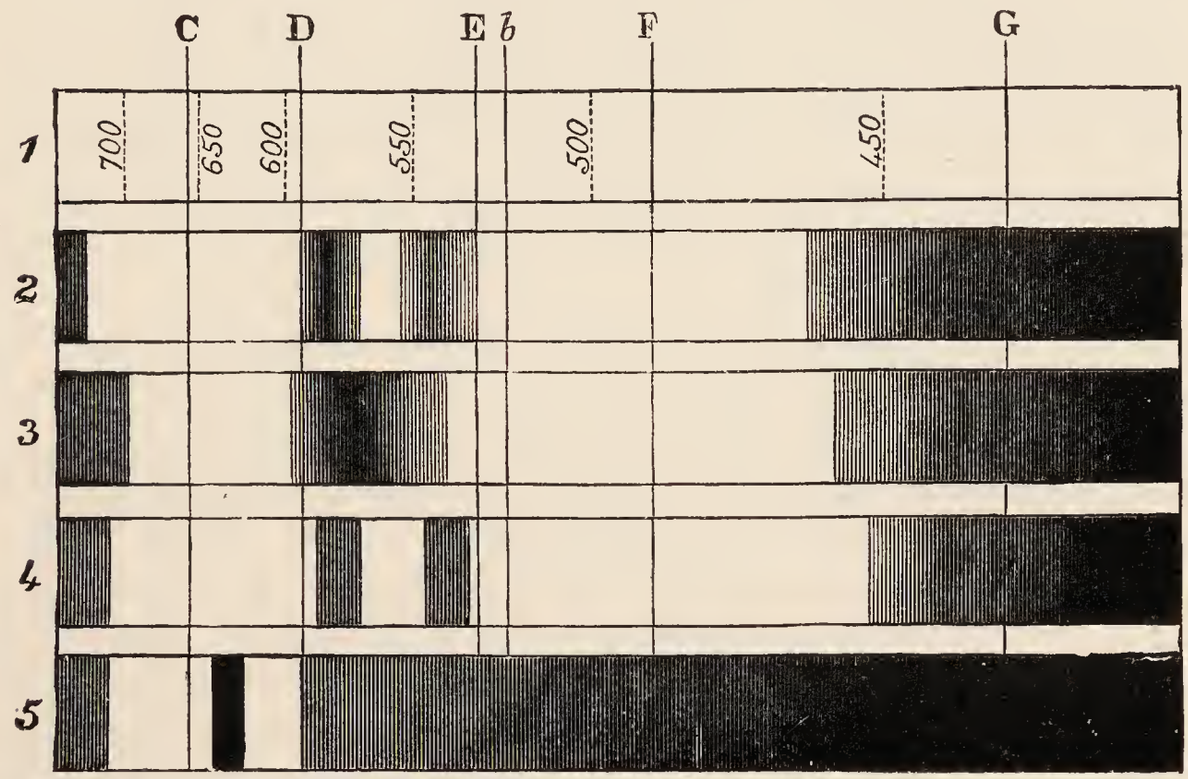


FIG. 37.—1, Solar spectrum; 2, spectrum of oxyhæmoglobin (0·37 per cent. solution); 3, spectrum of hæmoglobin; 4, spectrum of CO-hæmoglobin; 5, spectrum of methæmoglobin (concentrated solution).

of water, and even in more dilute solutions which seem to be colourless the α band is still visible.

Methæmoglobin.—This may be produced artificially by adding such reagents as potassium ferricyanide or amyl nitrite to a solution of oxyhæmoglobin; it may also occur in certain diseased conditions in the urine; it is therefore of considerable practical importance. It can be crystallised, and is found to contain the same amount of oxygen as oxyhæmoglobin, only combined differently. The oxygen is not removable by the air-pump, nor by a stream of a neutral gas like hydrogen. It can, however, by reducing agents like ammonium sulphide, be made to yield hæmoglobin. Methæmoglobin is of a brownish-red colour, and gives a characteristic absorption band in the red between the C and D lines (fig. 37, spectrum 5).

The ferricyanide of potassium or sodium not only causes the conversion of oxyhæmoglobin into methæmoglobin, but if the reagent is added to blood which has been previously laked by the addition of twice its volume of water there is an evolution of oxygen. If a small amount of sodium carbonate or ammonia is added as well to prevent the evolution of any carbonic acid, and the oxygen is collected and measured, it is found that all the oxygen previously combined in oxyhæmoglobin is discharged. This is at first sight puzzling, because, as just stated, methæmoglobin contains the same amount of oxygen that is present in oxyhæmoglobin. What occurs is that, after the oxygen is discharged from oxyhæmoglobin, an equal quantity of oxygen takes its place from the reagents added. The oxygen atoms of the methæmoglobin must be attached to a different part of the hæmatin group from the oxygen atoms of the oxyhæmoglobin, so that the hæmatin group when thus altered loses its power of combining with oxygen and carbonic oxide to form compounds which are dissociable in a vacuum.

Haldane, to whom we owe these interesting results, gives the following provisional equation to represent what occurs :—

$$\underset{\text{[oxyhæmoglobin]}}{HbO_2} + 4\underset{\text{[sodium ferricyanide]}}{Na_3Cy_6Fe} + 4\underset{\text{[sodium bicarbonate]}}{NaHCO_3} = \underset{\text{[methæmoglobin]}}{HbO_2} + 4\underset{\text{[sodium ferrocyanide]}}{Na_4Cy_6Fe}.$$

$$+ 4\underset{\text{[carbonic acid]}}{CO_2} + 2\underset{\text{[water]}}{H_2O} + \underset{\text{[oxygen]}}{O_2}.$$

Carbonic Oxide Hæmoglobin may be readily prepared by passing a stream of carbonic oxide or coal gas through blood or through a solution of oxyhæmoglobin. It has a peculiar cherry-red colour. Its absorption spectrum is very like that of oxyhæmoglobin, but the two bands are slightly nearer the violet end of the spectrum (fig. 37, spectrum 4). Reducing agents, like ammonium sulphide, do not change it; the gas is more firmly combined than the oxygen in oxyhæmoglobin. CO-hæmoglobin forms crystals like those of oxyhæmoglobin : it resists putrefaction for a very long time.

Carbonic oxide is given off during the imperfect combustion of carbon such as occurs in charcoal stoves : it is a powerful poison combining with the hæmoglobin of the blood, and thus interfering with normal respiratory processes. The colour of the blood and its resistance to reducing agents are in such cases characteristic.

Nitric Oxide Hæmoglobin.—When ammonia is added to blood, and then a stream of nitric oxide is passed through it, this compound is formed. It may be obtained in crystals isomorphous with oxy- and CO-hæmoglobin. It also has a similar spectrum. It is even more

stable than CO-hæmoglobin; it has little practical interest, but is of theoretical importance as completing the series.

Very dilute solutions of hæmoglobin and its derivatives show an absorption band in the ultra-violet region in addition to those just described in the visible regions of the spectrum (see more fully Advanced Course).

CHEMISTRY OF RESPIRATION

The consideration of the blood, and especially of its pigment, is so closely associated with respiration that a brief account of that process follows conveniently here.

The lungs consist essentially of numerous little hollow sacs in the walls of which is a close plexus of capillary blood vessels. These air sacs, or alveoli, communicate with the external air by the trachea, bronchi, and bronchial tubes. Inspiration is due to a muscular effort that enlarges the thorax, the closed cavity in which the lungs are situated. Owing to the atmospheric pressure the lungs become distended. Expiration is ordinarily brought about by the elastic rebound of the lungs and chest walls, and is only a muscular effort when forced; but even the most vigorous expiratory effort is unable to expel the alveolar air. This air and the blood in the capillaries are only separated by the thin capillary and alveolar walls. The blood parts with its excess of carbonic acid and watery vapour to the alveolar air; the blood at the same time receives from the alveolar air the oxygen which renders it arterial.

The intake of oxygen is the commencement, and the output of carbonic acid the end, of the series of changes known as respiration. The intermediate steps take place all over the body, and constitute what is known as *internal* or *tissue respiration*. The exchange of gases which occurs in the lungs is sometimes called in contradistinction *external respiration*. We have already seen that the oxyhæmoglobin is only a loose compound, and in the tissues it parts with its oxygen. The oxygen does not necessarily undergo immediate union with carbon to form carbonic acid, and with hydrogen to form water, but in most cases, as in muscle, is held in reserve by the tissue itself. Ultimately, however, these two oxides are formed: they are the chief products of combustion. Certain other products which represent the combustion of nitrogenous material (urea, uric acid, &c.) ultimately leave the body by the urine. All these substances pass into the venous blood, and the gaseous products, carbonic acid, and a portion of the water find an outlet by the lungs.

Inspired and Expired Air.—The composition of the inspired and expired air may be compared in the following table:—

—	Inspired or atmospheric Air	Expired Air
Oxygen . . .	20·96 vols. per cent.	16·03 vols. per cent.
Nitrogen . . .	79 ,, ,,	79 ,, ,,
Carbonic acid . .	0·04 ,, ,,	4·4 ,, ,,
Watery vapour . .	variable	saturated
Temperature . .	,,	that of body (37° C.)

The nitrogen remains unchanged. The recently discovered gases, argon, crypton, &c., are in the above table reckoned in with the nitrogen. They are, however, only present in minute quantities The chief change is in the proportion of oxygen and carbonic acid. The loss of oxygen is about 5, the gain in carbonic acid $4\frac{1}{2}$. If the inspired and expired airs are carefully measured at the same temperature and barometric pressure, the volume of expired air is thus rather less than that of the inspired. The conversion of oxygen into carbonic acid would not cause any change in the volume of the gas, for a molecule of oxygen (O_2) would give rise to a molecule of carbonic acid (CO_2) which would occupy the same volume (Avogadro's law). It must, however, be remembered that carbon is not the only element which is oxidised. Fats contain a number of atoms of hydrogen which during metabolism are oxidised to form water; a certain small amount of oxygen is also used in the formation of urea. Carbohydrates contain sufficient oxygen in their own molecules to oxidise their hydrogen; hence the apparent loss of oxygen is least when a vegetable diet (that is, one consisting largely of starch and other carbohydrates) is taken, and greatest when much fat and proteid are eaten. The quotient $\frac{CO_2 \text{ given off}}{O_2 \text{ absorbed}}$ is called the *respiratory quotient.* Normally it is $\frac{4·5}{5}=0·9$, but this varies considerably with diet, as just stated. It varies also with muscular exercise, when the output of carbonic acid is much increased both absolutely and relatively to the amount of oxygen used up.

Gases of the Blood.—From 100 volumes of blood about 60 volumes of gas can be removed by the mercurial air-pump (see Appendix). The average composition of this gas from dog's blood is—

—	Arterial Blood	Venous Blood
Oxygen . . .	20	8 to 12
Nitrogen . . .	1 to 2	1 to 2
Carbonic acid . .	40	46

The nitrogen in the blood is simply dissolved from the air just as water would dissolve it : it has no physiological importance. The other two gases are present in much greater amount than can be explained by simple solution ; they are, in fact, chiefly present in loose chemical combinations. Less than one volume of the oxygen and about two of carbonic acid are present in simple solution in the plasma.

Oxygen in the Blood.—The amount of gas dissolved in a liquid varies with the pressure of the gas ; double the pressure and the amount of gas dissolved is doubled. Now this does not occur in the case of oxygen and blood ; very nearly the same amount of oxygen is dissolved whatever be the pressure. We thus have a proof that oxygen is not merely dissolved in the blood, but is in chemical union ; and the fact that the oxygen of oxyhæmoglobin can be replaced by equivalent quantities of other gases, like carbonic oxide, is a further proof of the same statement. The tension or partial pressure of oxygen in the air of the alveoli is less than that in the atmosphere, but greater than that in venous blood ; hence oxygen passes from the alveolar air into the blood ; the oxygen immediately combines with the hæmoglobin, and thus leaves the plasma free to absorb more oxygen ; and this goes on until the hæmoglobin is entirely, or almost entirely, saturated with oxygen. The reverse change occurs in the tissues where the partial pressure of oxygen is lower than in the plasma, or the lymph that bathes the tissue elements ; the plasma parts with its oxygen to the lymph, the lymph to the tissues ; the oxyhæmoglobin then undergoes dissociation to supply more oxygen to the plasma and lymph, and this in turn to the tissues once more. This goes on until the oxyhæmoglobin loses a great portion of its store of oxygen, but even in asphyxia it does not lose all.

The following values are given by Fredericq for the tension of oxygen in percentages of an atmosphere. His experiments were made on dogs :—

External air	20·96
Alveolar air	18
Arterial blood	14
Tissues	0

The arrow shows the direction in which the gas passes.

The methods of obtaining the gases of the blood and analysing them are described in the Appendix. When the gases are being pumped off from the blood, very little oxygen comes off until the pressure is greatly reduced, and then, at a certain point, it is suddenly disengaged. This shows it is not in simple solution, but is united

chemically to the hæmoglobin as oxyhæmoglobin, which is dissociated when the pressure is extremely low.

The avidity of the tissues for oxygen is shown by Ehrlich's experiments with methylene blue and similar pigments. Methylene blue is more stable than oxyhæmoglobin ; but if it is injected into the circulation of a living animal, and the animal killed a few minutes later, the blood is found dark blue, but the organs colourless. On exposure to oxygen the organs become blue. In other words the tissues have removed the oxygen from methylene blue to form a colourless reduction product ; on exposure to the air this once more unites with oxygen to form methylene blue.

Carbonic Acid in the Blood.—What has been said for oxygen holds good in the reverse direction for carbonic acid. Compounds are formed in the tissues where the tension of the gas is high : these pass into the lymph, then into the blood, and in the lungs the compounds undergo dissociation, carbonic acid passing into the alveolar air where the tension of the gas is comparatively low, though it is greater here than in the expired air.

The relations of this gas and the compounds it forms are more complex than in the case of oxygen. If blood is divided into plasma and corpuscles, it will be found that both yield carbonic acid, but the yield from the plasma is the greater. If we place blood in a vacuum it bubbles, and gives out all its gases ; addition of a weak acid causes no further liberation of carbonic acid. If plasma or serum is similarly treated the gas comes off, but from 10 to 18 per cent. of the carbonic acid is fixed—that is, the addition of some stronger acid, like phosphoric acid, is necessary to displace it. Fresh red corpuscles will, however, take the place of the phosphoric acid, and thus it has been surmised that oxyhæmoglobin has the properties of an acid.

One hundred volumes of venous blood contain forty-six volumes of carbonic acid. Whether this is in solution or in chemical combination is determined by ascertaining the tension of the gas in the blood. One hundred volumes of blood plasma would dissolve more than an equal volume of the gas at atmospheric pressure, if its solubility in plasma were equal to that in water.[1] If, then, the carbonic acid were in a state of solution, its tension would be very high, but it proves to be only equal to 5 per cent. of an atmosphere. This means that when venous blood is brought into an atmosphere containing 5 per cent. of carbonic acid, the blood neither gives off

[1] To be exact, the solubility of carbon dioxide in plasma is a little less than in pure water.

any carbonic acid nor takes up any. Hence the remainder of the gas, 95 per cent., is in a condition of chemical combination. The chief compound appears to be sodium bicarbonate.

The carbonic acid and phosphoric acid of the blood are in a state of constant struggle for the possession of the sodium. The salts formed by these two acids depend on their relative masses. If carbonic acid is in excess, we get sodium carbonate (Na_2CO_3), and mono-sodium phosphate (NaH_2PO_4); but if the carbonic acid is diminished, the phosphoric acid obtains the greater share of sodium to form disodium phosphate (Na_2HPO_4). In this way, as soon as the amount of free carbonic acid diminishes, as in the lungs, the amount of carbonic acid in combination also decreases; whereas in the tissues, where the tension of the gas is highest, a large amount is taken up into the blood, where it forms sodium bicarbonate.

The tension of the carbonic acid in the tissues is high, but one cannot give exact figures; we can measure the tension of the gas in certain secretions; in the urine it is 9, in the bile 7 per cent. The tension in the cells themselves must be higher still.

The following figures (from Fredericq) give the tension of carbonic dioxide in percentages of an atmosphere :—

Tissues	5 to 9	
Venous blood	3·8 to 5·4	in dog ⇊
Alveolar air	2·8	
External air	0·04	

The arrow indicates the direction in which the gas passes, namely in the direction of pressure from the tissues to the atmosphere.

In some other experiments, also on dogs, the following are the figures given :—

Arterial blood	2·8	
Venous blood	5·4	⇊
Alveolar air	3·56	
Expired air	2·8	

It will be seen from these figures that the tension of carbonic acid in the venous blood (5·4) is higher than in the alveolar air (3·56); its passage into the alveolar air is therefore intelligible by the laws of diffusion. Diffusion, however, should cease when the tension of the gas in the blood and alveolar air are equal. But the transference goes beyond the establishment of such an equilibrium, for the tension of the gas in the blood continues to sink until it is, when the blood is arterial, ultimately less (2·8) than in the alveolar air.

The whole question is beset with great difficulties and contradictions. Analyses by different observers have given very different

results, but if such figures as those just quoted are ultimately found to be correct, we can only explain this apparent reversal of a law of nature by supposing with Bohr that the alveolar epithelium possesses the power of excreting carbonic acid, just as the cells of secreting glands are able to select certain materials from the blood and reject others. Recent work by Bohr and Haldane has also shown that in all probability the same explanation—epithelial activity—must be called in to account for the absorption of oxygen. Haldane, in fact, states that the tension of oxygen in the blood is greater than in the atmosphere. In the swim-bladder of fishes (which is analogous to the lung of mammals) the oxygen is certainly far in excess of anything that can be explained by mere diffusion. The storage of oxygen, moreover, ceases when the vagus nerves which supply the swim-bladder are divided.

Some Continental observers have stated that certain noxious substances are ordinarily contained in expired air which are much more poisonous than carbonic acid, but researches in this country have entirely failed to substantiate this. If precautions be taken by absolute cleanliness to prevent admixture of the air with exhalations from skin, teeth, and clothes, the expired air only contains one noxious substance, and that is carbonic acid.

Tissue-Respiration.—Before the processes of respiration were fully understood the lungs were looked upon as the seat of combustion ; they were regarded as the stove for the rest of the body where effete material was brought by the venous blood to be burnt up. When it was shown that the venous blood going to the lungs already contained carbonic acid, and that the temperature of the lungs is not greater than that of the rest of the body, this explanation had of necessity to be dropped.

Physiologists next transferred the seat of the combustion to the blood ; but since then innumerable facts and experiments have shown that it is in the tissues themselves, and not in the blood, that combustion occurs. The methylene-blue experiments already described (p. 112) show this ; and the following experiment is also quite conclusive. A frog can be kept alive for some time after salt solution is substituted for its blood. The metabolism goes on actively if the animal is kept in pure oxygen. The taking up of oxygen and giving out of carbonic acid must therefore occur in the tissues, as the animal has no blood.

The Aërotonometer.—This instrument was invented by Pflüger, and of the modifications that have been introduced since then, that of Fredericq is the simplest. It merely consists of a long glass tube surrounded by a water-jacket at body temperature. The tube is filled with a known mixture of gases, and the blood from the carotid artery is allowed to slowly trickle down the tube; the blood then returns to the jugular vein. In order that the

experiment may be continued for an hour or more, the animal's blood must be rendered incoagulable; this may be done by a previous injection of 'peptone.' This has the disadvantage of lowering the carbonic acid tension of the blood. Another way of rendering the blood incoagulable is to collect it, defibrinate it, and then return it to the circulation. The last step in the experiment consists in examining the composition of the gases left in the tube. The principle upon which the instrument acts is this:—If blood is brought into contact with a mixture of oxygen, nitrogen, and carbonic acid gas, it gives off some of its gases if their partial pressure is greater in the blood than the respective tensions of the gases in the mixture; if, on the other hand, the tension is lower in the blood than in the mixture for any gas or gases, they pass from the mixture to the blood. These interchanges go on until equilibrium is established. For example, suppose the aërotonometer contains at the start 10 per cent. of oxygen, 5 of carbonic acid, and 85 of nitrogen. Blood is then passed through it for an hour, and the gases are again analysed, and the mixture then contains 14 per cent. of oxygen, 2·8 of carbon dioxide, and the rest nitrogen; the tension of oxygen and carbon dioxide in the blood will then respectively be 14 and 2·8 per cent. of an atmosphere.

The Carbonic Oxide Method of Estimating the Oxygen Tension of Arterial Blood.—This method was devised by Haldane, and is considered by him and Lorrain Smith to give more trustworthy results than those obtained by the aërotonometer. If blood is exposed to a mixture of carbonic oxide and oxygen, the hæmoglobin will become saturated by these gases according to their relative tensions. If a number of experiments are performed using different percentages of carbonic oxide, the results may be expressed graphically as the curve of dissociation of carboxyhæmoglobin in air. When, in place of such experiments *in vitro*, an animal is made to breathe air containing a known percentage of carbonic oxide, the comparison of the saturation of its blood with the saturation of its blood *in vitro* exposed to the same percentage of carbonic oxide in air (which has an oxygen tension of 20·9 per cent.) gives us the means of discovering the oxygen tension in the arterial blood of the lung capillaries; this will be higher or lower than that of the air according as the saturation by carbonic oxide is correspondingly lower or higher. A small animal like a mouse is made to breathe air containing a known percentage of carbonic oxide. After a sufficient time the animal is killed and the amount of carboxyhæmoglobin is determined colorimetrically in a drop of its blood. The data thus obtained are compared with the data previously expressed in the curve of dissociation of carboxyhæmoglobin in air; it is then easy to calculate whether the oxygen tension in the blood is higher or lower than that of air. The results of the method show generally that the tension of oxygen in the arterial blood as it leaves the lungs is higher than could result from simple diffusion of the oxygen through the alveolar epithelium; in other words, the epithelial cells are capable of secreting oxygen into the blood until an oxygen pressure is reached considerably above that in the alveolar air.

The results expressed in percentages of an atmosphere are as follows:—

Oxygen tension of arterial blood in man, 38·5; in mouse, 22·6; in dog, 21; in cat, 35·3; in rabbit, 27·6, and in birds 30 to 50 per cent. The results in the case of man and larger animals probably require revision, as it is not certain that the time allowed for the establishment of the balance of carbonic oxide and oxygen has been sufficient in any of the experiments.

TESTS FOR BLOOD

These may be gathered from preceding descriptions. Briefly, they are microscopic, spectroscopic, and chemical. The best chemical test is the formation of hæmin crystals. The old test with tincture of guaiacum and hydrogen peroxide, the blood causing the red tincture to become green, is very untrustworthy, as it is also given by many other organic substances.

In medico-legal cases it is often necessary to ascertain whether a red fluid or stain upon clothing is or is not blood. In any such case it is advisable not to rely upon one test only, but to try every means of detection at one's disposal. To discover whether it is blood or not is by no means a difficult problem, but to distinguish human blood from that of the common mammals is possible only by the 'biological' test described at the end of the next section.

IMMUNITY

The chemical defences of the body against injury and disease are numerous. The property that the blood possesses of coagulating is a defence against hæmorrhage; the acid of the gastric juice is a protection against harmful bacteria introduced with food. Bacterial activity in urine is inhibited by the acidity of that secretion.

Far more important and widespread in its effects than any of the foregoing is the bactericidal (*i.e.* bacteria-killing) action of the blood and lymph; a study of this question has led to many interesting results, especially in connection with the important problem of immunity.

It is a familiar fact that one attack of many infective maladies protects us against another attack of the same disease. The person is said to be *immune*, either partially or completely, against that disease. Vaccination produces in a patient an attack of cowpox or vaccinia. This disease is either closely related to smallpox, or maybe it is smallpox modified and rendered less malignant by passing through the body of a calf. At any rate, an attack of vaccinia renders a person immune to smallpox, for a certain number of years. Vaccination is an instance of what is called *protective inoculation*, which is now practised with more or less success in reference to other diseases, like plague and typhoid fever. The study of immunity has also rendered possible what may be called *curative inoculation*, or the injection of antitoxic material as a cure for diphtheria, tetanus, snake poisoning, &c.

The power the blood possesses of slaying bacteria was first discovered when the effort was made to grow various kinds of bacteria in it; it was looked upon as probable that blood would prove a suitable soil or medium for this purpose. It was found in some instances to have exactly the opposite effect. The chemical characters of the substances which kill the bacteria are not fully known; but they appear to be proteid in nature. The bactericidal powers of blood are destroyed by heating it for an hour to 55° C. The balance of evidence appears to me to be in favour of the view that the substances in question originate from the leucocytes, and phagocytosis becomes more intelligible if this is accepted. The substances, whatever be their source or their chemical nature, are called *bacterio-lysins.*

Closely allied to the bactericidal power of blood, or blood-serum, is its globulicidal power. By this one means that the blood-serum of one animal has the power of dissolving the red blood-corpuscles of another species. If the serum of one animal is injected into the blood-stream of an animal of another species, the result is a destruction of its red corpuscles, which may be so excessive as to lead to the passing of the liberated hæmoglobin into the urine (hæmoglobinuria). The substance or substances in the serum that possess this property are called *hæmolysins,* and though there is some doubt whether bacterio-lysins and hæmolysins are absolutely identical, there is no doubt that they are closely related substances.

Another interesting chemical point in this connection is the fact that the bactericidal power of the blood is closely related to its alkalinity. Increase of alkalinity means increase of bactericidal power. Venous blood contains more diffusible alkali than arterial blood, and is more bactericidal; dropsical effusions are more alkaline than normal lymph and kill bacteria more easily. In a condition like diabetes, when the blood is less alkaline than it should be, the susceptibility to infectious diseases is increased. Alkalinity is probably beneficial because it favours those oxidative processes in the cells of the body which are so essential for the maintenance of healthy life.

Normal blood possesses a certain amount of substances which are inimical to the life of our bacterial foes. But suppose a person gets run down; everyone knows he is then more liable to 'catch anything.' This coincides with a diminution in the bactericidal power of his blood. But even a perfectly healthy person has not an unlimited supply of bacterio-lysin, and if the bacteria are sufficiently numerous he will fall a victim to the disease they produce. Here, however, comes in the remarkable part of the defence. In the struggle he

will produce more and more bacterio-lysin, and if he gets well it means that the bacteria are finally vanquished, and his blood remains rich in the particular bacterio-lysin he has produced, and so will render him immune to further attacks from that particular species of bacterium. Every bacterium seems to cause the development of a specific bacterio-lysin.

Immunity can more conveniently be produced gradually in animals, and this applies, not only to the bacteria, but also to the toxins they form. If, for instance, the bacilli which produce diphtheria are grown in a suitable medium, they produce the diphtheria poison, or toxin, much in the same way that yeast-cells will produce alcohol when grown in a solution of sugar. Diphtheria toxin is associated with a proteose, as is also the case with the poison of snake venom. If a certain small dose called a 'lethal dose' is injected into a guinea-pig the result is death. But if the guinea-pig receives a smaller dose it will recover; a few days after it will stand a rather larger dose; and this may be continued until, after many successive gradually increasing doses, it will finally stand an amount equal to many lethal doses without any ill effects. The gradual introduction of the toxin has called forth the production of an antitoxin. If this is done in the horse instead of the guinea-pig the production of antitoxin is still more marked, and the serum obtained from the blood of an immunised horse may be used for injecting into human beings suffering from diphtheria, and rapidly cures the disease. The two actions of the blood, antitoxic and antibacterial, are frequently associated, but may be entirely distinct.

The antitoxin is also a proteid probably of the nature of a globulin; at any rate it is a proteid of larger molecular weight than a proteose. This suggests a practical point. In the case of snake-poisoning the poison gets into the blood rapidly owing to the comparative ease with which it diffuses, and so it is quickly carried all over the body. In treatment with the antitoxin or antivenin, speed is everything if life is to be saved; injection of this material under the skin is not much good, for the diffusion into the blood is too slow. It should be injected straight away into a blood-vessel.

There is no doubt that in these cases the antitoxin neutralises the toxin much in the same way that an acid neutralises an alkali. If the toxin and antitoxin are mixed in a test-tube, and time allowed for the interaction to occur, the result is an innocuous mixture. The toxin, however, is merely neutralised, not destroyed; for if the mixture in the test-tube is heated to 68° C. the antitoxin is coagulated and destroyed and the toxin remains as poisonous as ever.

Immunity is distinguished into *active* and *passive*. Active immunity is produced by the development of protective substances in the body; passive immunity by the injection of a protective serum. Of the two the former is the more permanent.

Ricin, the poisonous proteid of castor-oil seeds, and *abrin*, that of the Jequirity bean, also produce when gradually given to animals an immunity, due to the production of antiricin and antiabrin respectively.

Ehrlich's hypothesis to explain such facts is usually spoken of as the *side-chain theory* of immunity. He considers that the toxins are capable of uniting with the protoplasm of the living cells by possessing groups of atoms like those by which nutritive proteids are united to cells during normal assimilation. He terms these *haptophor* groups, and the groups to which these are attached in the cells he terms *receptor* groups. The introduction of a toxin stimulates an excessive production of receptors, which are finally thrown out into the circulation, and the free circulating receptors constitute the antitoxin. The comparison of the process to assimilation is justified by the fact that non-toxic substances like milk or egg-white introduced gradually by successive doses into the blood-stream cause the formation of anti-substances capable of coagulating them.

Up to this point I have spoken only of the blood, but month by month workers are bringing forward evidence to show that other cells of the body may by similar measures be rendered capable of producing a corresponding protective mechanism.

One further development of the theory I must mention. At least two different substances are necessary to render a serum bactericidal or globulicidal. The bacterio-lysin or hæmolysin consists of these two substances. One of these is called the *immune body*, the other the *complement*. We may illustrate the use of these terms by an example. The repeated injection of the blood of one animal (*e.g.* the goat) into the blood of another animal (*e.g.* a sheep) after a time renders the latter animal immune to further injections, and at the same time causes the production of a serum which dissolves readily the red blood-corpuscles of the first animal. The sheep's serum is thus hæmolytic towards goat's blood-corpuscles. This power is destroyed by heating to 56° C. for half an hour, but returns when fresh serum of any animal is added. The specific immunising substance formed in the sheep is called the immune body; the ferment-like substance destroyed by heat is the complement. The latter is not specific, since it is furnished by the blood of non-immunised animals, but it is nevertheless essential for hæmolysis. Ehrlich believes that the immune body has two side groups—one which connects with the receptor of the red corpuscles, and one which unites with the haptophor group of the complement, and thus renders possible the ferment-like action of the complement on the red corpuscles. Various antibacterial serums, which have not been the success in treating disease they were expected to be, are probably too poor in complement, though they may contain plenty of the immune body.

To put it another way: the cell-dissolving substances cannot act on their objects of attack without an intermediate substance to anchor them on the substance in question. This intermediary substance, known as the immune body or amboceptor, is specific, and varies with the substance to be attacked (red corpuscles, bacterium, toxin, &c.). The complement may be compared to a person who wants to unlock a door; to do this effectively he must be provided with the proper key (amboceptor or immune body).

Quite distinct from the bactericidal, globulicidal, and antitoxic properties of blood is its agglutinating action. This is another result of infection with many kinds of bacteria or their toxins. The blood

acquires the property of rendering immobile and clumping together the specific bacteria used in the infection. The test applied to the blood in cases of typhoid fever, and generally called Widal's reaction, depends on this fact.

The substances that produce this effect are called *agglutinins*. They also are probably proteid-like in nature, but are more resistant to heat than the lysins. Prolonged heating to over 60° C. is necessary to destroy their activity.

Lastly, we come to a question which more directly appeals to the physiologist than the preceding, because experiments in relation to immunity have furnished us with what has hitherto been lacking, a means of distinguishing human blood from the blood of other animals.

The discovery was made by Tchistovitch (1899), and his original experiment was as follows:—Rabbits, dogs, goats, and guinea-pigs were inoculated with eel-serum, which is toxic; he thereby obtained from these animals an antitoxic serum. But the serum was not only antitoxic; it also produced a precipitate when added to eel-serum, though not when added to the serum of any other animal. In other words, not only has a specific antitoxin been produced, but also a specific *precipitin*. Numerous observers have since found that this is a general rule throughout the animal kingdom, including man. If, for instance, a rabbit is treated with human blood, the serum ultimately obtained from a rabbit contains a specific precipitin for human blood; that is to say, a precipitate is formed on adding such a rabbit's serum to human blood, but not when added to the blood of any other animal.[1] The great value of the test is its delicacy; it will detect the specific blood when it is greatly diluted, after it has been dried for weeks, or even when it is mixed with the blood of other animals.

[1] There may be a slight reaction with the blood of allied animals; for instance, with monkey's blood in the case of man.

LESSON X

URINE

1. Test the reaction of urine on litmus paper.
2. Determine its specific gravity by the urinometer.
3. Test for the following **inorganic salts** :

(*a*) *Chlorides.*—Acidulate with nitric acid and add silver nitrate; a white precipitate of silver chloride, soluble in ammonia, is produced. The object of acidulating with nitric acid is to prevent phosphates being precipitated by the nitrate of silver.

(*b*) *Sulphates.*—Acidulate with hydrochloric acid and add barium chloride. A white precipitate of barium sulphate is produced. Hydrochloric acid is again added first, to prevent precipitation of phosphates.

(*c*) *Phosphates.*—i. Add ammonia; a white crystalline precipitate of *earthy* (that is, calcium and magnesium) phosphates is produced. This becomes more apparent on standing. The *alkaline* (that is, sodium and potassium) phosphates remain in solution.

ii. Mix another portion of urine with half its volume of nitric acid; add ammonium molybdate, and boil. A yellow crystalline precipitate falls. This test is given by both kinds of phosphates.

4. **Urea.**—Take some urea crystals. Observe that they are readily soluble in water, and that effervescence occurs when fuming nitric acid (*i.e.* nitric acid containing nitrous acid in solution) is added to the solution. The effervescence is due to the breaking up of the urea. Carbonic acid and nitrogen come off. A similar bubbling, due to evolution of nitrogen, occurs when an alkaline solution of sodium hypobromite is added to another portion of the solution.

5. Heat some urea crystals in a dry test-tube. Biuret is formed, and ammonia comes off. Add a drop of copper-sulphate solution and a few drops of 20-per-cent. potash. A rose-red colour is produced.

6. Quantitative estimation of urea.

For this purpose Dupré's apparatus (fig. 38) is the most convenient. It consists of a bottle united to a measuring tube by indiarubber tubing. The measuring tube (an inverted burette will do very well) is placed within a cylinder of water, and can be raised and lowered at will. Measure 25 c.c. of alkaline solution of sodium hypobromite (made by mixing 2 c.c. of bromine with 23 c.c. of a 40-per-cent. solution of caustic soda) into the bottle. Measure 5 c.c. of urine into a small tube, and lower it carefully, so that no urine spills, into the bottle. Close the bottle securely with a stopper perforated by a glass tube ; this glass tube [1] is connected to the measuring tube by indiarubber tubing and a T-piece. The third limb of the T-piece is closed by a piece of indiarubber tubing and a pinch-cock, seen at the top of the

[1] The efficiency of the apparatus is increased by having a glass bulb blown on this tube to prevent froth passing into the rest of the apparatus. This is not shown in the figure.

figure. Open the pinch-cock and lower the measuring tube until the surface of the water with which the outer cylinder is filled is at the zero point of the graduation. Close the pinch-cock, and raise the measuring tube to ascertain whether the apparatus is air-tight. Then lower it again. Tilt the bottle so as to upset the urine, and shake well for a minute or so. During this time there is an evolution of gas. Then immerse the bottle in a large beaker containing water of the same temperature as that in the cylinder. After two or three minutes raise the measuring tube until the surfaces of the water inside and outside it are at the same level. Read off the amount of gas evolved. This is nitrogen. The carbonic acid resulting from the decomposition of urea has been absorbed by the excess of soda in the bottle. 35·4 c.c. of nitrogen are yielded by 0·1 gramme of urea. From this the quantity of urea in the 5 c.c. of urine and the percentage of urea can be calculated. If the total urea passed in the twenty-four hours is to be ascertained, the twenty-four hours' urine must be carefully measured and thoroughly mixed. A sample is then taken from the total for analysis; and then, by a simple sum in proportion, the total amount of urea is ascertained.

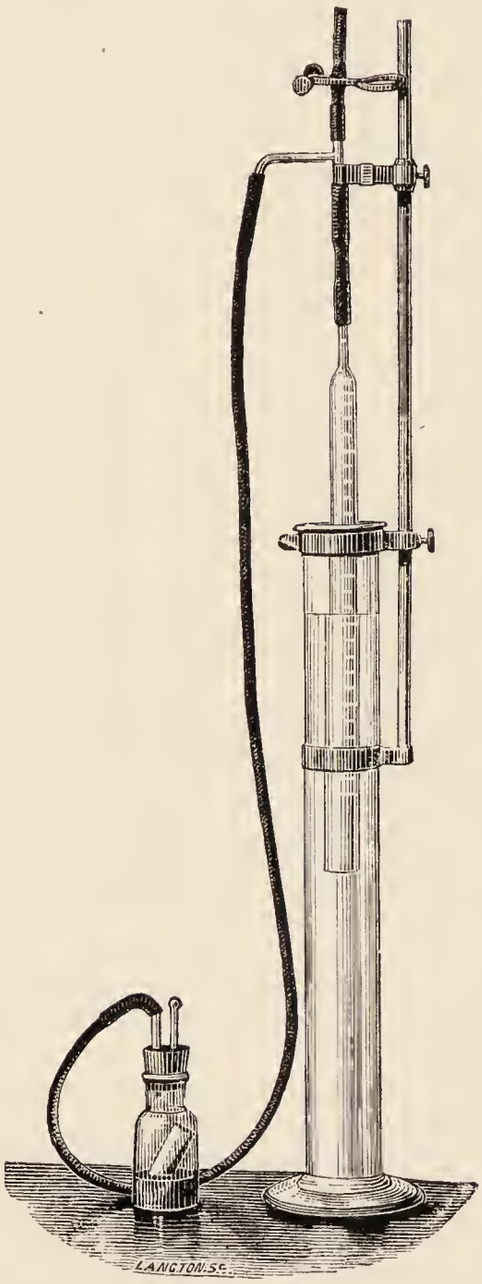


FIG. 38.—Dupré's urea apparatus.

7. **Creatinine.**—This substance may be detected by adding a little sodium nitro-prusside and caustic soda to the urine. A red colour develops, which fades on boiling.

The kidney is a compound tubular gland, the tubules of which it is composed differing much in the character of the epithelium that lines them in various parts of their course. The true secreting part of the kidney is the glandular epithelium that lines the convoluted portion of the tubules; there is in addition to this what is usually termed the filtering apparatus: tufts of capillary blood-vessels called the Malpighian glomeruli are supplied with afferent vessels from the renal artery; the efferent vessels that leave these have a smaller calibre, and thus there is high pressure in the Malpighian capillaries. Certain constituents of the blood, especially water and salts, pass through the thin walls of these vessels into the surrounding Bowman's capsule which forms the commencement of each renal tubule.

Bowman's capsule is lined by a flattened epithelium, which is reflected over the capillary tuft. Though the process which occurs here is generally spoken of as a filtration, yet it is no purely mechanical process, but the cells exercise a selective influence, and prevent the albuminous constituents of the blood from escaping. During the passage of this dilute urine through the rest of the renal tubule, it gains the constituents, urea, urates, &c., which are poured into it by the secreting cells of the convoluted tubules.

The term *excretion* is better than *secretion* as applied to the kidney, for the constituents of the urine are not actually formed in the kidney itself (as, for instance, the bile is formed in the liver), but they are formed elsewhere; the kidney is simply the place where they are picked out from the blood and eliminated from the body.

GENERAL CHARACTERS OF URINE

Quantity.—A man of average weight and height passes from 1,400 to 1,600 c.c., or about 50 oz., daily. This contains about 50 grammes (1½ oz.) of solids. The urine should be collected in a tall graduated glass vessel capable of holding 3,000 c.c., which should have a smooth-edged neck accurately covered by a ground-glass plate to exclude dust and avoid evaporation. From the total quantity thus collected in the twenty-four hours, samples should be drawn off for examination.

Colour.—This is some shade of yellow which varies considerably in health with the concentration of the urine. It appears to be due to a mixture of pigments: of these *urobilin* is the one of which we have the most accurate knowledge. Urobilin has a reddish tint and is ultimately derived from the blood pigment, and, like bile pigment, is an iron-free derivative of hæmoglobin. The bile pigment (and possibly also the hæmatin of the food) is in the intestines converted into stercobilin; most of the stercobilin leaves the body with the fæces; but some is reabsorbed and is excreted with the urine as urobilin. Urobilin is very like the artificial reduction product of bilirubin called hydrobilirubin (see p. 83). Normal urine, however, contains very little urobilin. The actual body present is a chromogen or mother substance called urobilinogen, which by oxidation (such as occurs when the urine stands exposed to the air) is converted into the pigment proper. In certain diseased conditions the amount of urobilin is considerably increased.

The most abundant urinary pigment is a yellow one called *urochrome*. It shows no absorption bands. It is probably an oxidation product of urobilin (Riva, A. E. Garrod). (See Lesson XXVI.)

Reaction.—The reaction of normal urine is acid. This is not due to free acid, as the uric and hippuric acids in the urine are combined as urates and hippurates respectively. The acidity is due to acid salts, especially acid sodium phosphate. Under certain circumstances the urine becomes less acid and even alkaline; the most important of these are as follows:—

1. During digestion. Here, there is a formation of free acid in the stomach, and a corresponding liberation of bases in the blood which passing into the urine diminish its acidity, or even render it alkaline. This is called *the alkaline tide*; the opposite condition, *the acid tide*, occurs after a fast—for instance, before breakfast.

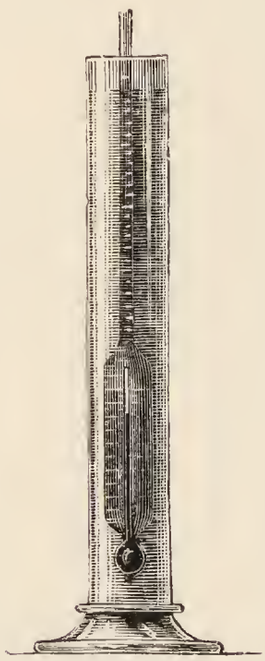


FIG. 39.—Urinometer floating in urine in a testing glass.



FIG. 40.—Crystals of urea: *a*, four-sided prisms; *b*, indefinite crystals, such as are usually formed from alcohol solutions.

2. In herbivorous animals and vegetarians. The food here contains excess of alkaline salts of acids like tartaric, citric, malic, &c. These acids are oxidised into carbonates, which passing into the urine give it an alkaline reaction.

Specific Gravity.—This should be taken in a sample of the twenty-four hours' urine with a good urinometer (see fig. 39).

The specific gravity varies inversely as the quantity of urine passed under normal conditions from 1015 to 1025. A specific gravity below 1010 should excite suspicion of hydruria; one over 1030 of a febrile condition, or diabetes, a disease in which it may rise to 1050. The specific gravity has, however, been known to sink as low as 1002 (after large potations, *urina potus*), or to rise as high as 1035 (after great sweating) in perfectly healthy persons.

Composition.—The following table gives the average amounts of

the urinary constituents passed by a man in the twenty-four hours :—

Water	1500·00 grammes
Total solids	72·00 ,,
Urea	33·18 ,,
Uric acid	0·55 ,,
Hippuric acid	0·40 ,,
Creatinine	0·91 ,,
Pigment and other organic substances	10·00 ,,
Sulphuric acid	2·01 ,,
Phosphoric acid	3·16 ,,
Chlorine	7·50 ,,
Ammonia	0·77 ,,
Potassium	2·50 ,,
Sodium	11·09 ,,
Calcium	0·26 ,,
Magnesium	0·21 ,,

The most abundant constituents of the urine are water, urea, and sodium chloride. In the foregoing table the student must not be misled by seeing the names of the acids and metals separated. The acids and the bases are combined to form salts :—urates, chlorides, sulphates, phosphates, &c.

UREA

Urea or **Carbamide**, $CO(NH_2)_2$, is isomeric (that is, has the same empirical, but not the same structural formula) with ammonium cyanate $(NH_4)CNO$, from which it was first prepared synthetically by Wöhler in 1828. Since then it has been prepared synthetically in other ways. Wöhler's observation derives interest from the fact that this was the first organic substance which was prepared synthetically by chemists.

When separated from the urine, it is found to be readily soluble both in water and in alcohol : it has a saltish taste, and is neutral to litmus paper. The form of its crystals is shown in fig. 40.

When treated with nitric acid, nitrate of urea ($CON_2H_4.HNO_3$) is formed ; this crystallises in octahedra, lozenge-shaped tablets, or hexagons (fig. 41, *a*). When treated with oxalic acid, prismatic crystals of urea oxalate ($CON_2H_4.H_2C_2O_4 + H_2O$) are formed (fig. 41, *b*).

These crystals may be readily obtained in an impure form by adding excess of the respective acids to urine which has been concentrated to a third or a quarter of its bulk.[1]

Under the influence of an organised ferment, the torula or micro-

[1] The preparation of urea nitrate and urea oxalate is postponed to the next lesson, when other microscopic crystals will also be under examination.

coccus ureæ, which grows readily in stale urine, urea takes up water, and is converted into ammonium carbonate [$CON_2H_4+2H_2O=(NH_4)_2CO_3$]. Hence the ammoniacal odour of putrid urine.

By means of nitrous acid, urea is broken up into carbonic acid, water, and nitrogen, $CON_2H_4+2HNO_2=CO_2+3H_2O+2N_2$. This may be used as a test for urea. Add fuming nitric acid (*i.e.* nitric acid containing nitrous acid in solution) to a solution of urea, or to urine; an abundant evolution of gas bubbles takes place.

Hypobromite of soda decomposes urea in the following way:—

$$\underset{\text{[urea]}}{CON_2H_4} + \underset{\text{[sodium hypobromite]}}{3NaBrO} = \underset{\text{[carbonic acid]}}{CO_2} + \underset{\text{[nitrogen]}}{N_2} + \underset{\text{[water]}}{2H_2O} + \underset{\text{[sodium bromide]}}{3NaBr}$$

This reaction is important, for on it one of the readiest methods for estimating urea depends. There have been various pieces of appa-

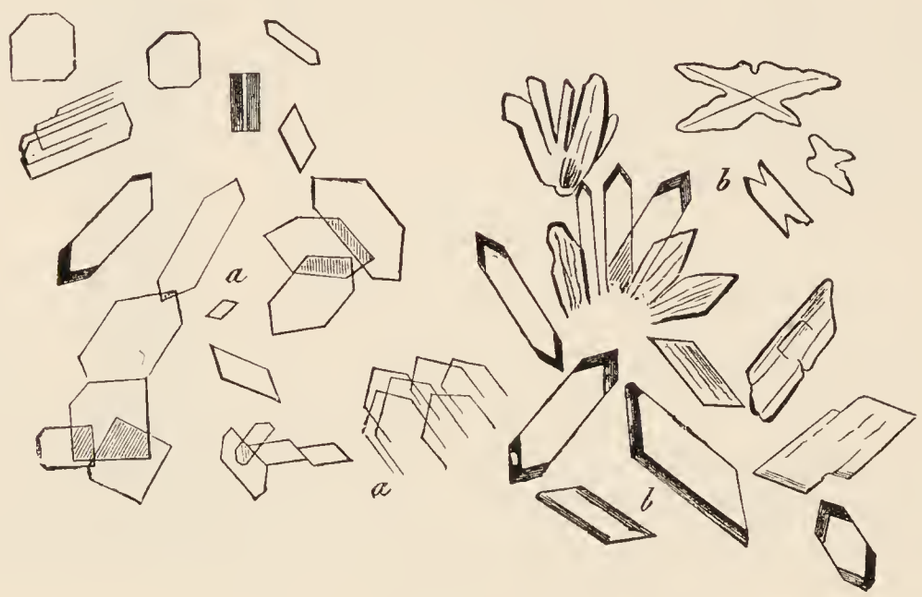


FIG. 41.—*a*, nitrate; *b*, oxalate of urea.

ratus invented for rendering the analysis easy; but the one described in the practical exercise at the head of this lesson is the best. If the experiment is performed as directed, nitrogen is the only gas that comes off, the carbonic acid being absorbed by excess of soda. The amount of nitrogen is a measure of the amount of urea.

The quantity of urea excreted is somewhat variable, the chief cause of variation being the amount of proteid food ingested. In a man in a state of equilibrium the quantity of urea excreted daily averages 33 grammes (500 grains). The normal percentage in human urine is 2 per cent.; but this also varies, because the concentration of the urine varies considerably in health. In dogs it may be 10 per cent. The excretion of urea is usually at a maximum three hours after a meal, especially after a meal rich in proteids. The urea does not

come, however, direct from the food; the food must be first assimilated, and become part of the body before it can break down to form urea. An exception to this rule is to be found in the case of the amino-acids, especially leucine and arginine, which are formed in the intestinal canal from proteids during digestion. These substances are carried to the liver and converted into urea; but only a very small fraction of the urea in the urine is formed in this way. Food increases the elimination of urea because it stimulates the tissues to increased activity; their waste nitrogenous products are converted into urea, which, passing into the blood, is directly excreted by the kidneys. The greater the amount of proteid food given, the more waste products do the tissues discharge from their protoplasm, in order to make room for the new proteid which is built into its substance.

Muscular exercise has little immediate effect on the amount of urea discharged. During intense muscular work there is a slight immediate increase of urea, but this is quite insignificant when compared with the increase of work. This is strikingly different from what occurs in the case of carbonic acid; the more the muscles work, the more carbonic acid do they send into venous blood, which is rapidly discharged by the expired air. Careful research has, however, shown that an increase of nitrogenous waste does occur on muscular exertion, but appears as urea in the urine to only a slight extent on the day of the work; the greater part is excreted during the next day.

Where is Urea formed?—The older authors considered that it was formed in the kidneys, just as they also erroneously thought that carbonic acid was formed in the lungs. Prévost and Dumas were the first to show that after complete extirpation of the kidney the formation of urea goes on, and that it accumulates in the blood and tissues. Similarly, in those cases of disease in which the kidneys cease work, urea is still formed and accumulates. This condition is called *uræmia* (or urea in the blood), and unless the urea be discharged from the body the patient dies.

Uræmia.—This term was originally applied on the erroneous supposition that it is urea or some antecedent of urea which acts as the poison. There is no doubt that the poison is not any constituent of normal urine; if the kidneys of an animal are extirpated, the animal dies in a few days, but there are no symptoms of uræmia. In man, also, if the kidneys are healthy or approximately so, and suppression of urine occurs from the simultaneous blocking of both renal arteries by clot, or of both ureters by stones, again uræmia does not follow. On the other hand, uræmia may occur even while a patient with diseased kidneys is passing a considerable amount of urine. What the poison is that is responsible for the convulsions and coma is unknown. It is doubtless some abnormal katabolic product, but whether this is produced by the diseased kidney cells, or in some other part of the body, is also unknown.

Where, then, is the seat of urea formation? Nitrogenous waste occurs in all the living tissues, and the principal final result of this proteid metabolism is urea. It may not be that the formation of urea is perfected in each tissue, for if we look to the most abundant tissue, the muscular tissue, very little urea is to be found. Yet there can be no doubt that the chief place from which urea ultimately comes is the muscular tissue. Some intermediate step occurs in the muscles; the final steps occur elsewhere.

In muscles we find a substance called creatine in fairly large quantities. If creatine is injected into the blood it is discharged as creatinine. But there is very little creatinine in normal urine: what little there is can be nearly all accounted for by the creatine in the food; if the muscular creatine and creatinine are discharged as urea, they must undergo some further change before they leave the muscle.

Similarly, other cellular organs, spleen, lymphatic glands, secreting glands, participate in the formation of urea; but the most important appears to be the liver: at any rate this is the organ where the final changes take place. The urea is then carried by the blood to the kidney, and is there excreted.

The facts of experiment and of pathology point very strongly in support of the theory that urea is formed in the liver. The principal are the following:—

1. After removal of the liver in such animals as frogs, urea formation almost ceases, and ammonia is found in the urine instead.

2. In mammals, the extirpation of the liver is such a serious operation that the animals die. But the liver of mammals can be very largely thrown out of gear by the operation known as Eck's fistula, which consists in connecting the portal vein directly to the inferior vena cava. Under these circumstances the liver receives blood only by the hepatic artery. The amount of urea is lessened, and its place is taken by ammonia.

3. When degenerative changes occur in the liver, as in *cirrhosis* of that organ, the urea formed is much lessened, and its place is taken by ammonia. In *acute yellow atrophy* urea is almost absent in the urine, and, again, there is considerable increase in the ammonia. In this disease leucine and tyrosine are also found in the urine; undue stress should not be laid upon this latter fact, for the small amounts of leucine and tyrosine found doubtless originate in the intestine, and, escaping further decomposition in the degenerated liver, pass as such into the urine.

We have to consider next the intermediate stages between proteid

and urea. A few years ago Drechsel succeeded in artificially producing urea from casein. More recent work has shown that this is true for other proteids also. If a proteid is decomposed by hydrochloric acid, a little stannous chloride being added to prevent oxidation, a number of products are obtained, such as ammonium salts, leucine, tyrosine, aspartic and glutaminic acids. This was known before, so the chief interest centres round two new substances, precipitable by phosphotungstic acid. One of these is called *lysine* ($C_6H_{14}N_2O_2$, probably di-amino-caproic acid); the other was first called *lysatinine*. Hedin then showed that lysatinine is a mixture of lysine with another base called *arginine* ($C_6H_{14}N_4O_2$); it is from the arginine that the urea comes in the experiment to be next described. Arguing from some resemblances between this substance and creatine, Drechsel expected to be able to obtain urea from it, and his expectation was confirmed by experiment. He took a silver compound of the base, boiled it with barium carbonate, and after twenty-five minutes boiling obtained urea. Drechsel's comparison of arginine to creatine has turned out to be correct; on decomposition it breaks up into di-amino-valerianic acid and cyanamide ($CN.NH_2$) from which the urea originates. (Schulze and Winterstein.) Lysine and arginine are two of the hexone bases (see p. 40).

It is, however, extremely doubtful whether the chemical decompositions produced in laboratory experiments on proteids are comparable with those occurring in the body. Many physiologists consider that the amino- acids are intermediate stages in the metabolic processes that lead to the formation of urea from proteids. We have already alluded to this question in relation to the creatine of muscle, and we are confronted with the difficulty that injection of creatine into the blood leads to an increase not of urea, but of creatinine in the urine. If creatine is an intermediate step, it must undergo some further change before it leaves the muscle. Other amino- acids, such as glycine (amino-acetic acid) and leucine (amino-caproic acid) and arginine, are to be included in the same category. The facts upon which such a theory depends are (1) that the introduction of glycine or leucine into the bowel, or into the circulation, leads to an increase of urea in the urine; there is, however, no evidence that tyrosine acts in this way; and (2) that amino- acids appear in the urine of patients suffering from acute yellow atrophy of the liver. Then again it is perfectly true that, in the laboratory, urea can be obtained from creatine, and also from uric acid, but such experiments do not prove that creatine and uric acid are normally intermediate products of urea formation in the body. Still, if we admit for the sake of argu-

ment that amino- acids are normally intermediate stages in proteid metabolism, and glance at their formulæ—

Glycine, $C_2H_5NO_2$	Creatine, $C_4H_9N_3O_2$
Leucine, $C_6H_{13}NO_2$	Arginine, $C_6H_{14}N_4O_2$

—we see that the carbon atoms are more numerous than the nitrogen atoms. In urea, CON_2H_4, the reverse is the case. The amino-acids must therefore be split into simpler compounds, which unite with one another to form urea. Urea formation is thus in part synthetic. There have been various theories advanced as to what these simpler compounds are. Some have considered that cyanate, others that carbamate, and others still that carbonate of ammonium is formed. Schröder's work proves that ammonium carbonate is one of the urea precursors, if not the principal one. The equation which represents the reaction is as follows:—

$$\underset{\text{[ammonium carbonate]}}{(NH_4)_2CO_3} - \underset{\text{[water]}}{2H_2O} = \underset{\text{[urea]}}{CON_2H_4}$$

Schröder's principal experiment was this: a mixture of defibrinated blood and ammonium carbonate was injected into the liver by the portal vein; the blood leaving the liver by the hepatic vein was found to contain urea in great abundance. This does not occur when the same experiment is performed with any other organ of the body, so that Schröder's experiments also prove the great importance of the liver in urea formation. Similar results were obtained by Nencki with ammonium carbamate.

There is, however, no necessity to suppose that the formation of amino- acids is a necessary preliminary to urea formation. The conversion of the leucine and arginine formed in the intestine into ammonium salts and then into urea does certainly occur, but this only accounts for quite an insignificant fraction of the urea in the urine. If this also occurs in tissue metabolism, we ought to find considerable quantities of leucine, glycine, creatine, arginine, and such substances in the blood leaving the various tissues and entering the liver; but we do not. We do, however, constantly find ammonia, which, after passing into the blood or lymph, has united with carbonic acid to form either carbonate or carbamate of ammonium. It is quite probable that the nitrogenous waste which leaves the muscles and other tissues is split off from them as ammonia, and not in the shape of large molecules of amino- acid which are subsequently converted into ammonia. The experiments outside the body which most closely imitate those occurring within the body are those of Drechsel,

in which he passed strong alternating currents through solutions of proteid-like materials. Such alternating currents are certainly absent in the body, but their effect, which is a rapidly changing series of small oxidations and reductions, is analogous to metabolic processes; under such circumstances the carbon atoms are burnt off as carbon dioxide, the nitrogen being split off in the form of ammonia, and by the union of these two substances ammonium carbonate is formed.

The following structural formulæ show the relationship between ammonium carbonate, ammonium carbamate, and urea :—

$O=C<^{O.NH_4}_{O.NH_4}$ [ammonium carbonate] $O=C<^{NH_2}_{O.NH_4}$ [ammonium carbamate] $O=C<^{NH_2}_{NH_2}$ [urea or carbamide]

The loss of one molecule of water from ammonium carbonate produces ammonium carbamate; the loss of a second molecule of water produces urea.

AMMONIA

The urine of man and carnivora contains small quantities of ammonium salts. In man the daily amount of ammonia excreted varies between 0·3 and 1·2 grammes: the average is 0·7 gramme. The ingestion of ammonium carbonate does not increase the amount of ammonia in the urine, but increases the amount of urea, into which substance the ammonium carbonate is easily converted. But if a more stable salt, like ammonium chloride, is given, it appears as such in the urine.

Under normal circumstances the amount of ammonia depends on the adjustment between the production of acid substances in metabolism and the supply of bases in the food. Ammonia formation is the physiological remedy for deficiency of bases.

When the production of acids is excessive (as in diabetes), or when mineral acids are given by the mouth or injected into the blood-stream, the result is an increase of the physiological remedy, and excess of the ammonia passes over in the urine. Under normal circumstances ammonia is kept at a minimum, being finally converted into the less toxic substance urea, which the kidneys easily excrete. The defence of the organism against acids which are very toxic is an increase of ammonia formation, or, to put it more correctly, less of the ammonia formed is converted into urea.

Under the opposite conditions, namely, excess of alkali, either in food or given as such, the ammonia disappears from the urine, all being converted into urea. Hence the diminution of ammonia in the

urine of man on a vegetable diet, and its absence in the urine of herbivorous animals.

Not only is this the case, but if ammonium chloride is given to a herbivorous animal like a rabbit, the urinary ammonia is but little increased. It reacts with sodium carbonate in the tissues, forming ammonium carbonate (which is excreted as urea) and sodium chloride. Herbivora also suffer much more from, and are more easily killed by, acids than carnivora, their organisation not permitting a ready supply of ammonia to neutralise excess of acids.

THE INORGANIC CONSTITUENTS OF URINE

The inorganic or mineral constituents of urine are chiefly chlorides, phosphates, sulphates, and carbonates; the metals with which these are in combination are sodium, potassium, ammonium, calcium, and magnesium. The total amount of these salts varies from 19 to 25 grammes daily. The most abundant is sodium chloride, which averages in amount 10 to 13 grammes per diem. These substances are derived from two sources—first from the food, and secondly as the result of metabolic processes. The chlorides and most of the phosphates come from the food; the sulphates and some of the phosphates are a result of metabolism. The salts of the blood and of the urine are much the same, with the important exception that, whereas the blood contains only traces of sulphates, the urine contains abundance of these salts. The sulphates are derived from the changes that occur in the proteids of the body; the nitrogen of proteids leaves the body as urea and uric acid; the sulphur of the proteids is oxidised to form sulphuric acid, which passes into the urine in the form of sulphates. The excretion of sulphates, moreover, runs parallel to that of urea. The chief tests for the various salts have been given in the practical exercises at the head of this lesson.

Chlorides.—The chief chloride is that of sodium. The ingestion of sodium chloride is followed by its appearance in the urine, some on the same day, some on the next day. Some is decomposed to form the hydrochloric acid of the gastric juice. The salt, in passing through the body, fulfils the useful office of stimulating metabolism and excretion.

Sulphates.—The sulphates in the urine are principally those of potassium and sodium. They are derived from the metabolism of proteids in the body. Only the smallest trace enters the body with the food. Sulphates have an unpleasant bitter taste (for instance, Epsom salts); hence we do not take food that contains them. The sulphates vary in amount from 1·5 to 3 grammes daily.

In addition to these sulphates there is a small quantity of sulphuric acid comprising about one-tenth of the total which is combined with organic radicles; the compounds are known as ethereal sulphates, and they originate from putrefactive processes occurring in the intestine. The most important of these ethereal sulphates are phenyl sulphate of potassium and indoxyl sulphate of potassium. The latter originates from the indole formed in the intestine, and as it yields indigo when treated with certain reagents it is sometimes called *indican*. It is very important to remember that the indican of urine is not the same thing as the indican of plants. Both yield indigo, but there the resemblance ceases.

The equation representing the formation of potassium phenyl-sulphate is as follows :—

$$\underset{\text{[phenol]}}{C_6H_5OH} + \underset{\text{[potassium hydrogen sulphate]}}{SO_2{<}^{OH}_{OK}} = \underset{\text{[potassium phenyl-sulphate]}}{SO_2{<}^{OC_6H_5}_{OK}} + \underset{\text{[water]}}{H_2O}$$

The formation of potassium indoxyl-sulphate may be represented as follows :— Indole, $C_6H_4{<}^{CH.CH}_{NH}$, on absorption is converted into indoxyl : $C_6H_4{<}^{C.OH.CH.}_{NH}$

Indoxyl then interacts with potassium hydrogen sulphate as follows :—

$$\underset{\text{[indoxyl]}}{C_8H_7NO} + \underset{\text{[potassium hydrogen sulphate]}}{SO_2{<}^{OH}_{OK}} = \underset{\text{[potassium indoxyl-sulphate]}}{SO_2{<}^{OC_8H_6N}_{OK}} + \underset{\text{[water]}}{H_2O}$$

The formation of such sulphates is important; the aromatic substances liberated by putrefactive processes in the intestine are poisonous, but their conversion into ethereal sulphates renders them innocuous.

Carbonates.—Carbonates and bicarbonates of sodium, calcium, magnesium, and ammonium are only present in alkaline urine. They arise from the carbonates of the food, or from vegetable acids (malic, tartaric, &c.) in the food. They are, therefore, found in the urine of herbivora and vegetarians, whose urine is thus rendered alkaline. Urine containing carbonates becomes, like saliva, cloudy on standing, the precipitate consisting of calcium carbonate, and also phosphates.

Phosphates.—Two classes of phosphates occur in normal urine :—

(1) Alkaline phosphates—that is, phosphates of sodium (abundant) and potassium (scanty).

(2) Earthy phosphates—that is, phosphates of calcium (abundant) and magnesium (scanty).

The composition of the phosphates in urine is liable to variation. In acid urine the acidity is due to the acid salts. These are chiefly sodium dihydrogen phosphate, NaH_2PO_4, and calcium dihydrogen phosphate, $Ca(H_2PO_4)_2$.

In neutral urine, in addition, disodium hydrogen phosphate (Na_2HPO_4), calcium hydrogen phosphate, $CaHPO_4$, and magnesium hydrogen phosphate, $MgHPO_4$, are found. In alkaline urine there may be instead of, or in addition to the above, the normal phosphates of sodium, calcium, and magnesium [Na_3PO_4, $Ca_3(PO_4)_2$, $Mg_3(PO_4)_2$].

The earthy phosphates are precipitated by rendering the urine alkaline by ammonia. In decomposing urine ammonia is formed from the urea: this also precipitates the earthy phosphates. The phosphates most frequently found in the white creamy precipitate which occurs in decomposing urine are—

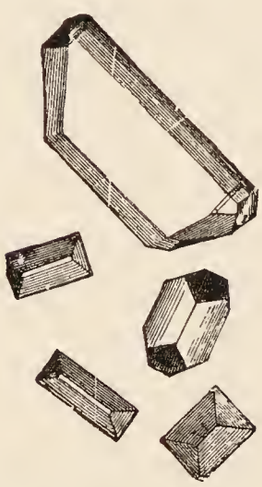


FIG. 42.—Ammonio-magnesium or triple phosphate.

(1) Triple phosphate or ammonio-magnesium phosphate ($NH_4MgPO_4 + 6H_2O$). This crystallises in 'coffin-lid' crystals (see fig. 42) or feathery stars.

(2) Stellar phosphate, or calcium phosphate, which crystallises in star-like clusters of prisms.

As a rule, normal urine gives no precipitate when it is boiled; but sometimes neutral, alkaline, and occasionally faintly acid urines give a precipitate of calcium phosphate when boiled; this precipitate is amorphous, and is liable to be mistaken for albumin. It may be distinguished readily from albumin, as it is soluble in a few drops of acetic acid, whereas coagulated proteid does not dissolve.

The phosphoric acid in the urine chiefly originates from the phosphates of the food, but is partly a decomposition product of the phosphorised organic materials in the body, such as lecithin and nuclein. The amount of P_2O_5 in the twenty-four hours' urine varies from 2·5 to 3·5 grammes, of which the earthy phosphates contain about half (1 to 1·5 gr.).

LESSON XI

URINE (*continued*)

1. **Urea Nitrate.**—Evaporate some urine in a capsule to a quarter of its bulk. Pour the concentrated urine into a watch-glass ; let it cool, and add a few drops of strong, but not fuming, nitric acid. Crystals of urea nitrate separate out. Examine these microscopically.

2. **Urea Oxalate.**—Concentrate the urine as in the last exercise, and add oxalic acid. Crystals of urea oxalate separate out. Examine these microscopically.

3. **Uric Acid.**—Examine microscopically the crystals of uric acid in some urine, to which 5 per cent. of hydrochloric acid has been added twenty-four hours previously. Note that they are deeply tinged with pigment, and to the naked eye look like granules of cayenne pepper.

When microscopically examined, the crystals are seen to be large bundles, principally in the shape of barrels, with spicules projecting from the ends, and whetstones. If oxalic acid is used instead of hydrochloric acid in this experiment, the crystals are smaller, and more closely resemble those observed in pathological urine in cases of uric acid gravel (see fig. 43).

Dissolve the crystals in caustic potash and then carefully add excess of hydrochloric acid. Small crystals of uric acid again form.

Murexide Test.—Place a little uric acid, or a urate (for instance, serpent's urine), in a capsule ; add a little dilute nitric acid and evaporate to dryness. A yellowish-red residue is left. Add a little ammonia carefully. The residue turns to violet. This is due to the formation of murexide or purpurate of ammonia. On the addition of potash the colour becomes bluer.

Schiff's Test.—Dissolve some uric acid in sodium carbonate solution. Put a drop of this on blotting paper, add a drop of silver nitrate, and warm gently ; the black colour of reduced silver is seen on the paper.

4. **Deposits of Urates or Lithates (Lateritious Deposit).**—The specimen of urine from the hospital contains excess of urates, which have become deposited on the urine becoming cool. They are tinged with pigment (uroerythrin), and have a pinkish colour, like brickdust ; hence the term 'lateritious.' Examine microscopically. The deposit is usually amorphous—that is, non-crystalline. Sometimes crystals of calcium oxalate (envelope crystals—octahedra) are seen also ; these are colourless.

The deposit of urates dissolves on heating the urine.

5. **Deposit of Phosphates.**—Another specimen of pathological urine contains excess of phosphates, which have formed a white deposit on the urine becoming alkaline. This precipitate does not dissolve on heating ; it may be increased. It is, however, soluble in acetic acid. Examine microscopically for coffin-lid crystals of triple phosphate (ammonio-magnesium phosphate), for crystals of stellar (calcium) phosphate, and for mucus. Mucus is flocculent to the naked eye, amorphous to the microscope.

N.B.—On boiling neutral, alkaline, or even faintly acid urine it may become turbid from deposition of phosphates. The solubility of this deposit in

a few drops of acetic acid distinguishes it from albumin, for which it is liable to be mistaken.

Some of the facts described in the foregoing exercises have been already dwelt upon in the preceding lesson. They are, however, conveniently grouped together here, as all involve the use of the microscope.

We have now studied urea, the principal nitrogenous constituent of urine, at some length. There are still left for our consideration a number of other nitrogenous constituents, the most important of which are uric acid, hippuric acid, and creatinine.

URIC ACID

Uric acid ($C_5N_4H_4O_3$) is in mammals the medium by which only a small quantity of nitrogen is excreted from the body. It is, however, in birds and reptiles the principal nitrogenous constituent of their urine. It is not present in the free state, but is combined with bases to form urates.

It may be obtained from human urine by adding 5 c.c. of hydrochloric acid to 100 c.c. of the urine, and allowing the mixture to stand for twelve to twenty-four hours. The crystals which form are deeply tinged with urinary pigment, and though by repeated solution in caustic soda or potash, and reprecipitation by hydrochloric acid, they may be obtained fairly free from pigment, pure uric acid is more readily obtained from the solid urine of a serpent or bird, which consists principally of the acid ammonium urate. This is dissolved in soda, and then the addition of hydrochloric acid produces as before the crystallisation of uric acid from the solution.



FIG. 43.—Uric acid crystals.

The pure acid crystallises in colourless rectangular plates or prisms. In striking contrast to urea it is a most insoluble substance, requiring for its solution 1,900 parts of hot and 15,000 parts of cold water. The forms which uric acid assumes when precipitated from human urine, either by the addition of hydrochloric acid or in certain pathological processes, are very various, the most frequent being the whetstone shape; there are also bundles of crystals resembling sheaves, barrels, and dumb-bells (see fig. 43).

The murexide test which has just been described among the practical exercises is the principal test for uric acid. The test has

received the name on account of the resemblance of the colour to the purple of the ancients, which was obtained from certain snails of the genus *Murex*.

Another reaction that uric acid undergoes (though it is not applicable as a test) is that on treatment with certain oxidising reagents urea and oxalic acid can be obtained from it. It is, however, doubtful whether a similar oxidation occurs in the normal metabolic processes of the body (see p. 129).

Uric acid is dibasic, and thus there are two classes of urates—the normal urates and the acid urates. A normal urate is one in which two atoms of the hydrogen are replaced by two of a monad metal like sodium; an acid urate is one in which only one atom of hydrogen is thus replaced. The formulæ would be—

$C_5H_4N_4O_3$=uric acid
$C_5H_3NaN_4O_3$=acid sodium urate
$C_5H_2Na_2N_4O_3$=normal sodium urate.

The acid sodium urate is the chief constituent of the pinkish deposit of urates, which, as we have already stated, is called the lateritious deposit.

If uric acid is represented by H_2U, the normal urates may be represented by M_2U and the acid urates by MHU. Bence Jones, and later Sir W. Roberts, considered that the urates which actually occur in urine are quadriurates $MHU.H_2U$. There is much doubt whether such compounds actually exist; if they do they are readily decomposed into acid urates MHU and free uric acid.

The quantity of uric acid excreted by an adult varies from 7 to 10 grains (0·5 to 0·75 gramme) daily.

The best method for determining the quantity of uric acid in the urine is that of Hopkins. Ammonium chloride in crystals is added to the urine until no more will dissolve. This saturation completely precipitates all the uric acid in the form of ammonium urate. After standing for two hours the precipitate is collected on a filter, washed with saturated solution of ammonium chloride, and then dissolved in weak alkali. From this solution the uric acid is precipitated by neutralising with hydrochloric acid. The precipitate of uric acid is collected on a weighed filter, dried and weighed, or titration may be performed with potassium permanganate (see Advanced Course).

Origin of Uric Acid.—Uric acid is not made by the kidneys. When the kidneys are removed uric acid continues to be formed and accumulates in the organs, especially in the liver and spleen. The liver has been removed from birds, and uric acid is then hardly formed

at all, its place being taken by ammonia and lactic acid. It is therefore probable that in these animals ammonia and lactic acid are normally synthesised in the liver to form uric acid.

The principal conditions which lead to an increase of uric acid in the urine are—

1. Increase of meat diet and diminution of oxidation processes, such as occur in people with sedentary habits.

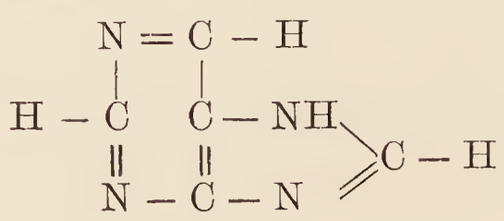
2. Increase of white corpuscles in the blood, especially in the disease known as leucocythæmia. This latter fact is of great interest, as leucocytes contain large quantities of nuclein. Nuclein yields nitrogenous bases which are closely related to uric acid.

Purine Substances.—The bases just alluded to are sometimes called the xanthine bases, because xanthine is a prominent member of the group. Sometimes they are termed the nuclein bases, because of their origin from nuclein. Another name they have received is the alloxuric bases, because they contain in combination two radicles, one of alloxan, and the other of urea. Emil Fischer's recent work, however, has shown that they are to be rather regarded as derivatives of a substance he has named purine; hence the name by which they are generally called now, the purine bases. The empirical formulæ for purine, the purine bases, and uric acid are as follows :—

Purine . .	$C_5H_4N_4$		
Hypoxanthine .	$C_5H_4N_4O$	Monoxypurine	Purine bases.
Xanthine . .	$C_5H_4N_4O_2$	Dioxypurine	
Adenine . .	$C_5H_3N_4.NH_2$	Amino-purine	
Guanine . .	$C_5H_3N_4O.NH_2$	Amino-oxypurine	
Uric acid . .	$C_5H_4N_4O_3$	Trioxypurine	

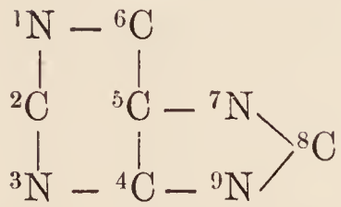
There are a vast number of purine derivatives, but only a few of them have at present any physiological importance. Others in addition to those already enumerated are theophylline (dimethyl-xanthine), theobromine (also a dimethyl-xanthine), caffeine (trimethyl-xanthine); these are of interest, as they occur in tea, cocoa, and coffee. A few words more may be added in respect to those in our list.

Purine itself has never been discovered in the body. It has the following structural formula :



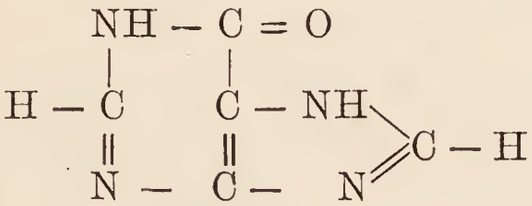
The purine nucleus is depicted in the next formula, and its atoms

have been empirically numbered as shown for convenience of description :—

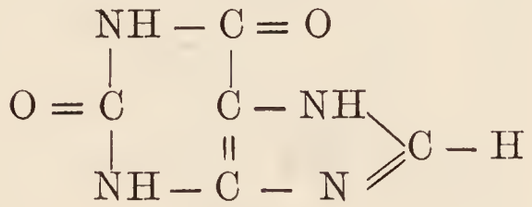


Hypoxanthine or **Sarcine** is found in the body tissues and fluids, and in the urine. It is derived from some nucleins, especially those from fishes' spermatozoa. It may be termed 6-oxypurine, as the oxygen is attached to the atom numbered 6 in the purine nucleus.

Xanthine is found with hypoxanthine in the body, and has been obtained from a number of nucleins (from spermatozoa, thymus, pancreas, &c.). It is 2, 6-dioxypurine, its oxygen atoms being attached to the atoms numbered 2 and 6 in the purine nucleus.



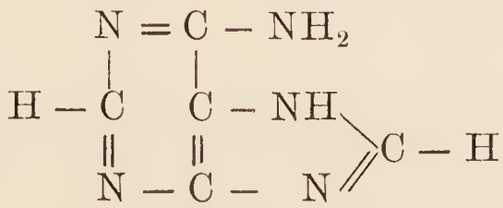
[hypoxanthine]



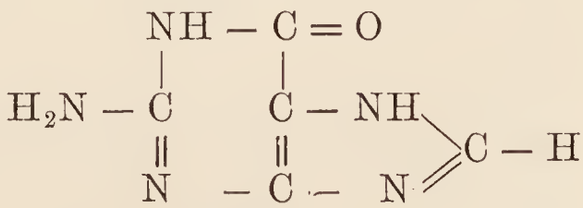
[xanthine]

Adenine in found in the tissues, blood, and urine. It is obtained from several nucleins, but especially from the nuclein derived from the thymus. It is 6-amino-purine.

Guanine is also a decomposition product of nucleins, especially of that obtained from the pancreas. Combined with calcium it gives the brilliancy to the scales of fishes, and is also found in the bright tapetum of the eyes in these animals. It is a constituent of guano, and here is probably derived from the fish eaten by marine birds. It is 2-amino-6-oxypurine.

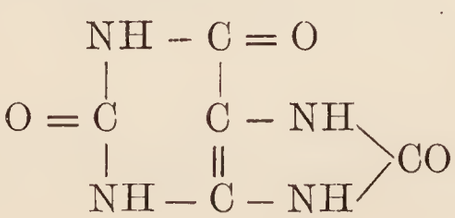


[adenine]



[guanine]

Uric acid is 2, 6, 8-trioxypurine.



[urie acid]

We have here a way in which uric acid may arise by oxidation from the nuclein bases, and thus ultimately from the nuclei of cells. Certain forms of diet increase uric acid formation by leading to an increase of leucocytes and consequently increase in the metabolism of their nuclei; in some cases, however, the increase is chiefly due to nuclein in the food. Uric acid, which comes from nuclein or purine substances in the food, is termed *exogenous*; that which arises from metabolism is termed *endogenous*. Although special attention has been directed to the nuclei of leucocytes because these can be readily examined during life, it must be remembered that the nuclein metabolism of all cells may contribute to uric acid formation. The synthetic origin of uric acid from ammonia and lactic acid, which is so important in birds, occurs in mammals to a slight extent only.



FIG. 44.—Hippuric acid crystals.

HIPPURIC ACID

Hippuric acid ($C_9H_9NO_3$), combined with bases to form hippurates, is present in small quantities in human urine, but in large quantities in that of herbivora. This is due to the food of herbivora containing substances belonging to the aromatic group—the benzoic acid series. If benzoic acid is given to a man, it unites with glycine with the elimination of a molecule of water, and is excreted as hippuric acid—

$$\underset{\text{[benzoic acid]}}{C_6H_5COOH} + \underset{\text{[glycine]}}{\begin{matrix}CH_2NH_2\\|\\COOH\end{matrix}} = \underset{\text{[hippuric acid]}}{\begin{matrix}CH_2NH.CO.C_6H_5\\|\\COOH\end{matrix}} + \underset{\text{[water]}}{H_2O}$$

This is a well-marked instance of synthesis carried out in the animal body, and experimental investigation shows that it is accomplished by the living cells of the kidney itself; for if a mixture of glycine, benzoic acid, and blood is injected through the kidney (or mixed with a minced kidney just removed from the body of an animal), their place is found to have been taken by hippuric acid.

It may be crystallised from horse's urine by evaporating to a syrup and saturating with HCl. The crystals are dissolved in boiling water, filtered, and on cooling the acid again crystallises out. It melts at 186° C., and on further heating gives rise to the odour of bitter almond oil.

CREATININE

The **creatinine** in the urine is nearly all derived from the creatine contained in the meat of the food. There is, however, a small amount in the urine even during starvation : this represents a small percentage of creatine from the muscles.

Sarcosine ($C_3H_7NO_2$) united to cyanamide ($CN.NH_2$) yields creatine ($C_4H_9N_3O_2$). Cyanamide *plus* water yields urea (CON_2H_4).

The formation of creatinine from creatine is represented thus :—

$$\underset{\text{[creatine]}}{C_4H_9N_3O_2} - \underset{\text{[water]}}{H_2O} = \underset{\text{[creatinine]}}{C_4H_7N_3O}$$

Creatine and creatinine are of considerable interest, because urea can be obtained from them as one of their decomposition products ; the equation which represents the formation of urea from creatine is as follows :—

$$\underset{\text{[creatine]}}{C_4H_9N_3O_2} + \underset{\text{[water]}}{H_2O} = \underset{\text{[sarcosine]}}{C_3H_7NO_2} + \underset{\text{[urea]}}{CON_2H_4}$$

The second substance formed is sarcosine or methyl-glycine—that is, amino-acetic acid in which one H is replaced by methyl (CH_3)

$$CH_2 \begin{matrix} NH.CH_3 \\ COOH \end{matrix}$$

It is, however, doubtful whether decompositions of this kind occur in the body (see p. 129). With sodium nitro-prusside creatinine gives a colour reaction described on p. 122.

Creatinine with zinc chloride gives a characteristic crystalline precipitate (groups of fine needles) with composition $C_4H_7N_3O.ZnCl_2$.

According to the researches of G. S. Johnson, urinary creatinine, though isomeric with the creatinine obtained artificially from the creatine of flesh, differs from it in some of its properties, such as reducing power, solubility, and character of its gold salts. The reducing action of urinary creatinine has led to some confusion, for some physiologists have supposed that the reducing action on Fehling's solution and picric acid of normal urine is due to sugar, whereas it is really chiefly due to creatinine. The readiest way of separating creatinine from urine is the following :—To the urine a twentieth of its volume of a saturated solution of sodium acetate is added, and then one-fourth of its volume of a saturated solution of mercuric chloride : this produces an immediate abundant precipitate of urates, sulphates, and phosphates, which is removed by filtration ; the filtrate is then allowed to stand for twenty-four hours, when the precipitation of a mercury salt of creatinine

$(C_4H_5HgN_3OHCl)3HgCl_2+2H_2O$ occurs in the form of minute spheres, quite typical on microscopic examination. This compound lends itself very well to quantitative analysis. It may be collected, dried, and weighed, and one-fifth of the weight found is creatinine.[1] Creatinine may be obtained from it by suspending it in water, decomposing it with sulphuretted hydrogen, and filtering. The filtrate



FIG. 45.—Creatine crystals.

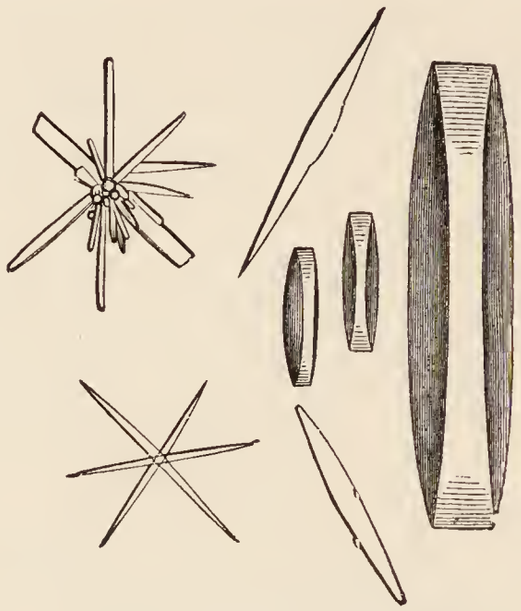


FIG. 46.—Creatinine crystals.

deposits creatinine hydrochloride, from which lead hydrate, $Pb(OH)_2$, liberates creatinine. An important point in Johnson's process is that all the operations are carried out in the cold ; if heat is applied one obtains the creatinine of former writers, which has no reducing power.

URINARY DEPOSITS

The different substances that may occur in urinary deposits are formed elements and chemical substances.

The **formed** or **anatomical elements** may consist of blood corpuscles, pus, mucus, epithelium cells, spermatozoa, casts of the urinary tubules, fungi, and entozoa. All of these, with the exception of a small quantity of mucus, which forms a flocculent cloud in the urine, are pathological, and the microscope is chiefly employed in their detection.

The **chemical substances** are uric acid, urates, calcium oxalate, calcium carbonate, and phosphates. Rarer forms are leucine, tyrosine, xanthine, and cystin. We shall, however, here only consider the commoner deposits, and for their identification the microscope and chemical tests must both be employed.

[1] If only a quantitative analysis is required, the process may be considerably accelerated by boiling the first filtrate for ten minutes instead of letting it stand for twenty-four hours.

Deposit of Uric Acid.—This is a sandy reddish deposit resembling cayenne pepper. It may be recognised by its crystalline form (fig. 43, p. 136) and by the murexide reaction. The presence of these crystals generally indicates an increased formation of uric acid, and, if excessive, may lead to the formation of stones or calculi in the bladder.

Deposit of Urates.—This is much commoner, and may, if the urine is concentrated, occur in nomal urine when it cools. It is generally found in the concentrated urine of fevers; and there appears to be a kind of fermentation, called the acid fermentation, which occurs in the urine after it has been passed, and which leads to the same result. The chief constituent of the deposit is the acid

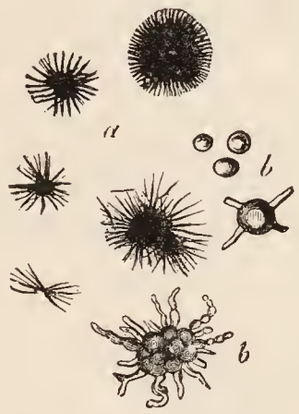


FIG. 47.—Acid sodium urate.



FIG. 48.—Acid ammonium urate

sodium urate, the formation of which from the normal sodium urate of the urine may be represented by the equation—

$$2C_5H_2Na_2N_4O_3 + H_2O + CO_2 = 2C_5H_3NaN_4O_3 + Na_2CO_3$$

[normal sodium urate] [water] [carbonic acid] [acid sodium urate] [sodium carbonate]

This deposit may be recognised as follows:—

1. It has a pinkish colour; the pigment called *uro-erythrin* is one of the pigments of the urine, but its relationship to the other urinary pigments is not known (see further Lesson XXVI.).

2. It dissolves upon warming the urine.

Microscopically it is usually amorphous, but crystalline forms similar to those depicted in figs. 47 and 48 may occur.

Crystals of calcium oxalate may be mixed with this deposit (see fig. 49).

Deposit of Calcium Oxalate.—This occurs in envelope crystals (octahedra) or dumb-bells.

It is insoluble in ammonia, and in acetic acid. It is soluble with difficulty in hydrochloric acid.

Deposit of Cystin.—Cystin ($C_6H_{12}N_2S_2O_4$) is recognised by its

colourless six-sided crystals (fig. 50). These are rare: they occur only in acid urine, and they may form concretions or calculi. Cystinuria (cystin in the urine) is hereditary.

Deposit of Phosphates.—These occur in alkaline urine. The urine may be alkaline when passed, due to fermentative changes

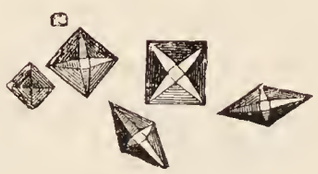


FIG. 49.—Envelope crystals of calcium oxalate.

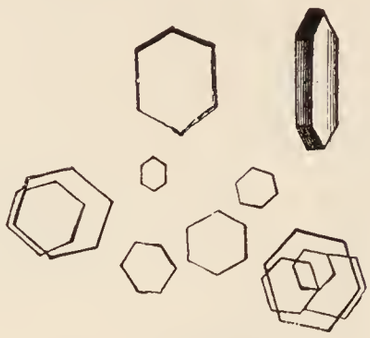


FIG. 50.—Cystin crystals.

occurring in the bladder. All urine, however, if exposed to the air (unless the air is perfectly pure, as on the top of a snow mountain), will in time become alkaline owing to the growth of the *micrococcus ureæ*. This forms ammonium carbonate from the urea.

$$\underset{\text{[urea]}}{CON_2H_4} + \underset{\text{[water]}}{2H_2O} = \underset{\text{[ammonium carbonate]}}{(NH_4)_2CO_3}$$

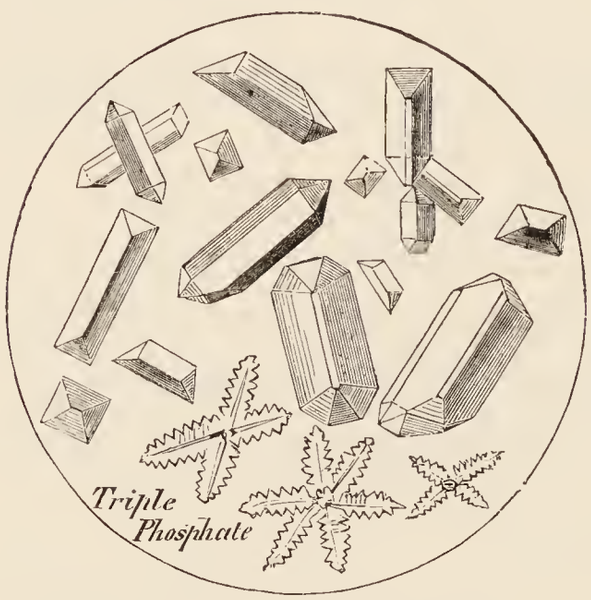


FIG. 51.—Triple phosphate crystals.



FIG. 52.—Crystals of phosphate of lime (stellar phosphate).

The ammonia renders the urine alkaline, and precipitates the earthy phosphates. The chief forms of phosphates that occur in urinary deposits are—

1. Calcium phosphate, $Ca_3(PO_4)_2$; amorphous.

2. Triple or ammonio-magnesium phosphate, $MgNH_4PO_4$; coffin-lids and feathery stars (fig. 51).

3. Crystalline phosphate of calcium, $CaHPO_4$, in rosettes of prisms, in spherules, or in dumb-bells (fig. 52).

4. Magnesium phosphate, $Mg_3(PO_4)_2 + 2H_2O$, occurs occasionally, and crystallises in long plates.

All these phosphates are dissolved by acids, such as acetic acid, without effervescence.

They do not dissolve on heating the urine ; in fact, the amount of precipitate may be increased by heating.

A solution of ammonium carbonate (1-in-5) eats magnesium phosphate away from the edges ; it has no effect on the triple phosphate. A phosphate of calcium ($CaHPO_4 + 2H_2O$) may occasionally be deposited in acid urine. Pus in urine is apt to be mistaken for phosphates, but can be distinguished by the microscope.

Deposit of calcium carbonate, $CaCO_3$, appears but rarely as whitish balls or biscuit-shaped bodies. It is commoner in the urine of herbivora (see p. 133). It dissolves in acetic or hydrochloric acid, with effervescence.

The following is a summary of the chemical sediments that may occur in urine :—

CHEMICAL SEDIMENTS IN URINE

In Acid Urine

Uric Acid.—Whetstone, dumb-bell, or sheaf-like aggregations of crystals deeply tinged by pigment (fig. 43).

Urates.—Generally amorphous. The acid urate of sodium (fig. 47) and of ammonium (fig. 48) may sometimes occur in star-shaped clusters of needles or spheroidal clumps with projecting spines. Tinged brick-red. Soluble on warming.

Calcium Oxalate.—Octahedra, so-called envelope crystals (fig. 49). Insoluble in acetic acid.

Cystin.—Hexagonal plates (fig. 50). Rare.

Leucine and Tyrosine.—Rare.

Calcium Phosphate, $CaHPO_4 + 2H_2O$.—Rare.

In Alkaline Urine

Phosphates.—Calcium phosphate, $Ca_3(PO_4)_2$. Amorphous.

Triple phosphate, $MgNH_4PO_4 + 6H_2O$. Coffin-lids or feathery stars (figs. 42 and 51).

Calcium hydrogen phosphate, $CaHPO_4$. Rosettes, spherules, or dumb-bells (fig. 52).

Magnesium phosphate, $Mg_3(PO_4)_2 + 22H_2O$. Long plates.

All soluble in acetic acid without effervescence.

Calcium Carbonate, $CaCO_3$.—Biscuit-shaped crystals. Soluble in acetic acid with effervescence.

Ammonium Urate, $C_5H_2(NH_4)_2.N_4O_3$. — 'Thorn-apple' spherules.

Leucine and Tyrosine.—Very rare.

LESSON XII

PATHOLOGICAL URINE

1. **Urine A** is pathological urine containing albumin. It gives the usual proteid tests. The two following are most frequently used in practice.

(*a*) Boil the top of a long column of urine in a test-tube. If the urine is acid, the albumin is coagulated. If the quantity of albumin is small, the cloudiness produced is readily seen, as the unboiled urine below it is clear. This is insoluble in a few drops of acetic acid, and so may be distinguished from phosphates. If the urine is alkaline, it should be first rendered acid with a little dilute acetic acid.

(*b*) *Heller's Nitric-Acid Test.*—Pour some of the urine gently on to the surface of some nitric acid in a test-tube. A ring of white precipitate occurs at the junction of the two liquids. This test is used for small quantities of albumin.

If the urine is cloudy, it should be filtered before applying these tests.

2. **Estimation of Albumin by Esbach's Albuminometer.**—Esbach's reagent for precipitating the albumin is made by dissolving 10 grammes of picric acid and 20 grammes of citric acid in 800 or 900 c.c. of boiling water, and then adding sufficient water to make up to a litre (1,000 c.c.).



FIG. 53.—Albuminometer of Esbach.

Pour the urine into the tube up to the mark U; then the reagent up to the mark R. Close the tube with a cork, and to ensure complete mixture tilt it to and fro a dozen times without shaking. Allow the corked tube to stand upright twenty-four hours; then read off on the scale the height of the coagulum. The figures indicate grammes of dried albumin in a litre of urine. The percentage is obtained by dividing by 10. Thus, if the coagulum stands at 3, the amount of albumin is 3 grammes per litre, or 0·3 gr. in 100 c.c. If the sediment falls between any two figures, the distance $\frac{1}{4}$, $\frac{1}{2}$, or $\frac{3}{4}$ from the upper or lower figure can be read off with sufficient accuracy. Thus the surface of the sediment, being midway between 3 and 4, would be read as 3·5. When the albumin is so abundant that the sediment is above 4, a more accurate result is obtained by first diluting the urine with one or two volumes of water, and then multiplying the resulting figure by 2 or 3, as the case may be. If the amount of albumin is less than 0·05 per cent., it cannot be accurately estimated by this method.

3. **Urine B** is diabetic urine. It has a high specific gravity. The presence of sugar is shown by the reduction (yellow precipitate of cuprous oxide) that occurs on boiling with Fehling's solution. Fehling's solution is an alkaline solution of copper sulphate to which Rochelle salt has been added. The Rochelle salt (double tartrate of potash and soda) holds the cupric hydrate in

solution. Fehling's solution should always be freshly prepared, as, on standing, an isomeride is formed from the tartaric acid, and this substance itself reduces the cupric to cuprous oxide. Fehling's solution should, therefore, always be tested by boiling before it is used. If it remains unaltered by boiling, it is in good condition.

4. **Quantitative Determination of Sugar in Urine.**—Fehling's solution is prepared as follows—34·639 grammes of copper sulphate are dissolved in about 200 c.c. of distilled water; 173 grammes of Rochelle salt are dissolved in 600 c.c. of a 14-per-cent. solution of caustic soda. The two solutions are mixed and diluted to a litre. Ten c.c. of this solution are equivalent to 0·05 gramme of dextrose. Dilute 10 c.c. of this solution with about 40 c.c of water, and boil it in a white porcelain dish. Run into this from the burette (see fig. 54) the urine (which should be previously diluted with nine times its volume of distilled water) until the blue colour of the copper solution disappears—that is, till all the cupric hydrate is reduced. The mixture in the basin should be boiled after every addition.[1] The quantity of diluted urine used from the burette contains 0·05 gramme of sugar. Calculate the percentage from this, remembering that urine has been diluted to ten times its original volume.

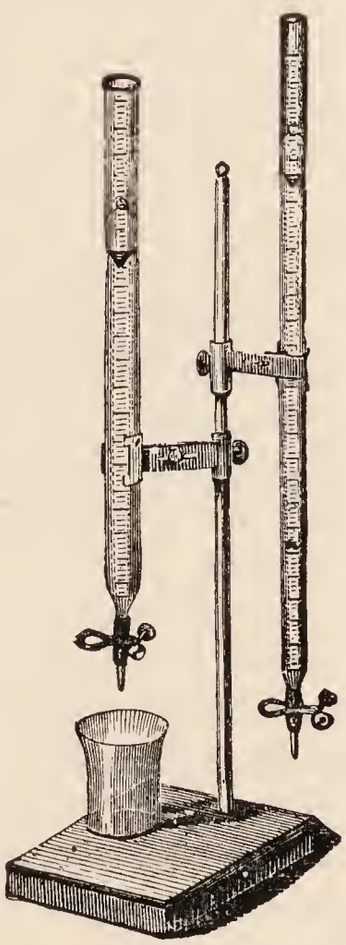


Fig. 54.—Two burettes on stand. (Sutton.)

The following formula will be found useful in converting grammes into grains:—

x = number of grains of sugar in the 24 hours.

a = number of ounces of urine in the 24 hours.

1 ounce = 28·396 c.c.

b = number of c.c. of urine used from the burette to decompose 10 c.c. of Fehling's solution (equivalent to 0·05 gramme = 0·77 grain of sugar).

Then

$$x = \frac{a}{b} \times 28{\cdot}396 \times 0{\cdot}77 = \frac{a}{b} \times 21{\cdot}865.$$

Pavy's modification of Fehling's solution is sometimes used. Here ammonia holds the cuprous oxide in solution, so that no precipitate forms on boiling Pavy's solution with a reducing sugar. The reduction is complete when the blue colour disappears: 10 c.c. of Pavy's solution = 1 c.c. of Fehling's solution = 0·005 gramme of dextrose. In some cases of diabetic urine where there is excess of ammonio-magnesic phosphate, the full reduction is not obtained with Fehling's solution, and when the quantity of sugar is small it may be missed. In such a case excess of soda or potash should be first added; the precipitated phosphates filtered off; and the filtrate, after it has been well boiled, may then be titrated with Fehling's or Pavy's solution.

5. **Picric Acid Test.**—The work of Sir George Johnson and G. S. Johnson has shown the value of this reagent in detecting both albumin and sugar in the urine. The same reagent may be employed for the detection of both substances. The method of testing for albumin has been already studied with Esbach's tubes. To test for sugar perform the following experiment. Take a drachm (about 4 c.c) of diabetic urine; add to it an equal volume of saturated aqueous solution of picric acid, and half the volume (*i.e.* 2 c.c.) of the liquor potassæ of the British Pharmacopœia. Boil the mixture for about

[1] On cooling the blue colour reappears, owing to re-oxidation

a minute, and it becomes so intensely dark red as to be opaque. Now do the same experiment with normal urine. An orange-red colour appears even in the cold, and is deepened by boiling, but it never becomes opaque, and so the urine for clinical purposes may be considered free from sugar. This reduction of picric acid by normal urine is due to creatinine (see p. 141).

The full significance and causes of pathological urine cannot be appreciated until a theoretical and practical acquaintance with disease is obtained, and we shall briefly consider only those abnormal constituents which are most frequently met with.

PROTEIDS IN THE URINE

There is no proteid matter in normal urine, and the most common cause of the appearance of albumin in the urine is disease of the kidney (Bright's disease). The best methods of testing for and estimating the albumin are given in the practical heading to this lesson. The term 'albumin' is the one used by clinical observers. Properly speaking, it is a mixture of serum albumin and serum globulin.

A condition called 'peptonuria,' or peptone in the urine, is observed in certain pathological states, especially in diseases where there is a formation of pus, and particularly if the pus is decomposing owing to the action of a bacterial growth called staphylococcus; one of the products of disintegration of pus cells appears to be peptone; and this leaves the body by the urine. The term 'peptone,' however, is in the strict sense incorrect; the proteid present is deutero-proteose. In the disease called 'osteomalacia' a proteose is usually found in the urine, which more nearly resembles hetero-proteose in its characters.

SUGAR IN THE URINE

Normal urine contains no sugar, or so little that for clinical purposes it may be considered absent. It occurs in the disease called diabetes mellitus, which can be artificially produced by the methods described on p. 78.

The methods usually adopted for detecting and estimating the sugar are given at the head of this lesson. The sugar present is dextrose. Lactose may occur in the urine of nursing mothers. Diabetic urine also contains hydroxybutyric acid, and may contain or yield on distillation acetone and ethyl-diacetic acid.

Fehling's test is not absolutely trustworthy. Often a normal urine will decolorise Fehling's solution, though seldom a red precipitate is formed. This is due to excess of urates and creatinine. Another substance called

glycuronic acid ($C_6H_{10}O_7$) is, however, very likely to be confused with sugar by Fehling's test; the cause of its appearance is sometimes the administration of drugs (chloral, camphor, &c.); but sometimes it appears independently of drug treatment (see p. 79).

Then, too, in the rare condition called alcaptonuria, confusion may similarly arise. Alcapton is a substance which probably originates from tyrosine by an unusual form of metabolism. It gives the urine a brown tint, which darkens on exposure to the air. It is an aromatic substance, and the researches of Baumann and Wolkow have identified it with homogentisinic acid ($C_6H_3.(OH)_2CH_2.COOH$).

The best confirmatory tests for sugar are the *phenyl-hydrazine test* (see Lesson XIII.), and the *fermentation test* (Lesson II.).

Sir W. Roberts introduced a method for estimating sugar in urine, by the diminution in specific gravity which it undergoes on fermentation with yeast. Every degree lost in the specific gravity corresponds to one grain of sugar per fluid ounce. Suppose that the specific gravity of the unfermented urine is 1040, and that of the urine which has undergone fermentation is 1030: the number of degrees lost is ten; *i.e.* the urine contained 10 grains of sugar per ounce. The percentage of sugar may be ascertained by multiplying the degrees of specific gravity lost by 0·22; thus the percentage in the example just given will be 0·22 × 10 = 2·2. The method, however, is rough and has dropped out of use.

BILE IN THE URINE

This occurs in jaundice. The urine is dark-brown, greenish, or in extreme cases almost black in colour. The most readily applied test is Gmelin's test for the bile pigments. Pettenkofer's test for the bile acids seldom succeeds in urine if the test is done in the ordinary way. The best method is to warm a thin film of urine and cane-sugar solution in a flat porcelain dish. Then dip a glass rod in strong sulphuric acid, and draw it across the film. Its track is marked by a purplish line. Hay's sulphur test (p. 70) is very trustworthy. Excess of urobilin should not be mistaken for bile pigment.

BLOOD AND BLOOD PIGMENT IN THE URINE

When hæmorrhage occurs in any part of the urinary tract, blood appears in the urine. It is found in the acute stage of Bright's disease. If a large quantity is present, the urine is deep red. Microscopic examination then reveals the presence of blood corpuscles, and on spectroscopic examination the bands of oxyhæmoglobin are seen.

If only a small quantity of blood is present, the secretion—especially if acid—has a characteristic reddish-brown colour, which physicians term 'smoky.'

The blood pigment may, under certain circumstances, appear in

the urine without the presence of any blood corpuscles at all. This is produced by a disintegration of the corpuscles occurring in the circulation, and the most frequent cause of this is a disease allied to ague, which is called *paroxysmal hæmoglobinuria*. The pigment is in the condition of methæmoglobin mixed with more or less oxyhæmoglobin, and the spectroscope is the means used for identifying these substances (see p. 107).

PUS IN THE URINE

Pus occurs in the urine as the result of suppuration in any part of the urinary tract. It forms a white sediment resembling that of phosphates, and, indeed, is always mixed with phosphates. The pus corpuscles may, however, be seen with the microscope; their nuclei are rendered evident by treatment with 1-per-cent. acetic acid, and the pus corpuscles are seen to resemble white blood corpuscles, which, in fact, they are in origin. Some of the proteid constituents of the pus cells—and the same is true for blood—pass into solution, so that the urine pipetted off from the surface of the deposit gives the tests for albumin. On the addition of liquor potassæ to the deposit of pus cells a ropy gelatinous mass is obtained. This is distinctive. Mucus treated in the same way is dissolved.

DETECTION OF PHYSIOLOGICAL PROXIMATE PRINCIPLES

Subsequent lessons may be very usefully employed by the class in testing for the various substances the properties of which they have studied. The following scheme will form a rough guide to the tests to be employed for the most important of the proximate principles:—

1. Note reaction, colour, clearness or opalescence, taste, smell. Coloured liquids suggest blood, bile, urine, &c. Opalescent liquids suggest starch, glycogen, or certain proteids.

2. Add iodine. A colour is produced:

If blue: **Starch.** Confirm by converting into a reducing sugar by saliva at 40° C., or boiling with dilute sulphuric acid.

If reddish brown: **Glycogen** or **dextrin.** Glycogen forms an opalescent solution in water, and is readily precipitated by alcohol. It is precipitated by basic lead acetate. Dextrin forms a clear solution: it is not precipitated by basic lead acetate unless ammonia is added also. It is not precipitated by alcohol unless a large excess is added. Both dextrin and glycogen are, like starch, convertible into a reducing sugar.

3. Add copper sulphate and caustic potash.

(*a*) Blue solution: boil; yellow or red precipitate. **Dextrose, levulose, maltose,** or **lactose** (for distinguishing tests see Lesson XIII.).

(*b*) Blue solution; no reduction on boiling; boil some of the original solution with 25-per-cent. sulphuric acid, and then boil with copper sulphate and caustic potash; abundant yellow or red precipitate: **Cane sugar.** Confirm by HCl test (see p. 13).

(*c*) Violet solution: **Proteids** (albumins, globulins, albuminates). In presence of magnesium sulphate the potash causes also a white precipitate of magnesia.

(*d*) Pink solution; biuret reaction. **Peptones** or **albumoses (proteoses).** In presence of ammonium sulphate very large excess of potash is necessary for this test. Only a trace of copper sulphate must be used.

4. When proteids are present proceed as follows: Boil the original solution (after adding a trace of 2-per-cent. acetic acid).

(*a*) Precipitate produced: **Albumins** or **globulins.**

(*b*) No precipitate: **Albuminates, proteoses,** or **peptones.**

5. If albumin, or globulin, or both are present, saturate a fresh portion with magnesium sulphate or half saturate with ammonium sulphate; filter; the precipitate contains the **globulin,** the filtrate the **albumin.** Test temperature of heat coagulation.

6. If proteids are present, but albumin or globulin absent:

(*a*) Neutralisation causes a precipitate soluble in excess of weak acid or alkali. **Acid albumin** or **alkali albumin**, according as the reaction of the original liquid is acid or alkaline respectively. If the original liquid is neutral, acid albumin and alkali albumin must be both absent.

(*b*) Neutralisation produces no such precipitate: **Proteose** or **peptone.**

7. If proteose, or peptone, or both are present, saturate a fresh portion with ammonium sulphate:

(*a*) Precipitate: **Proteose.** (*b*) No precipitate: **Peptone.**

If both are present, the precipitate contains the proteose, and the filtrate the peptone.

8. To a fresh portion add nitric acid (proteids having been proved to be present).

(*a*) No precipitate, even though excess of sodium chloride be also added: **Peptone.**

(*b*) No precipitate, until excess of sodium chloride is added: **Deutero-proteose.**

(*c*) Precipitate which disappears on heating and reappears on cooling: **Proteoses.** This is the distinctive test of all the proteoses or albumoses, and is given by all of them. For one of them, however (deutero-proteose), excess of sodium chloride must be added also.

(*d*) Precipitate little altered by heating: **Albumin** or **globulin.**

In all four cases nitric acid *plus* heat causes a yellow colour, turned orange by ammonia.

9. Confirmatory tests for proteids:—

(*a*) Millon's test.

(*b*) Adamkiewicz's reaction.

(*c*) Ferrocyanide of potassium and acetic acid cause a precipitate (except in the case of peptones and some proteoses).

(*d*) To test for fibrinogen:—

i. It coagulates by heat at 56° C.

ii. It is changed into fibrin by fibrin ferment and calcium chloride.

(*e*) To test for caseinogen:—

i. It is not coagulated by heat.

ii. It is changed into casein by rennet and calcium chloride.

10. If **blood** is suspected:

(*a*) Examine spectroscopically, diluting if necessary.

i. Oxyhæmoglobin shows two bands between D and E.

ii. Add ammonium sulphide; one band only appears.

iii. Carbonic oxide hæmoglobin shows two bands also, but will not reduce with ammonium sulphide.

iv. Methæmoglobin gives a typical band in the red between C and D.

v. Hæmatin &c. show special spectra (see Advanced Course).

(*b*) Dry: boil with glacial acetic acid and a crystal of sodium chloride on a glass slide under a cover glass. When cold, hæmin crystals are seen.

(*c*) If the blood is old and dry, and its hæmoglobin converted into hæmatin:

i. Try hæmin test.

ii. Dissolve it in potash; add ammonium sulphide, and examine for spectrum of hæmochromogen.

11. If **bile** is suspected:

(*a*) Try Gmelin's test for bile pigments.

(*b*) Try Pettenkofer's test for bile salts.

(*c*) Try Hay's sulphur test.

12. Miscellaneous substances.

(*a*) **Mucin.** Precipitated by acetic acid or by alcohol. The precipitate is soluble in lime water. By collecting the precipitate and boiling it with 25-per-cent. sulphuric acid, a reducing sugar-like substance is obtained. Mucin gives the proteid colour tests.

(*b*) **Nucleo-proteid.**—Precipitated by acetic acid or by alcohol. The precipitate is often viscous. It is soluble in dilute alkalis like 1-per-cent. sodium carbonate. This solution causes intravascular clotting. If the precipitate is collected and subjected to gastric digestion, an insoluble deposit of nuclein is left, which is rich in phosphorus. Nucleo-proteid gives the proteid colour tests.

(*c*) **Gelatin.** This also gives most of the proteid colour tests. It is not coagulated, but dissolved in hot water. The solution gelatinises when cold.

(*d*) **Urea.** Very soluble in water. The solution effervesces when sodium hypobromite or fuming nitric acid is added. Concentrate a fresh portion, add nitric acid, and examine for crystals of urea nitrate. Solid urea heated in a dry test-tube gives off ammonia, and the residue is called biuret. This gives a rose-red colour with copper sulphate and caustic potash.

(*e*) **Uric acid.** Very insoluble in water; soluble in potash, and precipitated from this solution in crystals by hydrochloric acid. Uric acid crystals from human urine are deeply pigmented red. Try murexide test.

(*f*) **Cholesterin.** Characteristic flat crystalline plates. For various colour tests see p. 70.

13. **Urine.** Normal constituents:

(*a*) Chlorides. Acidulate with nitric acid; add silver nitrate; white precipitate.

(*b*) Sulphates. Acidulate with nitric or hydrochloric acid: add barium chloride; white precipitate.

(*c*) Phosphates. Acidulate with nitric acid; add ammonium molybdate; boil; and a yellow crystalline precipitate forms. To another portion add ammonia; earthy (*i.e.* calcium and magnesium) phosphates are precipitated.

(*d*) Urea (see above).

(*e*) Uric acid. To 100 c.c. of urine add 5 c.c. of hydrochloric acid; leave for twenty-four hours, and pigmented crystals of uric acid are formed. For tests see above.

(*f*) Hippuric acid. Evaporate the urine with nitric acid, and heat the residue in a dry test-tube. A smell of oil of bitter almonds is given off.

(*g*) Creatinine. Take 100 c.c. of urine: add 5 c.c. of a saturated solution of sodium acetate and 20 c.c. of a saturated solution of mercuric chloride. Filter. Set the filtrate aside for twenty-four hours, and the spherical mercury compound of creatinine crystallises out. Examine this with the microscope.

For colour test with sodium nitro-prusside see p. 122.

14. **Urine.** Abnormal constituents.

(*a*) *Blood.* Microscope (blood corpuscles). Spectroscope (for oxyhæmoglobin or methæmoglobin). Hæmin test.

(*b*) *Blood pigment* may be present without blood corpuscles. Spectroscope.

(*c*) *Bile.* Gmelin's test. Hay's sulphur test.

(*d*) *Pus.* White deposit. Microscope (pus cells). Add potash; it becomes stringy.

(*e*) *Albumin.* (i.) Precipitated, if acid, by boiling; precipitate insoluble in acetic acid, so distinguishing it from phosphates. (ii.) Precipitated by nitric acid in the cold. (iii.) Precipitated by picric acid.

(*f*) *Sugar.* (i.) Brown colour with potash and heat (Moore's test). (ii.) Ferments with yeast. (iii.) Reduces Fehling's solution. (iv.) Urine has a high specific gravity. (v.) Add picric acid, potash, and boil; the urine becomes a dark opaque red; the similar slight coloration in normal urine is due to creatinine.

(*g*) *Mucus.* Flocculent cloud; may be increased by acetic acid; soluble in alkalis. A little mucus in urine is not abnormal.

(*h*) *Deposits.*

i. Examine microscopically for blood corpuscles, pus cells, crystals, &c.

ii. Phosphates. White deposit often mixed with mucus or pus. Insoluble on heating; soluble in acetic acid. Urine generally alkaline. Examine microscopically for coffin-lids of triple phosphate and star-like clusters of stellar (calcium) phosphate.

iii. Urates. Pink deposit, usually amorphous; may be mixed with envelope crystals of calcium oxalate. Deposit soluble on heating urine. Murexide test.

iv. Uric acid. Deposit like cayenne pepper. Microscope. Tests as above.

ADVANCED COURSE

INTRODUCTION

It will be presupposed that students who take the following lessons have already been through the elementary course. The order in which the subjects are treated is the same as that already adopted. The instructions given will be mainly practical; theoretical matter on which they depend, or to which they lead, is, as a rule, too lengthy to be discussed in a short manual like the present volume. The Appendix contains a description of various instruments which are not generally contained in sufficient numbers in a physiological laboratory to admit of each student being able to use them in a class. It also contains a description of certain methods of research which should always be shown in demonstrations, though there may be practical difficulties in allowing each member of the class to perform the experiments. The description of the phenomena of polarised light and their application has been considerably extended, and a section added on osmosis, a subject which is becoming of more and more importance to the student of physiology.

The few experiments in which living animals are employed will also necessarily be of the nature of demonstrations.

LESSON XIII

CARBOHYDRATES

1. **Glycogen.**—A rabbit which has been fed five or six hours previously on carrots is killed by bleeding. The chest and abdomen are opened quickly and a cannula inserted into the portal vein, and another into the vena cava inferior. A stream of salt solution is then allowed to pass through the liver until it is uniformly pale. The washings are collected in three beakers labelled *a*, *b*, and *c*.

The liver is cut out quickly, chopped into small pieces, and thrown into boiling water acidulated with acetic acid. The acidulated water extracts a small quantity of glycogen. The pieces of scalded liver are then ground up in a mortar with hot water, and thoroughly extracted with boiling water. Filter. A strong solution of glycogen is thus obtained.

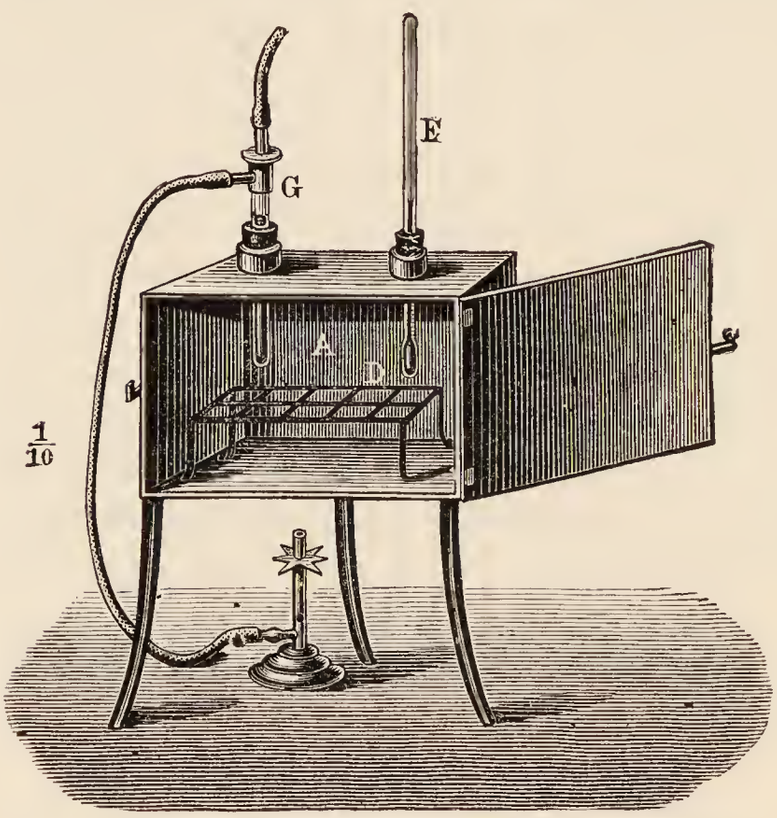


FIG. 55.—Hot-air oven with gas regulator (G). (Gscheidlen.)

Test the solution when cold with iodine.

To separate the glycogen evaporate the solution to a small bulk on the water-bath and add excess of alcohol; the glycogen is precipitated as a flocculent powder, which is collected on a filter and dried in an oven at the temperature of 100° (see fig. 55).

If the experiment is to be a quantitative one, the piece of liver taken and the glycogen obtained must be weighed.[1]

2. Examine the washings of the liver in the beakers *a*, *b*, and *c* for sugar. This may be done in a rough quantitative manner as follows:—Take equal

[1] This method of preparation of glycogen has the advantage that only traces of proteid are mixed with it. In Külz's method (extraction with dilute potash) there is more proteid. This is precipitated by the alternate addition of hydrochloric acid and potassio-mercuric iodide. Pavy and also Pflüger recommend extraction with strong potash, and subsequent precipitation with a certain percentage of alcohol; this method extracts all the glycogen easily.

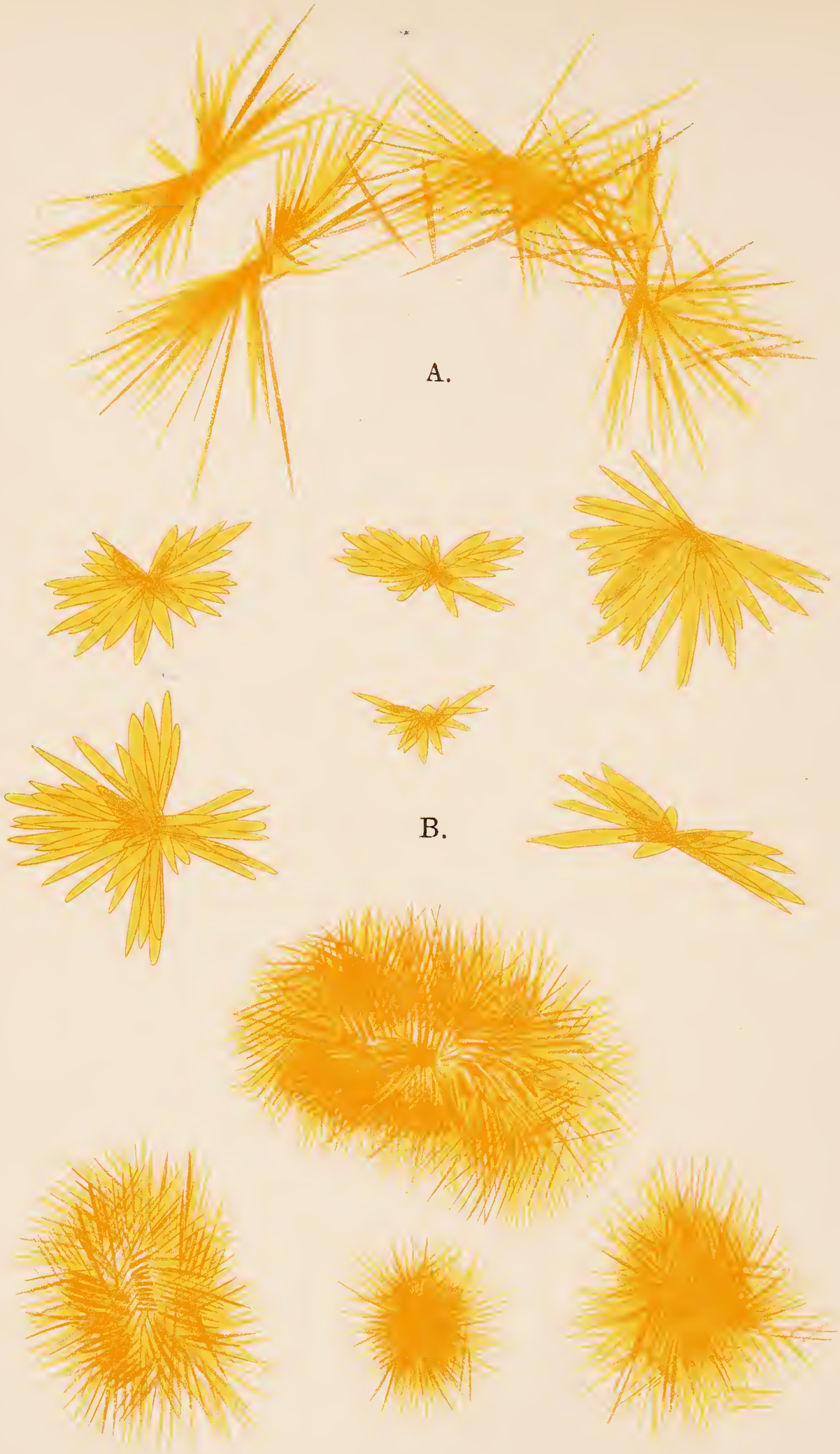


Fig. 56.—Plate of osazone crystals highly magnified.

A, phenyl-glucosazone. *B, phenyl-maltosazone.* *C, phenyl-lactosazone.*

quantities of *a*, *b*, and *c* in three test-tubes; to each add an equal amount of Fehling's solution, and boil: *a* will give a heavy precipitate, *b* one not so heavy, and *c* least of all, or none at all.

3. **Micro-chemical detection of Glycogen.**—A thin piece of the same liver is hardened in 90 per cent. alcohol. Sections are cut by the free hand, or after embedding in paraffin. If paraffin is used, this is got rid of by means of turpentine; and the sections prepared by either method are treated with chloroform in which iodine is dissolved, and mounted in chloroform balsam containing some iodine. The glycogen is stained brown, and is most abundant in the cells around the radicles of the hepatic vein.

4. **Phenyl-Hydrazine Test for Sugars.**—To 5 c.c. of the suspected fluid (*e.g.* diabetic urine) add 1 decigramme of phenyl-hydrazine hydrochloride, 2 decigrammes of sodium acetate, and heat on the water-bath at 100° C. for 30 to 60 minutes. On cooling, if not before, a crystalline or amorphous precipitate separates out. If amorphous, dissolve it in hot alcohol; dilute the solution with water, and boil to expel the alcohol, whereupon the osazone separates out in yellow crystals. Examine the crystals with the microscope (see accompanying plate).

Dextrose gives a precipitate of phenyl-glucosazone $C_6H_{10}O_4(N_2H.C_6H_5)_2$, which crystallises in yellow needles (melting-point 205° C.).

Levulose yields an osazone identical with this.

Galactose yields a very similar osazone (phenyl-galactosazone). It differs from phenyl-glucosazone by melting at 190–193°, and in being optically inactive when dissolved in glacial acetic acid.

Cane sugar does not form a compound with phenyl-hydrazine.

Lactose yields phenyl-lactosazone $C_{12}H_{20}O_9(N_2H.C_6H_5)_2$. It crystallises in needles, usually in clusters (melting-point 200° C.). It is soluble in 80–90 parts of boiling water. Lactose in urine does not give this test readily.

Maltose yields phenyl-maltosazone ($C_{24}H_{32}N_4O_9$). It crystallises in yellow needles much wider than those yielded by glucose or lactose (melting-point 206° C.). Unlike phenyl-glucosazone, it dissolves in 75 parts of boiling water, and is still more soluble in hot alcohol.

The chemistry of the phenyl-hydrazine reaction is represented in the following equations, dextrose being taken as an example of the sugar used:—

I. $$\underset{\text{[dextrose]}}{CH_2OH[CH(OH)]_3CH(OH)COH} + \underset{\text{[phenyl-hydrazine]}}{H_2N.NH(C_6H_5)} = \underset{\text{[hydrazone]}}{CH_2OH[CH(OH)]_3CH(OH)C(=N-NH(C_6H_5))} + \underset{\text{[water]}}{H_2O}$$

II. $$\underset{\text{[hydrazone]}}{CH_2OH[CH(OH)]_3CH(OH)C(=N-NH(C_6H_5))} + \underset{\text{[phenyl-hydrazine]}}{C_6H_5.NH-NH_2} = \underset{\text{[osazone]}}{CH_2OH[CH(OH)]_3C(=N-C_6H_5.NH)-C(=N-NH.C_6H_5)} + \underset{\text{[hydrogen]}}{H_2} + \underset{\text{[water]}}{H_2O}$$

5. **Barfoed's Reagent.**—Dissolve 1 part of cupric acetate in 15 parts of water; to 200 c.c. of this solution add 5 c.c. of acetic acid containing 38 per cent. of glacial acetic acid. Dextrose reduces this reagent on boiling; maltose and lactose do not. This test is not very delicate, and the reagent must be freshly prepared.

6. **The Polarimeter.**—Estimate the strength of a solution of dextrose by means of the polarimeter (see Appendix).

7. **Formation of Mucic Acid.**—Take 1 gramme of lactose and heat it in a porcelain capsule with 12 c.c. of nitric acid on a water-bath until the fluid is reduced to one-third of its original volume. A precipitate of mucic acid separates out. Cane sugar, maltose, dextrose, dextrin, and starch treated in the same way yield an isomeric acid called saccharic acid, which, being soluble, does not separate out. Lactose yields both acids; galactose mucic acid only.

8. **Pentoses** give the ordinary reduction tests for sugar and yield osazones, but do not ferment with yeast. They give the two following characteristic tests; they may be performed with gum arabic (which contains arabinose) or pine-wood shavings (which contain xylose).

(*a*) Phloroglucin reaction. Warm some distilled water with an equal volume of concentrated hydrochloric acid in a test-tube and add phloroglucin until a little remains undissolved. Add a small quantity of gum arabic, and keep the mixture warm in the water-bath at 100° C. The solution becomes cherry-red, and a precipitate settles out, which is soluble in amyl alcohol. This solution gives an absorption band between the D and E lines.

(*b*) Orcin reaction. Substitute orcin for phloroglucin in the foregoing experiment. The solution becomes violet on warming, then blue, red, and finally green. A bluish green precipitate settles out, soluble in amyl alcohol. This solution gives an absorption band between C and D.

9. **Glycuronic acid** gives all the above reactions; it may be distinguished as follows:—

Take 50 c.c. of glycuronic acid solution in a capsule; add 1 gramme of *p*-bromphenyl-hydrazine and rather more than the same amount of sodium acetate. Keep the mixture in the water-bath at 100° C. for a quarter of an hour, when yellow crystals of *p*-bromphenyl-hydrazone separate out. After cooling filter off the crystals and wash them with absolute alcohol, in which they are insoluble. Under the same conditions carbohydrates yield *p*-bromphenyl-osazones, but these are soluble in absolute alcohol. The *p*-bromphenyl-hydrazone is soluble in absolute alcohol to which pyridine has been added; the rotatory power of this solution is greater than that of any of the osazones.

LESSON XIV

CARBOHYDRATES : ACTION OF MALT UPON STARCH

1. Prepare a 0·5-per-cent. solution of starch.

2. Prepare some malt extract by digesting 10 grammes of powdered malt with 50 c.c. of water at 50° C. for three hours, and subsequently straining. This extract contains the diastatic or malting ferment.

Solutions 1 and 2 may be conveniently prepared beforehand by the demonstrator.

3. To the starch solution add one-tenth of its volume of malt extract, and place the mixture in a water-bath at 40° C. From time to time test portions of the liquid by mixing a drop with a drop of iodine solution on a testing slab. The blue colour at first seen is soon replaced by a violet (mixture of blue and red), and then by a red reaction (due to *erythrodextrin*) which gradually vanishes. Alcohol added to the liquid when all starch and erythrodextrin have gone still causes a precipitate of a dextrin, which, as it gives no colour with iodine, is called *achroö-dextrin*. The liquid also contains a reducing sugar, maltose.

4. Take 50 c.c. of a solution of maltose and determine how much of it is necessary to reduce 10 c.c. of Fehling's solution.

5. Take another 50 c.c. and boil it with 1 c.c. of strong sulphuric acid for half an hour in a flask. This converts it into dextrose. After cooling bring the liquid to its original volume (50 c.c.) by adding water, and again determine its increased reducing power with Fehling's solution. If x = c.c. of maltose solution necessary to reduce 10 c.c. of Fehling's solution, then $\frac{2x}{3}$ = c.c. of dextrose solution necessary for the same purpose. The strength of the maltose solution can be calculated from the fact that 10 c.c. of Fehling's solution correspond to 0·05 gramme of dextrose.

LESSON XV

CRYSTALLISATION OF EGG ALBUMIN

Fresh egg-white is mixed with an equal bulk of fully saturated, filtered, neutral ammonium sulphate solution. 100 c.c. of the former are measured into a porcelain basin or strong beaker, and 100 c.c. of ammonium sulphate solution are added in successive quantities of 10 or 15 c.c., the mixture being thoroughly churned with an egg whisk after each addition. The whole should be finally so thoroughly beaten up as to form a large proportion of light froth. After the greater part of the froth has broken down the mixture is thrown on a folded filter-paper, moderately rapid filtration being obtained without the use of a filter-pump. The filtrate is strongly alkaline to litmus, and smells of ammonia. To the filtrate in a flask, or to as much of it as can be obtained in a convenient time of filtration, further ammonium sulphate solution is very cautiously added (best, drop by drop from a burette) until a slight permanent precipitate remains, and this precipitate is afterwards just redissolved by the equally cautious addition of water. Dilute acetic acid (10 per cent.) from a burette is now added drop by drop until such a stage of reaction is reached that a precipitate forms and only just redissolves. Finally one or two drops (not more) of acid are added in excess of this, whereupon a bulky white precipitate falls. The flask is now corked and allowed to stand. In 24 hours or less the precipitate, which will have increased in quantity, will be found to consist entirely of acicular crystals. Small portions should be examined under a $\frac{1}{6}$th objective, avoiding pressure on the cover slip. (F. G. Hopkins.)

LESSON XVI

MILK

1. Caseinogen in milk exists in the form of a salt (calcium caseinogenate). Add acetic acid to milk, and this salt is decomposed, and free caseinogen (with entangled fat) is precipitated. Collect the precipitate so produced from a pint of milk in a filter, and wash thoroughly with distilled water: grind it up with calcium carbonate in a mortar, and add about a pint of distilled water; allow the mixture to stand for about an hour. The fat rises to the top: the excess of calcium carbonate falls to the bottom. The intermediate fluid contains the caseinogen in solution; it is usually very opalescent. Take some of this solution and divide it into three parts, A, B, and C.

To A add rennet.

To B add a few drops of 2-per-cent. solution of calcium chloride.

To C add both rennet and calcium chloride.

Put all three in the water-bath at 40° C. A clot of casein forms in C, but not in A if all calcium salts have been successfully washed away, nor in B.

2. The formation of casein from caseinogen is a double process; the first action is that of the ferment, which converts the caseinogen into what may be called soluble casein; the second action is that of the calcium salt, which precipitates the casein in an insoluble form, or curd. This may be shown by taking some of the caseinogen solution and adding rennet. Warm to 40° C.; no visible change occurs, but nevertheless soluble casein and not caseinogen is now present. Then boil this mixture to destroy the rennet, cool, and add calcium chloride. A formation of insoluble curd now occurs.

3. Caseinogen may be precipitated as a salt from milk by the addition of alcohol. This reagent also precipitates the other milk proteids.

4. The method of salting out described in Lesson VI., Exercise 10 (p. 41), may also be used. Add to some milk an equal volume of saturated solution of ammonium sulphate. Caseinogen as a salt is thus precipitated, and entangles the fat with it. Filter off the precipitate and examine the filtrate as follows:—Saturate it with sodium chloride; a small amount of precipitate comes down. This is the so-called lacto-globulin. This contains only a trace of true globulin; it is mostly caseinogen previously left in solution together with calcium sulphate. Filter it off; acidify the filtrate with a few drops of 2-per-cent. acetic acid and heat it in a water-bath gradually. About 77° C. the remaining proteid (lactalbumin) is coagulated.

LESSON XVII

THE ALBUMOSES

1. Witte's peptone contains very little true peptone, but consists chiefly of albumoses, which are soluble like peptone, in neutral saline solutions.

2. Make a solution of this substance in 10-per-cent. sodium chloride solution, and filter. Very little residue is left on the filter. This consists of dysalbumose, an insoluble form of hetero-albumose, formed during the process of preparing the substance. If hot saline solution is used instead of cold as a solvent, this amount of insoluble residue is increased, hetero-albumose being to a slight extent precipitated by heat.

3. The solution gives the following tests :—

(*a*) It does not coagulate on heating.

(*b*) Biuret reaction (due both to peptone and albumoses).

(*c*) A drop of nitric acid, best added by a glass rod, gives a precipitate which dissolves upon heating and reappears on cooling. (This is due to the albumoses present.)

The precipitate produced by the addition of acetic acid and a drop of potassium ferrocyanide is also soluble on heating, and reappears on cooling.

4. For the separation of the albumoses and peptone proceed as follows :—

(*a*) Saturate the solution with ammonium sulphate, and filter. The filtrate contains the peptone, and the precipitate the albumoses. The peptone is not precipitated by nitric acid, nor by most of the reagents that precipitate other proteids. It is precipitated completely by alcohol, tannin, and potassio-mercuric iodide ; imperfectly by phospho-tungstic and phospho-molybdic acid.

It gives the biuret reaction, *but in the presence of ammonium sulphate a large excess of caustic potash is necessary.*

(*b*) Dialyse another portion of the solution; hetero-albumose is precipitated.

(*c*) Saturate another portion of the solution with sodium chloride (or half saturate with ammonium sulphate) after faintly acidulating with acetic acid. Proto-albumose and hetero-albumose are precipitated. Filter. The filtrate contains the deutero-albumose and peptone.

The proto- and hetero-albumose may be redissolved by adding distilled water, and may be separated from each other by dialysis (see *b*).

Deutero-albumose may be separated from the peptone by saturation with ammonium sulphate, or by the addition of a crystal of phosphoric acid. These reagents precipitate the deutero-albumose, but not the peptone.

Deutero-albumose gives the nitric acid reaction (see 3, *b*) characteristic of the albumoses only in the presence of excess of salt. If the salt is removed by dialysis, nitric acid then causes no precipitate.

5. Among the important reactions of proteids is Piotrowski's reaction—that is, the coloration produced by copper sulphate and a caustic alkali; the term 'biuret reaction' is applied to the rose-red colour which proteoses and peptones give with these reagents, because biuret (a derivative of urea) gives a similar colour. It does not, however, prove that biuret is contained in the proteid molecule. Biuret and proteid both contain some radicle to which the colour is due. Gnezda thought it was cyanogen, and that the cyanogen was differently combined in the peptones and native proteids (albumins and globulins) respectively; hence the rose-red given by one group and the violet by the other. More recent work by Pickering, however, points to a CONH group rather than cyanogen. Gnezda found that if a dilute solution of nickel sulphate is used instead of copper sulphate, the native proteids give different colours from the peptones and proteoses, and Pickering has found the same with cobalt. Their results may be given in the following table:—

Proteid	Copper sulphate and ammonia	Copper sulphate and potash	Nickel sulphate and ammonia	Nickel sulphate and potash	Cobalt sulphate and ammonia	Cobalt sulphate and potash
Albumins and globulins	Blue	Violet	Nil	Yellow	Nil	Heliotrope-purple
Proteoses and peptones	Rose-red	Rose-red	Yellow	Orange	Nil	Red brown

6. Another delicate test introduced by McWilliam may here be mentioned. Salicyl-sulphonic acid precipitates albumins and globulins: on heating the precipitate is coagulated. The same reagent precipitates proteoses. On heating the precipitate dissolves and reappears on cooling. It does not precipitate peptones.

7. The use of trichloracetic acid for the separation of various proteids may be illustrated by the following experiment. Take some blood and add to it some solution of Witte's peptone (*i.e.* proteoses and peptone). Add to this mixture an equal volume of a 10-per-cent. solution of trichloracetic acid. There is an abundant precipitate. Boil rapidly and filter hot. The filtrate contains the proteoses and peptone, all the other proteids being contained in the precipitate. On cooling, the filtrate deposits some of the proteose. The proteose and peptone may be detected in the usual way.

LESSON XVIII

DIGESTION

1. **Activity of Pepsin Solutions (Grützner's Method).**—Examine the comparative digestive power of the glycerin extracts of two stomachs. Take, in two test-tubes, an equal small weighed quantity of fibrin stained with carmine. Add to each 10 c.c. of 0·2-per-cent. hydrochloric acid. Add to one a measured quantity of one glycerin extract, and to the other an equal quantity of the other glycerin extract. As the fibrin is digested the carmine is set free, and colours the liquid; that which is more deeply stained is that which contains the more active preparation of pepsin. In the original method the amount of carmine set free is estimated by an artificial scale consisting of ten solutions of carmine of different known strengths.

The carmine solution for staining the fibrin is prepared by dissolving 1 gramme of carmine in about 1 c.c. of ammonia; to this 400 c.c. of water are added, and the mixture is kept in a loosely stoppered bottle till the smell of ammonia has become faint.

The fibrin is stained by taking it perfectly fresh and clean. It is chopped finely and placed in the carmine solution for twenty-four hours. The fluid is strained off and the fibrin washed in water till the washings are colourless. It is kept in a stoppered bottle with just enough ether to cover it.

2. **Mett's Tubes.**—A method which is now more generally employed for estimating the proteolytic activity of a digestive juice is one originally introduced by Mett. Pieces of capillary glass tubing of known length are filled with white of egg. This is set into a solid by heating to 95° C. They are then placed in the digestive fluid at 36° C., and the coagulated egg-white is digested. After a given time the tubes are removed; and if the digestive process has not gone too far, only a part of the little column of coagulated proteid will have disappeared; the length of the remaining column is easily measured, and the length that has been digested is a measure of the digestive strength of the fluid.[1]

3. **The Acid of Gastric Juice.**—The digestive powers of the acids are pro-

[1] Hamburger has used the same method in investigating the digestive action of juices on gelatin. The tubes are filled with warm gelatin solution, and this jellies on cooling. They are placed as before in the digestive mixture, and the length of the column that disappears can be easily measured. These experiments must, however, be performed at room temperature, for the usual temperature (36°–40° C.) at which artificial digestion is usually carried out would melt the gelatin. He has also used the same method for estimating amylolytic activity, by filling the tubes with thick starch paste.

portional to their dissociation and the number of H ions liberated. The anions, however, modify this by having different powers of retarding the action. The greater suitability of hydrochloric over lactic acid, for instance, in gastric digestion is due to the fact that the former acid more readily undergoes dissociation.

Hydrochloric acid is absent in some diseases of the stomach; the best colour tests for it are the following:—

(*a*) Gunsberg's reagent consists of 2 parts of phloroglucinol, 1 part of vanillin, and 30 parts of rectified spirit. A drop of filtered gastric juice is evaporated with an equal quantity of the reagent. Red crystals form, or, if much peptone is present, there will be a red paste. The reaction takes place with one part of hydrochloric acid in 10,000. The organic acids do not give the reaction.

(*b*) Tropæolin test. Drops of a saturated solution of tropæolin -OO in 94 per cent. methylated spirit are allowed to dry on a porcelain slab at 40° C. A drop of the fluid to be tested is placed on the tropæolin drop, still at 40° C.; and if hydrochloric acid is present, a violet spot is left when the fluid has evaporated. A drop of 0·006-per-cent. hydrochloric acid leaves a distinct mark.

(*c*) Töpfer's test. A drop of dimethyl-amido-azo-benzol is spread in a thin film on a white plate. A drop of dilute hydrochloric acid (up to 1 in 10,000) strikes with this in the cold a bright red colour.

Lactic acid is sometimes present in the gastric contents, being derived by fermentative processes from the food. It is soluble in ether, and is generally detected by making an ethereal extract of the stomach contents, and evaporating the ether. If lactic acid is present in the residue it may be identified by Uffelmann's reaction in the following way:—

A solution of dilute ferric chloride and carbolic acid is made as follows:—

10 c.c. of a 4-per-cent. solution of carbolic acid.

20 c.c. of distilled water.

1 drop of the liquor ferri perchloridi of the British Pharmacopœia.

On mixing a solution containing a mere trace (up to 1 part in 10,000) of lactic acid with this violet solution, it is instantly turned yellow. Larger percentages of other acids—for instance, more than 0·2 per cent. of hydrochloric acid—are necessary to decolorise the test solution.

4. **Demonstration of Pancreatic Secretion.**—In an anæsthetised dog insert a cannula into the main pancreatic duct, and collect the juice in a suitable vessel. Inject some 0·4-per-cent. hydrochloric acid into the duodenum, and note after some minutes the abundant flow of pancreatic juice. Next ligature off and remove two or three feet of the upper part of the small intestine, wash out the contents and slit it open; scrape off the mucous membrane with the back of a scalpel; preserve a small quantity of the scrapings for future use and label this A. Grind up the remainder in a mortar with clean sand or powdered glass, and add 0·4 per cent. hydrochloric acid. Transfer the mixture to a flask, boil thoroughly and when cool neutralise with a little caustic soda solution. Filter; the filtrate contains *secretin*, which has been formed by the acid from the *pro-secretin* of the intestinal epithelium. Inject some of this solution through a cannula into

the external jugular vein of the dog, and an abundant flow of pancreatic juice is an almost immediate result.

5. Characters of the juice so obtained:—

(*a*) It is a clear, colourless fluid, and very strongly alkaline.

(*b*) Mixed with starch solution and kept at 40° C. dextrin and maltose are rapidly formed.

(*c*) Mixed with milk there may be some curdling produced, but the most marked effect is that the milk rapidly becomes acid, and a smell of fatty acids is noticeable.

(*d*) Added to fibrin and kept at 40° proteid digestion occurs very slowly; next day, however, the fibrin will be in large measure digested.

(*e*) Mix some of the pancreatic juice with the scraping of the intestine which was preserved and labelled A. Then add fibrin. The fluid is now strongly proteolytic and at 40° C. the fibrin rapidly dissolves; trypsin has been liberated from the trypsinogen of the juice by the intestinal *entero-kinase*.

6. **Products of Pancreatic Digestion of Proteids.**—A finely divided ox-pancreas should be allowed to digest at 40° C. for twenty-four to thirty-six hours in a litre of 1-per-cent. solution of sodium carbonate to which the white of an egg has been added every ten hours. Unless thymol or some other suitable antiseptic has been added to prevent putrefaction, the odour of putrefaction will be very evident. This should be got ready by the demonstrator beforehand.

Instead of the above, it is on the whole more convenient to digest a large quantity of fibrin with artificial pancreatic juice, if the natural juice obtained by the injection of secretin is not available; in the latter case the addition of intestinal epithelium (entero-kinase) should not be forgotten. The products of digestion in one case should be examined, say, after six hours' digestion, and in another case after thirty-six hours' digestion or more. The digestion products should then be searched for, the early products of digestion (alkali-albumin, deutero-proteose, &c.) will become less abundant with the length of time that digestion has been allowed to progress, and the latter products (peptone, leucine, tyrosine, tryptophan) will become more abundant. The methods for testing most of these substances have been already given. The following are the tests for tryptophan, leucine, and tyrosine:—

(*a*) *Tryptophan.*—Add a few drops of bromine water; a violet colour is produced.

(*b*) *Leucine and Tyrosine.*—i. Examine microscopical specimens of these. The deposit generally found in rather old specimens of Benger's liquor pancreaticus will be a convenient source of these crystals.

ii. To some of the pancreatic digest add Millon's reagent and filter off the precipitated proteid. Boil the filtrate, and the presence of tyrosine is indicated by a red colour. If tyrosine is abundant the red colour appears without boiling. Leucine does not give this test.

iii. Faintly acidify another portion of the filtered digest with acetic acid and boil; if any proteid matter is still undigested it will be thus coagulated and can be filtered off. Reduce the filtrate to a small bulk until it begins to become syrupy. Leave overnight in a cool place, and crystals mainly of

tyrosine will separate out. Filter these off through fine muslin, and evaporate down the filtrate to the consistency of a thick syrup; leave this overnight again, and a second crop of crystals, forming a skin on the surface and consisting mainly of leucine, will have separated out.

7. **Zymogen Granules.**—Examine microscopically, mounting in aqueous humour or serum (or in glycerin after treatment with osmic acid vapour), small pieces of the pancreas, parotid, and submaxillary glands in a normal guinea-pig [1] and also in one in which profuse secretion had been produced by the administration of pilocarpine.

Note that zymogen granules are abundant in the former, and scarce in the latter, being situated chiefly at the free border of the cells.

Extremely good, though not permanent, microscopic specimens may be obtained by teasing in a 33-per-cent. solution of caustic potash.

[1] The guinea-pigs should be killed by bleeding, and the blood collected and defibrinated, and utilised for the preparation of oxyhæmoglobin crystals. This will give students an opportunity of seeing the exceptional form (tetrahedra) in which the blood-pigment of this animal crystallises.

The three methods of obtaining crystals described on p. 100 all give good results. If amyl nitrite is used instead of ether in the third method, crystals of methæmoglobin are obtained.

LESSON XIX

HÆMOGLOBIN AND ITS DERIVATIVES

Defibrinated ox-blood suitably diluted may be used in the following experiments as in those described in Lesson IX.

1. Place some in a hæmatoscope (see fig. 33, p. 104) in front of the large spectroscope. Note the position of the two characteristic bands of **oxyhæmoglobin**; these are replaced by the single band of **hæmoglobin** after reduction by the addition of Stokes's reagent (see footnote, p. 103) or ammonium sulphide. By means of a small rectangular prism a comparison spectrum showing the bright sodium line (in the position of the dark line named D in the solar spectrum) may be obtained, and focussed with the absorption spectrum.

2. Obtain similar comparison spectra by the use of the microspectroscope. For this purpose a cell containing a small quantity of oxyhæmoglobin solution may be placed on the microscope stage, and a test-tube containing **carbonic oxide hæmoglobin** in front of the slit in the side of the instrument. Notice that the two bands of carbonic oxide hæmoglobin are very like those of oxyhæmoglobin, but are a little nearer to the violet end of the spectrum.

Carbonic oxide hæmoglobin may be readily prepared by passing a stream of coal gas through the diluted blood. It has a cherry-red colour, and is not reduced by the addition of ammonium sulphide (fig. 57, spectrum 4).

3. **Methæmoglobin.**—Add a few drops of ferricyanide of potassium to dilute blood and warm gently. The colour changes to mahogany-brown. Place the test-tube in front of the small direct-vision spectroscope. Note the characteristic band in the red (fig. 57, spectrum 5). On dilution other bands appear (fig. 57, spectrum 6). Treat with ammonium sulphide, and the band of hæmoglobin appears.

4. **Acid Hæmatin.**—(*a*) Prepare the following mixture:—150 c.c. of 90-per-cent. alcohol and 6 c.c. of concentrated sulphuric acid; take about 5 c.c. of this mixture and boil it in a test-tube. While still hot drop into it a few drops of undiluted defibrinated blood, and filter. Note the brown colour of the filtrate. Compare the position of the absorption band in the red with that of methæmoglobin; that of acid hæmatin is further from the D line (fig. 57, spectrum 7).

(*b*) Add some glacial acetic acid to undiluted defibrinated blood. Extract this with ether by gently agitating it with that fluid. The ethereal extract should then be poured off and examined spectroscopically. The band in the red is seen, and on further diluting with ether three additional bands appear.

5. **Alkaline Hæmatin.**—(*a*) Add to diluted blood a small quantity of strong

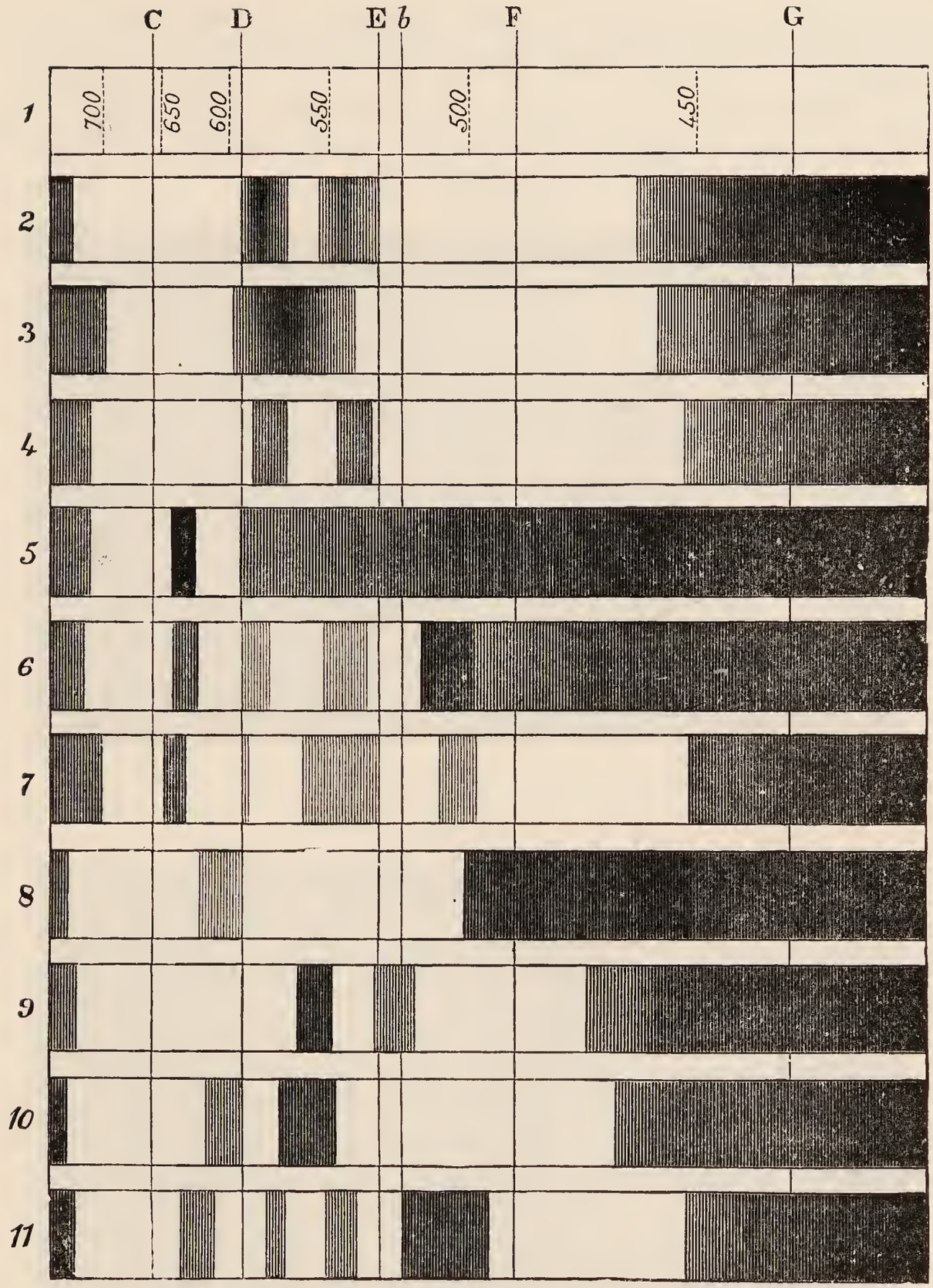


FIG. 57.—1, Solar spectrum. 2, Spectrum of oxyhæmoglobin (0·37 p.c. solution). First band, λ 589–564 ; second band, λ 555–517. 3, Spectrum of hæmoglobin. Band, λ 597–535. 4, Spectrum of CO-hæmoglobin. First band, λ 583–564 ; second band, λ 547–521. 5, Spectrum of methæmoglobin (concentrated solution). 6, Spectrum of methæmoglobin (dilute solution). First band, λ 647–622 ; second band, λ 587–571 ; third band, λ 552–532 ; fourth band, λ 514–490. 7, Spectrum of acid hæmatin (ethereal solution). First band, λ 656–615 ; second band, λ 597–577 ; third band, λ 557–529 ; fourth band, λ 517–488. 8, Spectrum of alkaline hæmatin. Band from λ 630–581. 9, Spectrum of hæmochromogen (reduced hæmatin). First band, λ 569–542 ; second band, λ 535–504. 10, Spectrum of acid hæmatoporphyrin. First band, λ 607–593 ; second band, λ 585–536. 11, Spectrum of alkaline hæmatoporphyrin. First band, λ 633–612 ; second band, λ 589–564 ; third band, λ 549–529 ; fourth band, λ 518–488. The above measurements (after MacMunn) are in millionths of a millimetre. The liquid was examined in a layer 1 centimetre thick. The edges of ill-defined bands vary a good deal with the concentration of the solutions.

caustic potash and boil. The colour changes to brown, and with the spectroscope a faint shading on the left side of the D line is seen (fig. 57, spectrum 8).

(*b*) The band is much better seen in an alcoholic solution. Prepare the following mixture:—150 c.c. of 90-per-cent. alcohol, and 18 c.c. of 50-per-

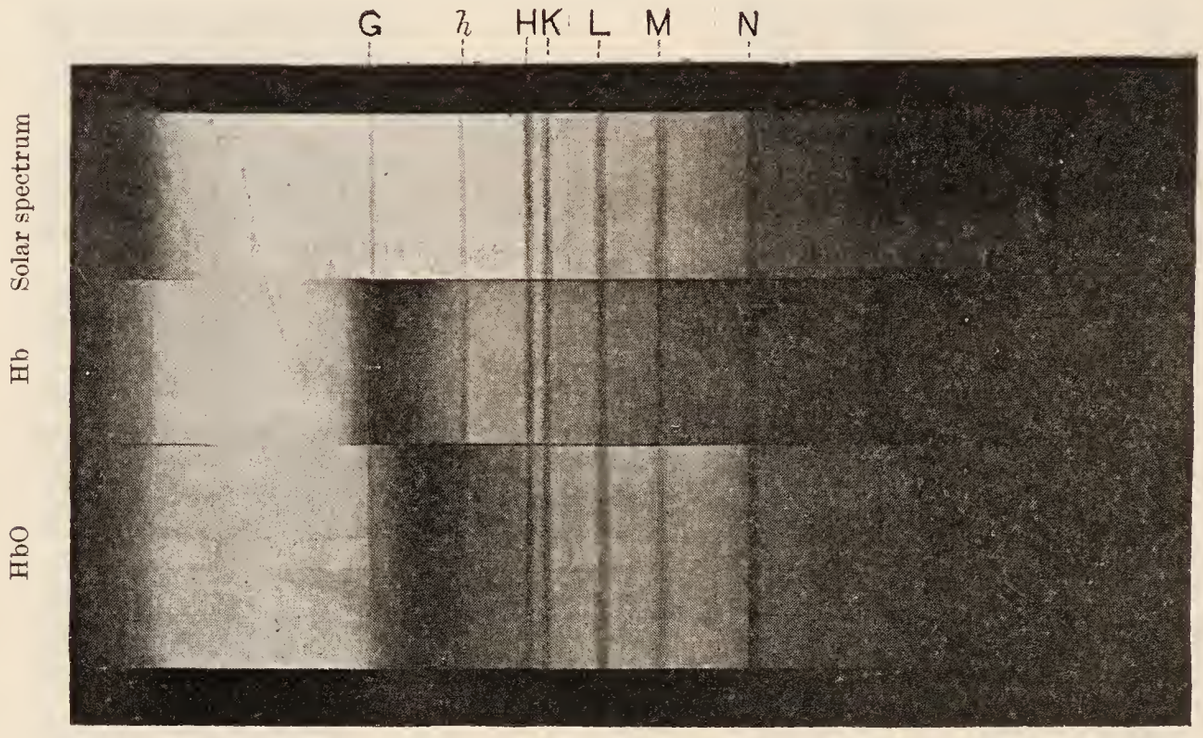


FIG. 58.—The photographic spectrum of hæmoglobin and oxyhæmoglobin. (Gamgee.)

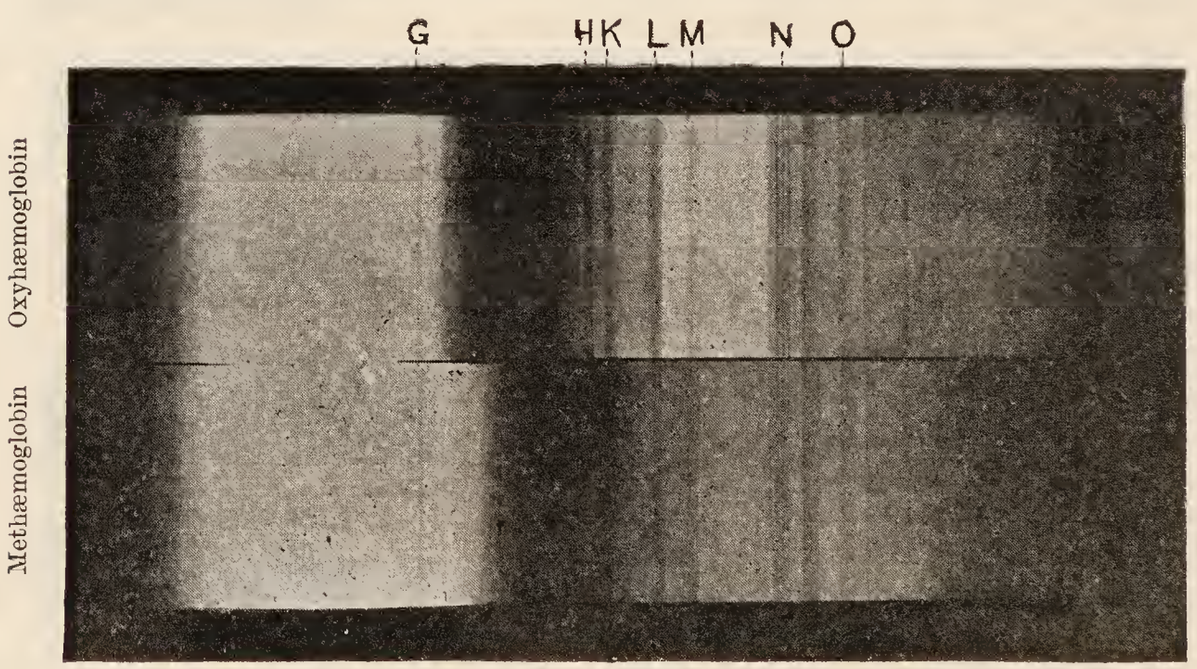


FIG. 59.—The photographic spectrum of oxyhæmoglobin and methæmoglobin. (Gamgee.)

cent. potash. Take about 5 c.c. of this mixture in a test-tube and boil it. While still hot drop into it a few drops of undiluted defibrinated blood. The fluid then shows the spectrum of alkaline hæmatin. This may then be used for the next experiment.

6. **Hæmochromogen.**—Add ammonium sulphide to the solution of alkaline hæmatin; the colour changes to red, and two bands are seen, one between D

and E, and the other nearly coinciding with E and *b* (fig. 57, spectrum 9). The spectrum of alkaline hæmatin reappears for a short time after vigorous shaking with air.

7. **Hæmatoporphyrin.**—To some strong sulphuric acid in a test-tube add a few drops of undiluted blood, and observe the spectrum of acid hæmatoporphyrin (iron-free hæmatin) (fig. 57, spectrum 10). Map out all the spectra you see on a chart.

8. **The Photographic Spectrum.**—Hæmoglobin and its compounds also show absorption bands in the ultra-violet portion of the spectrum. This portion of the spectrum is not visible to the eye, but can be rendered visible by allowing the spectrum to fall on a fluorescent screen, or on a sensitive photographic plate. In order to show absorption bands in this part of the spectrum very dilute solutions of the pigment must be used.

In order to demonstrate these bands, the telescope of a large spectroscope is removed, and a beam of sunlight or of light from the positive pole of an arc lamp is allowed to fall on the slit of the collimator. The spectrum is focussed on a fluorescent screen.[1] The slit is then opened very widely, and the coloured solution is interposed on the path of the beam falling on the slit.

Oxyhæmoglobin shows a band (Soret's band) between the lines G and H. In hæmoglobin, carbonic oxide hæmoglobin, and nitric oxide hæmoglobin, this band is rather nearer G. Methæmoglobin and hæmatoporphyrin show similar bands.

The two preceding figures show the 'photographic spectra' of hæmoglobin, oxyhæmoglobin, and methæmoglobin, and will serve as examples of the results obtained. I am greatly indebted to Prof. Gamgee, to whom we owe most of our knowledge on this subject, for permission to reproduce these two specimens of his numerous photographs.

9. **Preparation of Pure Oxyhæmoglobin.**—The following method is described in Stirling's 'Practical Physiology.' Centrifugalise dog's defibrinated blood and pour off the serum. Centrifugalise again with physiological saline solution repeatedly until the supernatant fluid contains only traces of proteid. Mix the magma of corpuscles with two or three volumes of water saturated with acid-free ether; the solution becomes clear. Then add a few drops of 1-per-cent. solution of acid sodium sulphate till the mixture looks tinted like fresh blood, owing to the precipitation of the stromata. These can be separated by centrifugalising. (I have found that they aggregate together and can be easily removed by filtration.) Pour off the clear red fluid: cool it to 0° C., add one-fourth of its volume of absolute alcohol previously cooled to 0° C. Shake well, and then let the mixture stand at 5°–15° C. for 24 hours. As a rule the whole passes into a glittering crystalline mass. Filter at 0° C. and wash with ice-cold 25-per-cent. alcohol. Redissolve the crystals in a small quantity of water, and recrystallise as before. The crystals may then be spread on plates of porous porcelain, and dried in a vacuum over sulphuric acid.

[1] Fluorescent screens, similar to those in common use in observations made with Röntgen rays, may be made by coating white cardboard with barium platinocyanide.

LESSON XX

SERUM

1. The following methods of precipitating serum globulin should be performed :—

(*a*) **Panum's Method.**—Dilute serum with fifteen times its bulk of water. It becomes cloudy owing to partial precipitation of the serum globulin. Add a few drops of 2-per-cent. acetic acid ; the precipitate becomes more abundant, and it dissolves in excess of the acid. It was formerly called 'serum casein.'

(*b*) **Alexander Schmidt's Method.**—Dilute serum with twenty times its bulk of water and pass a stream of carbonic acid through it. A fairly abundant precipitate of serum globulin falls. Let it settle and an additional precipitate can be obtained from the decanted liquid by treating it with a trace of acetic acid (the 'serum casein' mentioned above). Repeat the carbonic acid method without dilution ; no precipitate forms.

(*c*) **By Dialysis.**—Put some serum in a dialyser with distilled water in the outer vessel. The water must be frequently changed. In order to prevent decomposition a few crystals of thymol are added. In a day or two the salts have passed out ; the proteids remain behind : of these the serum albumin is still in solution ; the serum globulin is in part precipitated, as it requires a small quantity of salt to hold it in solution.

(*d*) **By Addition of Salts :—**

(i.) Schmidt's method. Saturate some serum with sodium chloride. A precipitate of serum globulin is produced.

(ii.) Hammarsten's method. Use magnesium sulphate instead of sodium chloride. A more abundant precipitate is produced, because this salt is a more perfect precipitant of serum globulin than sodium chloride. In order to obtain complete saturation with these salts it is necessary to shake the mixture of salt and serum for some hours.[1]

(iii.) Kauder's method. Half saturate serum with ammonium sulphate. This is done by adding to the serum an equal volume of saturated solution of ammonium sulphate. This precipitates the globulin. Complete saturation with the salt precipitates the albumin also.

2. **Heat Coagulation.**—Saturate serum with magnesium sulphate and filter off the precipitate ; preserve the filtrate and label it 'B.' Wash the precipitate on the filter with saturated solution of magnesium sulphate until the washings do not give the tests for albumin,[2] then dissolve the precipitate

[1] This may be conveniently done by a shaking machine before the class meets.

[2] On account of the prolonged nature of these operations, they must necessarily be performed by the demonstrator beforehand

by adding distilled water. It readily dissolves, owing to the salt adherent to it. The solution is opalescent. Label it 'A.'

Render A faintly acid with a drop of 2-per-cent. acetic acid, and heat in a test-tube. The temperature of the test-tube may be raised by placing it in a flask of water gradually heated over a flame. A thermometer is placed in the test-tube, and should be kept moving so as to ensure that all parts of the liquid are at the same temperature. The quantity of liquid in the test-tube should be just sufficient to cover the bulb of the thermometer. A flocculent precipitate of coagulated serum globulin separates out at about 75°.

Now take the filtrate B. This contains the serum albumin. Dilute it with an equal volume of water; render it faintly acid as before, testing the reaction with litmus paper. Heat. A flocculent precipitate (a) falls at about 73° C.; filter this off; note that the filtrate is less acid than that from which the precipitate has separated, or it may even be alkaline. If so, make it faintly acid again, and heat; a precipitate falls at 77–79° C. (β). A third precipitate is similarly obtained at 84–86° C. (γ). In the serum of the ox, sheep, and horse the a precipitate is absent: in cold-blooded animals, the β and γ varieties are absent.

3. Take a fresh portion of B, and saturate it with sodium sulphate. The serum albumin is precipitated (completely after a long shaking). This is due to the formation of sodio-magnesium sulphate. B was already saturated with magnesium sulphate ($MgSO_4 + 7H_2O$); on adding sodium sulphate a double salt ($MgSO_4.Na_2SO_4 + 6H_2O$) is formed. Shake some serum with sodium sulphate alone. A small precipitate of globulin is produced. Saturate another portion of the serum with sodio-magnesium sulphate; both globulin and albumin are precipitated.

Of the methods used for precipitating serum globulin practically only two are used now. These are Hammarsten's and Kauder's. The other methods only precipitate the globulin incompletely. Kauder's method is rapid and efficacious: if the globulin is filtered off, the albumin may be precipitated in the filtrate by complete saturation with the same salt, ammonium sulphate. This method avoids the trouble of using two salts as described under 3. This last method is instructive, but not nearly so quick as Kauder's.

With regard to the separation of serum albumin into a, β, and γ varieties by the use of the method of fractional heat coagulation, it must be mentioned that at present no further difference has been shown to exist between them, and the opinion has been very freely expressed that the results obtained are not trustworthy. I am convinced that the method is a good one, especially as in other cases (see MUSCLE) the proteids so separated can be shown to possess other differences. In the case of serum, however—and the same is true for egg albumin—the matter must still be considered *sub judice*.

Recent research has shown that serum globulin is not a single proteid. We have already seen that the precipitation which occurs by means of dialysis is incomplete. It has now been shown that serum globulin as 'salted out' by means of the sulphate of magnesium or ammonium really consists of two proteids; one of these (eu-globulin) is precipitable by dialysis: the other (pseudo-globulin) is not.

LESSON XXI

COAGULATION OF BLOOD

1. Salted plasma and oxalate plasma are the two kinds of blood-plasma which are most easily prepared (Lesson IX.). The separation of the plasma and corpuscles may most readily be carried out by the centrifugal machine (one form of this is represented in the next figure); the corpuscles settle and the supernatant plasma is pipetted off.

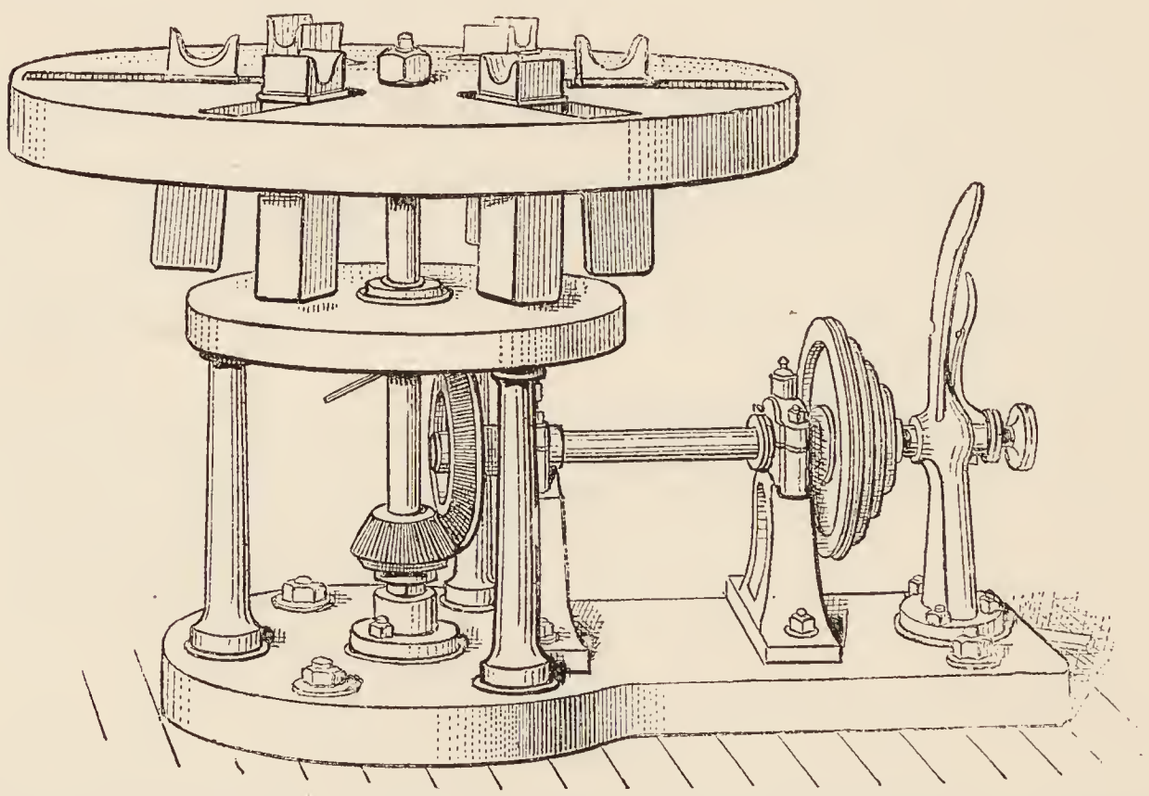


FIG. 60.—Centrifugal machine as made by Runne. Glass vessels containing the substances to be centrifugalised are placed within the six metallic tubes which hang vertically while the disc is at rest; when the machinery is set going they fly out into the horizontal position. A water motor or gas engine may be used to work the instrument.

The principal properties of these two forms of plasma have already been described in Lesson IX.; the following additional experiments may now be performed.

2. Heat a portion of the salted plasma to 60° C. The fibrinogen is coagulated by heat at 56° C. Filter. Dilute the filtrate as in Lesson IX. 3, and add fibrin ferment. No coagulation occurs.

3. Oxalate plasma or decalcified plasma coagulates when a little calcium chloride is added, but not if the oxalate plasma has been previously heated to 60° C. and filtered, because the fibrinogen has been thus removed.

4. **Hydrocele Fluid.**—This does not clot spontaneously, or only very slowly. Divide it into four parts—A, B, C, and D.

To A add an equal volume of serum.

To B add a few drops of fibrin ferment solution.

To C add a piece of buffy coat.

Put them into the warm bath; coagulation takes place in each. The serum or the buffy coat supplies the missing fibrin ferment. The serum does not produce its effect in virtue of the serum globulin it contains; hydrocele fluid contains both fibrinogen and serum globulin, as the following experiment shows:—

Take the portion D and half saturate it with sodium chloride by adding to it an equal bulk of saturated solution of sodium chloride. Fibrinogen is precipitated. The precipitate is a small one, and on standing aggregates together, and so becomes more apparent. Filter, and saturate the filtrate with sodium chloride, or, better, magnesium sulphate; serum globulin is precipitated.

5. **Intravascular Coagulation.**—A solution of nucleo-proteid from the thymus, testis, lymphatic glands, or kidney has been prepared beforehand by the demonstrator. It may be prepared in one or two ways.

(*a*) *Wooldridge's Method.*—The gland is cut up small and extracted with water for twenty-four hours. Weak acetic acid (0·5 c.c. of the acetic acid of the 'Pharmacopœia' diluted with twice its volume of water for every 100 c.c. of extract) is then added to the decanted liquid. After some hours the precipitated nucleo-proteid (called tissue-fibrinogen by Wooldridge) falls to the bottom of the vessel. This is collected and dissolved in 1-per-cent. sodium carbonate solution.

(*b*) *The Sodium Chloride Method.*—The finely divided gland is ground up in a mortar with about an equal volume of sodium chloride. The resulting viscous mass is poured into excess of distilled water. The nucleo-proteid rises to the surface of the water, where it may be collected and dissolved as before.

A rabbit is anæsthetised, and a cannula inserted into the external jugular vein. The solution is injected into the circulation through this. The animal soon dies from cessation of respiration; the eyeballs protrude and the pupils are widely dilated. On opening the animal the heart will be found still beating, and its cavities (especially on the right side) distended with clotted blood. The vessels, especially the veins, also are full of clot. The blood of the portal vein is usually clotted most. If a dog is employed instead of a rabbit in this experiment, coagulation is usually confined to the portal area. This is related to the greater venosity of the blood in this situation. If venosity is increased in any other area, as by tetanising the muscles of one leg, clotting will be found also in the veins of this region.

LESSON XXII

MUSCLE AND NERVOUS TISSUE

1. A rabbit has been killed and its muscles washed free from blood by a stream of salt solution injected through the aorta. The muscles have been quickly removed, chopped up small, and extracted with 5-per-cent. solution of magnesium sulphate. This extract is given out.

2. The extract will probably be faintly acid. The acid is sarco-lactic acid. It may be identified by Uffelmann's reaction (p. 165).

3. The coagulation of muscle is very like that of blood. This may be shown with the salted muscle plasma (the extract given out) as follows: Dilute some of it with four times its volume of water; divide it into two parts; keep one at 40° C. and the other at the ordinary temperature. Coagulation—that is, formation of a clot of myosin—occurs in both, but earliest in that at 40° C.

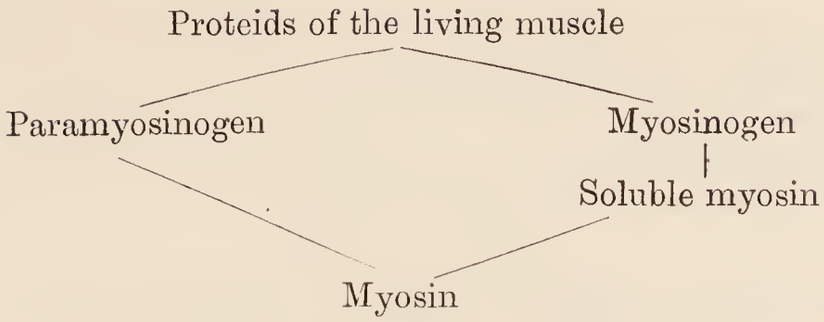
4. Remove the clot of myosin from 3; observe it is soluble in 10-per-cent. sodium chloride, and also in 0·2-per-cent. hydrochloric acid, forming syntonin.

5. Make an extract of muscle in the same way, using a small quantity of physiological salt solution instead of the strong solution of magnesium sulphate employed in the foregoing experiments. Such an extract contains the two principal muscular proteids, viz. paramyosinogen (v. Fürth's myosin) and myosinogen (v. Fürth's myogen), the two precursors of the muscle-clot or myosin (v. Fürth's muscle-fibrin). Small quantities of other proteids also present are mainly due to unavoidable mixture with small amounts of blood and lymph.

These two proteids differ in temperature of heat coagulation. Take the extract and heat it in a test-tube within a water-bath; at 47° C. paramyosinogen is coagulated; filter this off and heat the filtrate; at 56° C. flocculi of the coagulated myosinogen separate out.

6. Paramyosinogen is precipitable by dialysis and is a true globulin. Myosinogen is what is called an atypical globulin, and corresponds to the pseudo-globulin of blood serum and egg-white. Though readily salted out of solution like paramyosinogen it is not precipitable by dialysis.

7. In the process of clotting, such as occurs in *rigor mortis*, paramyosinogen is directly converted into myosin; whereas myosinogen first passes into a soluble modification (coagulable by heat at the remarkably low temperature of 40° C.) before myosin is formed. This is shown in a diagrammatic way in the following scheme:—



8. When a muscle is gradually heated, at a certain temperature it contracts permanently and loses its irritability. This phenomenon is known as *heat-rigor*, and is due to the coagulation of the proteids in the muscle. If a tracing is taken of the shortening, it is found that the first shortening occurs at the coagulation temperature of paramyosinogen (47°–50° C.), and if the heating is continued a second shortening occurs at 56° C., the coagulation temperature of myosinogen. If frog's muscles are used there are three shortenings—namely, at 40°, 47°, and 56° C.; frog's muscle thus contains an additional proteid which coagulates at 40° C. This additional proteid is the soluble myosin alluded to above, some of which, in the muscle of cold-blooded animals, is present before *rigor mortis* occurs.

In addition to the proteids mentioned, there is a small quantity of nucleo-proteid.

9. **Involuntary Muscle.**—The main facts just described for voluntary are true also for involuntary muscle. The chief distinction lies in the quantity of nucleo-proteid, which is more abundant in those forms of muscle, the fibres of which are least different from the mesoblastic cells from which all ultimately originate. This may be readily shown by the following simple experiment.

Take equal parts of voluntary muscle, heart muscle, and plain muscle (say from the stomach wall), and extract each for the same time with equal amounts of 0·15-per-cent. solution of sodium carbonate. Filter and add to each filtrate acetic acid, drop by drop. The extract of voluntary muscle gives only an opalescence; in the case of the plain muscle there is an abundant precipitate; the heart muscle gives a result intermediate between the other two.

10. **Pigments of Muscle:—**

(*a*) Notice the difference between the red and pale muscles of the rabbit.

(*b*) Examine a piece of red muscle (*e.g.* the diaphragm) spectroscopically for oxyhæmoglobin (or it may be more convenient to make an aqueous extract of the muscle and examine that).

(*c*) A piece of the pectoral muscle of a pigeon has been soaked in glycerin. Press a small piece between two glass slides and place it in front of the spectroscope. Observe and map out the bands of myohæmatin. This pigment is doubtless a derivative of hæmoglobin.

(*d*) Pieces of the same muscles have been placed in ether for twenty-four hours. The ether dissolves out a yellow lipochrome from the adherent fat. A watery fluid below contains modified myohæmatin. Filter it; compare its spectrum with that of hæmochromogen. The myohæmatin bands are

rather nearer the violet end of the spectrum (fig. 61, spectrum 2) than those of hæmochromogen (fig. 57, spectrum 9).

11. **Creatine**:—

(*a*) Take some of the red fluid described in 10, *d*, and let it evaporate to dryness in a desiccator over sulphuric acid (fig. 62).

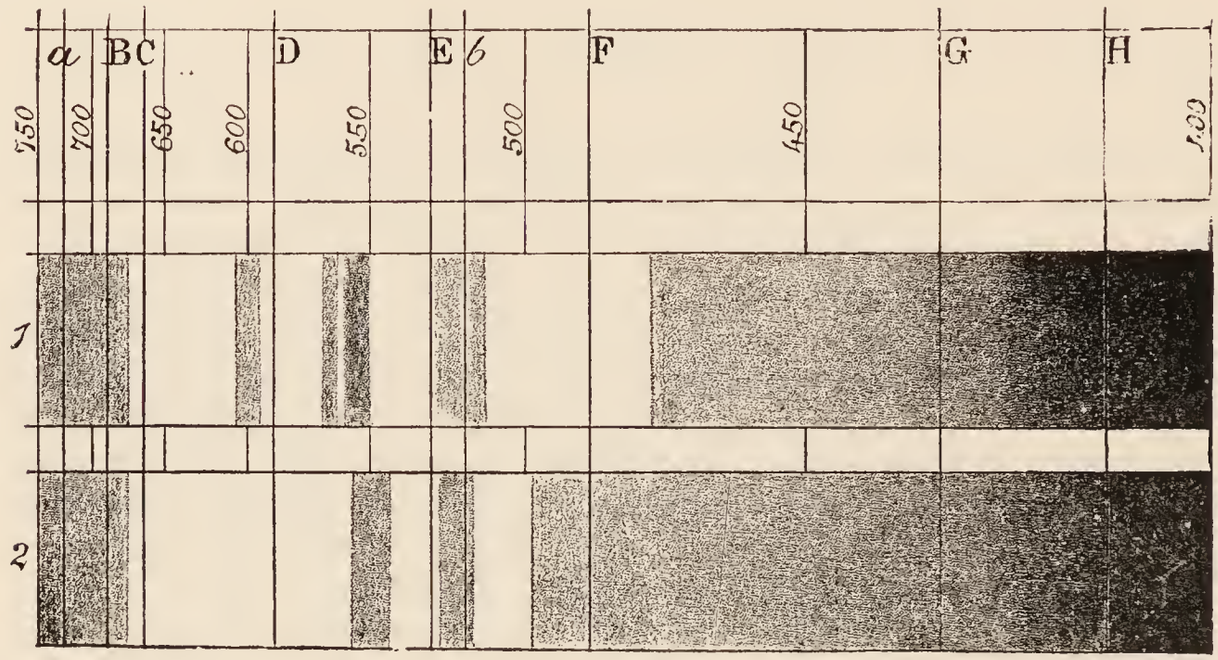


FIG. 61.—1, Absorption spectrum of myohæmatin, as seen in muscle rendered transparent by glycerin. 2, Absorption spectrum of modified myohæmatin.

In a day or two crystals of creatine tinged with myohæmatin separate out.

(*b*) Take an aqueous extract of muscle, like Liebig's extract or beef-tea; add baryta water to precipitate the phosphates, and filter. Remove excess

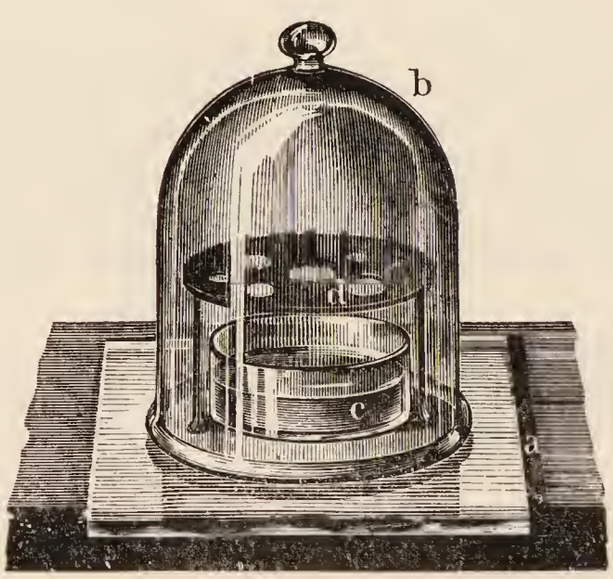


FIG. 62.—A desiccator. (Gscheidlen.) a, glass plate; b, bell-jar; c, dish of sulphuric acid; d, stand for capsules.

of baryta by a stream of carbonic acid; filter off the barium carbonate and evaporate the filtrate on the water-bath to a thick syrup. Set it aside to cool, and in a few days crystalline deposits of creatine will be found at the bottom of the vessel. These are washed with alcohol and dissolved in hot water.

On concentrating the aqueous solution crystals once more separate out, which may be still further purified by recrystallisation.

Creatinine can also be obtained from meat extracts by the methods described on p. 141.

NERVOUS TISSUES

The chemical investigation of nervous tissues is not well adapted to class exercises; still it may not be uninteresting to state briefly the principal known facts in relation to this subject. The most important points which any table of analysis will show are: (1) the large percentage of water, especially in the grey matter; (2) the large percentage of proteid. In grey matter, where the cells are prominent structures, this is most marked, and of the solids, proteid material here comprises more than half of the total. The following are some analyses which give the mean of a number of observations on the nervous tissues of human beings, monkeys, dogs, and cats:

—	Water	Solids	Percentage of Proteids in Solids
Cerebral grey matter . . .	83·5	16·5	51
„ white „ . . .	69·9	30·1	33
Cerebellum	79·8	20·2	42
Spinal cord as a whole . . .	71·6	28·4	31
Cervical cord	72·5	27·5	31
Dorsal cord	69·8	30·2	28
Lumbar cord	72·6	27·4	33
Sciatic nerves	65·1	34·9	29

The most important proteid is *nucleo-proteid*; there is also a certain amount of *globulin*, which, like the paramyosinogen of muscle, is coagulated by heat at the low temperature of 47° C. A certain small amount of neurokeratin (especially abundant in white matter) is included in the above table with the proteids. The granules in nerve cells (Nissl's bodies), which stain readily with methylene blue, are nucleo-proteid in nature. The next most abundant substances are of a fatty nature; the most prominent of these is the phosphorised fat called *lecithin* (see p. 25). In the nervous tissues lecithin is in part combined with cerebrin to form a complex substance called *protagon*, which crystallises out on cooling a hot alcoholic extract of brain or other nervous structures. *Cerebrin* is a term which probably includes several substances, which are nitrogenous glucosides; they yield on hydrolysis the sugar called galactose (see p. 18). They are sometimes called cerebrosides. There are other phosphorised fats as well, of which *kephalin* is the best known. The crystalline monatomic alcohol *cholesterin* (see p. 84) is also a fairly abundant constituent of nervous structures, especially of the white substance of Schwann. Finally there are smaller quantities of other extractives, and a small proportion of mineral salts (about 1 per cent. of the solids).

Fresh nervous tissues are alkaline, but, like most other living structures,

they turn acid after death. The change is particularly rapid in grey matter. The acidity is due to lactic acid.

Little or nothing is known of the chemical changes nervous tissues undergo during activity. We know that oxygen is very essential, especially for the activity of grey matter; cerebral anæmia is rapidly followed by loss of consciousness and death. Waller has suggested that small quantities of carbonic acid are produced during activity, because the increase in the action current (detected by the galvanometer) which occurs after a nerve has been repeatedly excited is very like the increase also noted on the application of small quantities of this gas. Waller's statement that the action current is increased by small amounts of carbonic acid has, however, received other explanations. Certainly large quantities of carbonic acid act like an anæsthetic, abolishing nervous activity. Of all parts of the nervous system, the cells in the grey matter are those which most readily manifest fatigue; the next most sensitive region is the termination of nerves in such endings as the end-plates. Fatigue in a medullated nerve-trunk has never yet been experimentally demonstrated; Waller's view that this is due to inter-nutritional changes between the axis cylinder and the investing medullary sheath can hardly be considered proved, for it is just as difficult to demonstrate fatigue in non-medullated nerves.

Chemistry of Nerve-degeneration.—Mott and I have shown that, in the disease General Paralysis of the Insane, the marked degeneration that occurs in the brain is accompanied by the passing of the products of degeneration into the cerebro-spinal fluid. Of these, nucleo-proteid and choline—a decomposition product of the lecithin (see p. 26)—are those which can be most readily detected. Choline can also be found in the blood. But this is not peculiar to the disease just mentioned, for in various other degenerative nervous diseases (combined sclerosis, disseminated sclerosis, meningitis, alcoholic neuritis, beri-beri, &c.) choline can also be detected in these situations. The tests employed to detect choline are mainly two: (1) a chemical test, namely the obtaining of the characteristic octahedral crystals of the platinum double salt from the alcoholic extract of the cerebro-spinal fluid or blood;[1] (2) a physiological test, namely the lowering of arterial blood-pressure (partly cardiac in origin, and partly due to dilatation of peripheral vessels) which a saline solution of the residue of the alcoholic extract produces: this fall is abolished, or even replaced by a rise of arterial pressure, if the animal has

[1] This test is performed as follows:—The fluid is diluted with about five times its volume of alcohol and the precipitated proteids are filtered off. The filtrate is evaporated to dryness at 40° C. and the residue dissolved in absolute alcohol and filtered; the filtrate from this is again evaporated to dryness, and again dissolved in absolute alcohol, and this should be again repeated. To the final alcoholic solution, an alcoholic solution of platinum chloride is added, and the precipitate so formed is allowed to settle and washed with absolute alcohol by decantation; the precipitate is then dissolved in 15-per-cent. alcohol, filtered, and the filtrate is allowed to slowly evaporate in a watch-glass at 40° C. The crystals can then be seen with the microscope. They are recognised not only by their yellow colour and octahedral form, and by their solubility in water and 15-per-cent. alcohol, but also by the fact that on incineration they yield 31 per cent. of platinum and give off the odour of trimethylamine. There is a danger of mistaking such crystals for those obtained from the chlorides of potassium and ammonium; but the presence of such contaminations may be minimised by the use of alcohol as water-free as possible.

been atropinised. It is possible that such tests may be of diagnostic value in the distinction between organic and so-called functional diseases of the nervous system.

A similar condition can be produced artificially in animals by a division of large nerve-trunks; and is most marked in those animals in which the degenerative process is at its height as tested histologically by the Marchi reaction.[1] A chemical analysis of the nerves themselves was also made. A series of cats was taken, both sciatic nerves divided, and the animals subsequently killed at intervals varying from 1 to 106 days. The nerves remain practically normal as long as they remain irritable: that is, up to three days after the operation. They then show a progressive increase in the percentage of water, and a progressive decrease in the percentage of phosphorus, until degeneration is complete. When regeneration occurs, the nerves return approximately to their previous chemical condition. One chemical feature of degeneration is the replacement of phosphorised by non-phosphorised fat. When the Marchi reaction disappears in the later stages of degeneration, the non-phosphorised fat has been absorbed. This absorption occurs earlier in the peripheral nerves than in the central nervous system. The non-phosphorised fat of degenerated myelin is also either richer in olein, or the olein is more loosely combined than in the healthy medullary sheath; hence the deeper reaction with osmic acid even in the presence of chromic acid as in the Marchi test. The following table gives details of these experiments:

Cat's sciatic nerves				Condition of blood	Condition of nerves
—	Water	Solids	Percentage of phosphorus in solids		
Normal	65·1	34·9	1·1	Minimal traces of choline present	Nerves irritable and histologically healthy
1 to 3 days after section	64·5	35·5	0·9		
4 to 6 „	69·3	30·7	0·9	Choline more abundant	Irritability lost; degeneration beginning
8 „	68·2	31·8	0·5	Choline abundant	Degeneration well shown by Marchi reaction
10 „	70·7	29·3	0·3		
13 „	71·3	28·7	0·2		
25–27 „	72·1	27·9	traces	Choline much less	Marchi reaction still seen, but absorption of degenerated fat has set in
29 „	72·5	27·5	0		
44 „	72·6	27·4	0	Choline almost disappeared	Absorption of fat practically complete
100 to 106 „	66·2	33·8	0·9		Return of function, nerves regenerated

The foregoing figures relate to the peripheral portions of the nerves. Noll has also shown that the phosphorised fat diminishes somewhat in the central ends of cut nerves due to 'disuse atrophy.'

[1] The Marchi reaction is the black staining that the medullary sheath of degenerated nerve fibres shows when, after being hardened in Müller's fluid, they are treated with Marchi's reagent, a mixture of Müller's fluid and osmic acid. Healthy nerve fibres are but little affected by the reagent, but degenerated myelin is blackened like the fat of normal adipose tissue.

Further, it has been found that in human spinal cords in which a unilateral degeneration of the pyramidal tract has been produced by a lesion in the opposite hemisphere, and which gives the Marchi reaction, there is a similar increase of water and diminution of phosphorus on the degenerated side.

Cerebro-spinal Fluid.—This plays the part of the lymph of the central nervous system, but differs considerably from all other forms of lymph. It is a very watery fluid, containing, besides some inorganic salts similar to those of the blood, a trace of proteid matter (globulin) and a small amount of a reducing substance, the nature of which was for a long time uncertain, but which seems now to have been proved to be sugar. It contains the merest trace of choline; but this is not devoid of significance, for this fact taken in conjunction with another— namely, that physiological saline solution will extract from perfectly fresh nervous matter a small quantity of choline— shows us that lecithin is not a stable substance, but is constantly breaking down and building itself up afresh; in fact, undergoing the process called metabolism. This is most marked in the most active region of the brain— viz., the grey matter.

LESSON XXIII

UREA AND CHLORIDES IN URINE

ESTIMATION OF UREA

If albumin is present it must be first separated by boiling after acidulation with acetic acid if necessary, and filtering off the flakes of coagulated proteid.

The three chief methods of estimating urea are—

(*a*) The mercuric nitrate or Liebig's method.

(*b*) The hypobromite, or Hüfner's method.

(*c*) The method of Mörner and Sjöqvist.

a. **Liebig's Method.**—The compound formed when mercuric nitrate is added to urea has the formula $(CON_2H_4)_2Hg(NO_3)_2(HgO)_3$. It forms a white precipitate, insoluble in water and weak alkaline solutions. It is therefore necessary to prepare a standard solution of mercuric nitrate, and to have an indicator by which to detect the point when all the urea has entered into combination with the mercury salt, and the latter slightly predominates. This indicator is sodium carbonate, which gives a yellow colour with the excess of mercuric nitrate owing to the formation of hydrated mercuric oxide.

Theoretically, 100 parts of urea should require 720 parts of mercuric oxide, but practically 772 of the latter are necessary to remove all the urea, and at the same time show the yellow colour with alkali; consequently the solution of mercuric nitrate must be of empirical strength in order to give accurate results.

The following solutions must be prepared—

i. Standard mercuric nitrate solution. Dissolve 77·2 grammes of red oxide of mercury (weighed after it has been dried over a water-bath), or 71·5 gr. of the metal itself, in dilute nitric acid. Expel excess of acid by evaporating the liquid to a syrupy consistence. Make up to 1,000 c.c. with distilled water, adding the water gradually. This solution is of such a strength that 19 c.c. will precipitate 10 c.c. of a 2-per-cent. urea solution. Add 52·6 c.c. of water to the litre of the mercuric nitrate solution and shake well; then 20 c.c. (instead of 19) = 10 c.c. 2-per-cent. urea solution, *i.e.* 1 c.c. = ·01 gr. urea.

ii. Baryta mixture. This is a mixture of two volumes of solution of barium hydrate with one of solution of barium nitrate, both saturated in the cold.

Analysis.—Take 40 c.c. of urine. Add to this 20 c.c. baryta mixture and filter off the precipitate of barium salts (phosphate and sulphate). Take 15 c.c. of the filtrate (this corresponds to 10 c.c. of urine) in a beaker. Run into it the mercuric nitrate solution from a burette, until it is found that, on

mixing a drop of the mixture with a drop of saturated solution of sodium carbonate on a white tile, a pale lemon colour is produced. Then read the amount used from the burette, and calculate thence the percentage of urea.

Corrections.—This method only approaches accuracy when the quantity of urea present is about 2 per cent., which is about the normal percentage of urea in urine. The chlorine in the urine must also be estimated, and the quantity of urea indicated reduced by the subtraction of 1 gramme of urea for every 1·3 gramme of sodium chloride found. If the urine contains less than 2 per cent. of urea, 1·0 c.c. of mercuric nitrate solution must be deducted for every 4 c.c. used; if more than 2 per cent. of urea, a second titration must be performed with the urine diluted with half as much water as has been needed of the mercurial solution above 20 c.c. Suppose that 28 c.c. have been used in the first titration, the excess is 8 c.c.; therefore 4 c.c. of water must be added to the urine before the second titration is made. When ammonium carbonate is present, first estimate the urea in one portion of urine, and the ammonia by titration with normal sulphuric acid in another; 0·017 gramme of ammonia = 0·030 of urea. The equivalent of ammonia must be added to the urea found in the first portion of urine.

b. **The Hypobromite Method.**—This method (see p. 121) holds its own, as it is easy and sufficiently exact for clinical purposes. When absolute accuracy is necessary, one of the recently introduced methods must be employed, and of these that of Mörner and Sjöqvist appears to be best.

(*c*) **Method of Mörner and Sjöqvist.**—The following reagents are necessary:

i. A saturated solution of barium chloride containing 5 per cent. of barium hydrate.

ii. A mixture of ether and alcohol in proportion 1 : 2.

iii. The apparatus, &c., necessary for carrying out Kjeldahl's method of estimating nitrogen (see p. 215).

Analysis.—Five c.c. of urine are mixed with 5 c.c. of the barium mixture and 100 c.c. of the mixture of ether and alcohol. By this means all nitrogenous substances except urea are precipitated. Twenty-four hours later this is filtered off, and the precipitate is washed with 50 c.c. of the ether-alcohol mixture, the filter-pump being used to accelerate the process. The washings are added to the filtrate; a little magnesia is added to this to drive off ammonia. The alcohol and ether are then driven off at a temperature of 55° C., and evaporation is continued at this temperature until the volume of the residue is 10–15 c.c. The nitrogen in this is estimated by Kjeldahl's method. The nitrogen found is multiplied by 2·143, and the result is the amount of urea.

PREPARATION OF UREA FROM URINE

(1) Evaporate the urine to a small bulk. Add strong pure nitric acid in excess, keeping the mixture cool during the addition of the acid. Pour off the excess of fluid from the crystals of urea nitrate which are formed; strain through muslin and press between filter paper. Add to the dry product barium carbonate in large excess. This forms barium nitrate and sets the urea free. Mix thoroughly with sufficient methylated spirit to form a paste. Dry on a water-bath and extract with alcohol; filter; evaporate the filtrate

on a water-bath and set aside. The urea crystallises out, and may be decolorised by animal charcoal and purified by recrystallisation.

(2) The following method is well adapted for the preparation of microscopic specimens of urea and urea nitrate: Take 20 c.c. of urine; add baryta mixture until no further precipitate is produced; filter, evaporate the filtrate to a thick syrup on the water-bath, and extract with alcohol; pour off and filter the alcoholic extract; evaporate it to dryness on the water-bath and take up the residue with water. Place a drop of the aqueous solution on a slide and allow it to crystallise; crystals of urea separate out. Place another drop on another slide and add a drop of nitric acid; crystals of urea nitrate separate out.

ESTIMATION OF CHLORIDES

The chlorides in the urine consist of those of sodium and potassium, the latter only in small quantities. The method adopted for the determination of the total chlorides consists in their precipitation by a standard solution of silver nitrate or mercuric nitrate.

Mohr's Method.—Precipitation by silver nitrate.

The following solutions must be prepared:

Standard silver nitrate solution. Dissolve 29·075 grammes of fused nitrate of silver in a litre (1,000 c.c.) of distilled water; 1 c.c. = 0·01 gramme of sodium chloride.

(*a*) Saturated solution of neutral potassium chromate.

Analysis.—Take 10 c.c. of urine; dilute with 100 c.c. of distilled water.

Add to this a few drops of the potassium chromate solution.

Drop into this mixture from a burette the standard silver nitrate solution; the chlorine combines with the silver to form silver chloride, a white precipitate. When all the chlorides are so precipitated, silver chromate (red in colour) goes down, but not while any chloride remains in solution. The silver nitrate must therefore be added until the precipitate has a pink tinge.

From the amount of standard solution used, the quantity of sodium chloride in 10 c.c. of urine, and thence the percentage, may be calculated.

Sources of Error and Corrections.—A high-coloured urine may give rise to difficulty in seeing the pink tinge of the silver chromate: this is overcome by diluting the urine more than stated in the preceding paragraph.

1 c.c. should always be subtracted from the total number of c.c. of the silver nitrate solution used, as the urine contains small quantities of certain compounds more easily precipitable than the chromate.

(*b*) To obviate such sources of error the following modification of the test, as described by Sutton, may be used: 10 c.c. of urine are measured into a thin porcelain capsule and 1 gramme of pure ammonium nitrate added; the whole is then evaporated to dryness, and gradually heated over a small spirit lamp to low redness till all vapours are dissipated and the residue becomes white. It is then dissolved in a small quantity of water, and the carbonates produced by the combustion of the organic matter neutralised by dilute acetic acid; a few grains of pure calcium carbonate to remove all free acid are then added, and one or two drops of potassium chromate. The mixture is then titrated with decinormal silver solution

(16·966 gr. of silver nitrate per litre) until the end reaction, a pink colour, appears. Each c.c. of silver solution represents 0·005837 gr. of salt; consequently, if 12·5 c.c. have been used, the weight of salt in the 10 c.c. of urine is 0·07296 gr., or 0·7296 per cent. If 5·9 c.c. of urine are taken for titration, the number of c.c. of silver solution used will represent the number of parts of salt per 1,000 parts of urine.

(c) *Liebig's Method.*—Precipitation by mercuric nitrate.

The following solutions must be first prepared:

i. Standard mercuric nitrate solution: Dissolve 20 grammes of pure mercury in boiling nitric acid; then dilute to nearly a litre. To dilute this to the right strength, preliminary experiments must be performed with a standard solution of pure sodium chloride, 20 grammes to the litre. Take 10 c.c. of the standard sodium chloride solution, add to this 2 c.c. of a 4-per-cent. solution of urea and 5 c.c. of a saturated solution of sodium sulphate. Into this mixture allow the mercuric nitrate solution to flow from a burette, stirring the mixture the while. A precipitate forms, which redissolves on stirring; add the mercuric nitrate solution till a permanent precipitate (not an opalescence) forms; the reaction is then complete. The strength of the mercurial solution is thus determined, and it is then diluted so that 20 c.c. = 0·2 gramme of sodium chloride = 10 c.c. of the standard sodium chloride solution; 1 c.c., therefore, corresponds to 0·01 gramme of sodium chloride, or 0·006059 gramme of chlorine.

ii. Baryta mixture.

iii. Dilute nitric acid (1 in 20).

Analysis.—Take 40 c.c. of urine.

Add 20 c.c. of baryta mixture. Filter off the precipitate.

Take 15 c.c. of the filtrate: this corresponds to 10 c.c. of the original urine.

Render this slightly acid with dilute nitric acid.

Run in the standard mercuric nitrate solution from a burette, stirring the mixture well until a permanent precipitate appears.

Read off the number of c.c. used; multiply by 0·01. This gives the amount of chloride as sodium chloride contained in 10 c.c. urine.

Explanation and Corrections.—This test depends on the fact that when mercuric nitrate and sodium chloride in solution are mixed, sodium nitrate and mercuric chloride, which are both soluble in water, are formed. It is not till all the chloride in the urine is so decomposed that mercuric nitrate begins to combine with the urea present to form a permanent white precipitate. Hence the necessity of estimating the chlorides when using Liebig's method for the determination of urea.

In order to obtain the exact point at which the precipitate becomes a permanent one, the process must be repeated in another specimen. The advantage of this process is its simplicity; its disadvantage is that the end point is rather obscure.

LESSON XXIV

ESTIMATION OF PHOSPHATES AND SULPHATES IN URINE

ESTIMATION OF PHOSPHATES

The phosphoric acid in the urine is combined with soda, potash, lime, and magnesia.

(*a*) *Estimation of the total phosphates.*

For this purpose the following reagents are necessary:

i. A standard solution of uranium nitrate. The uranium nitrate solution contains 35·5 grammes in a litre of water; 1 c.c. corresponds to 0·005 gramme of phosphoric acid (P_2O_5).

ii. Acid solution of sodium acetate. Dissolve 100 grammes of sodium acetate in 900 c.c. of water; add to this 100 c.c. of glacial acetic acid.

iii. Solution of potassium ferrocyanide.

Method.—Take 50 c.c. of urine. Add 5 c.c. of the acid solution of sodium acetate.[1] Heat the mixture to 80° C.

Run into it while hot the standard uranium nitrate solution from a burette until a drop of the mixture gives a distinct brown colour with a drop of potassium ferrocyanide placed on a porcelain slab. Read off the quantity of solution used and calculate therefrom the percentage amount of phosphoric acid in the urine.

(*b*) *Estimation of the phosphoric acid combined with lime and magnesia* (alkaline earths).

Take 200 c.c. of urine. Render it alkaline with ammonia. Lay the mixture aside for twelve hours. Collect the precipitated earthy phosphates on a filter; wash with dilute ammonia (1 in 3). Wash the precipitate off the filter with water acidified by a few drops of acetic acid. Dissolve with the aid of heat, adding a little more acetic acid if necessary. Add 5 c.c. of the acid solution of sodium acetate. Bring the volume up to 50 c.c., and estimate the phosphates in this volumetrically by the standard uranium nitrate as before. Subtract the phosphoric acid combined with the alkaline earths thus obtained from the total quantity of phosphoric acid, and the difference is the amount of acid combined with the alkalis, soda and potash.

(*c*) Instead of uranium nitrate, a standard solution of uranium acetate may be used. The directions for the making of these standard solutions will be found in Sutton's 'Volumetric Analysis.' As a rule, it is less troublesome, and not much more expensive, to purchase standard solutions ready-made.

[1] In using uranium nitrate it is imperative that sodium acetate should accompany the titration in order to avoid the possible occurrence of free nitric acid in the solution. If uranium acetate is used, it may be omitted.

ESTIMATION OF SULPHATES

The sulphates in the urine are of two kinds: the pre-formed sulphates—viz. those of soda and potash—and the combined or ethereal sulphates.

(*a*) For the determination of the total amount of sulphuric acid (SO_3) (*i.e.* pre-formed and combined sulphuric acid together) in the urine, one of two methods is adopted:

1. Volumetric method.
2. Gravimetric method.

Both methods will be given here; the former is, however, better suited for class experiments.

1. *Volumetric Determination.*—This process consists in adding to a given volume of the urine a standard solution of chloride of barium so long as a precipitate of barium sulphate is formed.

The following solutions are necessary:

i. Standard barium chloride solution: 30·5 grammes of crystallised chloride of barium in a litre of distilled water; 1 c.c. of this solution corresponds to 0·01 gramme of sulphuric acid (SO_3).

ii. Solution of sulphate of potash: 20 per cent.

iii. Pure hydrochloric acid.

Method.—100 c.c. of urine are taken in a flask. This is rendered acid by 5 c.c. of hydrochloric acid, and boiled. The combined sulphates are thus converted into ordinary sulphates, and give a precipitate like them with barium chloride. The chloride of barium solution is allowed to drop into this mixture as long as any precipitate occurs, the mixture being heated before every addition of barium chloride to it. After adding 5 to 8 c.c. of the standard solution, allow the precipitate to settle; pipette off a few drops of the clear supernatant fluid into a watch-glass; add to it a few drops of the standard barium chloride solution. If any precipitate occurs, return the whole to the flask and add more barium chloride; again allow the precipitate to settle, and test as before; go on in this way until no more barium sulphate is formed on the addition of barium chloride.

Excess of barium chloride must also be avoided; when only a trace of excess is present a drop of the clear fluid removed from the precipitate gives a cloudiness with a drop of the potassium sulphate solution placed on a glass plate over a black ground. If more than a cloudiness appears, too large a quantity of barium chloride has been added, and the operation must be repeated. From the quantity of barium chloride solution used, the percentage of sulphuric acid in the urine is calculated.

2. *Gravimetric Determination* (*i.e.* by weight).—This method consists in weighing the precipitate of barium sulphate obtained by adding barium chloride to a known volume of urine; 100 parts of sulphate of barium correspond to 34·33 parts of sulphuric acid (SO_3).

Method (Salkowski).—100 c.c. of urine are taken in a beaker. This is boiled with 5 c.c. of hydrochloric acid as before.

Chloride of barium is added till no more precipitate occurs.

The precipitate is collected on a small filter of known ash, and washed with hot distilled water till no more barium chloride occurs in the filtrate—

i.e. until the filtrate remains clear after the addition of a few drops of hydric sulphate. Then wash with hot alcohol, and afterwards with ether. Remove the filter, and place it with its contents in a platinum crucible. Heat to redness. Cool over sulphuric acid in a desiccator; weigh, and deduct the weight of the crucible and filter ash; the remainder is the weight of barium sulphate formed.

Error.—When the experiment is carried out as above there is a slight error from the formation of a small quantity of sulphide of barium. This may be corrected as follows: After the platinum crucible has become cool add a few drops of pure sulphuric acid (H_2SO_4). The sulphide is converted into sulphate. Heat again to redness to drive off excess of sulphuric acid.

(*b*) The following is Salkowski's method of estimating the *combined sulphuric acid*—that is, the amount of SO_3 in ethereal sulphates:—100 c.c. of urine are mixed with 100 c.c. of alkaline barium chloride solution, which is a mixture of two volumes of solution of barium hydrate with one of barium chloride, both saturated in the cold. The mixture is stirred, and after a few minutes filtered: 100 c.c. of the filtrate (= 50 c.c. of urine) are acidified with 10 c.c. of hydrochloric acid. boiled, kept at 100° C. on the water-bath for an hour, and then allowed to stand till the precipitate has completely settled; if possible, it should be left in this way for twenty-four hours. The further treatment of this precipitate (= combined sulphates) is then carried out as in the last case.

Calculation.—233 parts of barium sulphate correspond to 98 parts of H_2SO_4, or 80 parts of SO_3 or 32 parts of S. To calculate the H_2SO_4, multiply the weight of barium sulphate by $\frac{98}{233} = 0{\cdot}4206$; to calculate the SO_3, multiply by $\frac{80}{233} = 0{\cdot}34335$; to calculate the S multiply by $\frac{32}{233} = 0{\cdot}13734$. This method of calculation applies to the gravimetric estimation both of total sulphates and of combined sulphates.

(*c*) To obtain the amount of pre-formed sulphuric acid, subtract the amount of combined SO_3 from the total amount of SO_3. The difference is the pre-formed SO_3.

Example: 100 c.c. of urine gave 0·5 gramme of total barium sulphate. This multiplied by $\frac{80}{233} = 0{\cdot}171$ gr. = total SO_3. Another 100 c.c. of the same urine gave 0·05 gr. of barium sulphate from ethereal sulphates; this multiplied by $\frac{80}{233} = 0{\cdot}017$ gr. of combined SO_3. Total SO_3 − combined SO_3 $= 0{\cdot}171 - 0{\cdot}017 = 0{\cdot}154$ gr. of pre-formed SO_3.

LESSON XXV

URIC ACID AND CREATININE

1. **Preparation of Pure Uric Acid.**—If one wishes to prepare pure uric acid, the solid urine of a reptile or bird, which consists principally of the acid ammonium salt, should be selected; one has not then to separate any pigment. It is boiled with 10-per-cent. caustic soda or ammonia, diluted, and then allowed to stand. The clear fluid is decanted and poured into a large excess of water to which 10 per cent. of hydrochloric acid has been added; after twenty-four hours, crystals of uric acid are deposited. These may be purified by washing, re-solution in soda, and re-precipitation by acid.

2. **Estimation of Uric Acid** (Hopkins's method).—The following reagents are required: Pure chloride of ammonium, finely powdered.

A wash bottle containing a filtered saturated solution of the same salt.

A twentieth normal solution of potassium permanganate made by dissolving 1·581 grammes of permanganate in a litre of water.

Measure 100 c.c. of urine into a beaker of about 150 c.c. capacity.

Add to this 25 grammes (approximately weighed) of ammonium chloride, stirring briskly till all the salt is dissolved. Now add 2 c.c. of strong ammonium hydrate, and allow the mixture to stand until the precipitate of ammonium urate, which rapidly forms, has wholly settled to the bottom of the beaker; its subsidence is promoted by occasional brisk stirring.

Adjust a small filter paper (7 cm. diam.) in a funnel of such size that only a small margin of glass projects above the edge of the folded paper, and transfer to this the ammonium urate precipitate.

Filtration should not be commenced until the precipitate has settled satisfactorily. The precipitate should be as far as possible retained in the beaker until the greater part of the clear liquid has filtered through; finally transfer the whole to the filter with the help of a wash bottle containing saturated ammonium chloride solution. After the filter has thoroughly drained, wash the precipitate twice again with the same solution.

While the last washings are running through the paper, distilled water should be heated to boiling in a wash bottle provided with a *fine* jet. The funnel containing the filter is now held horizontally over a small porcelain basin (of about 50 c.c. capacity) and the precipitate washed into the latter with a jet of hot water, the filter itself being afterwards opened out over the basin in order that any urate adhering to its folds may be washed off. Not more than 20–30 c.c. of water need be employed in this transference: if much more has been used the liquid should be concentrated over the water-bath at this stage. A little strong HCl (1 c.c.) is next added to the contents of the basin, and the whole is then heated over a burner until it just reaches the boiling point. It is then set aside for the uric acid to crystallise out.

If the mixture is artificially cooled all the uric acid will separate out in two hours, otherwise it is best allowed to stand overnight or longer.

The crystals are filtered off through a very small filter paper (4 cm. diam.) ; the filtrate is received into a graduated cylinder so that the amount of mother liquid may be noted (see below). The uric acid is next washed with cold distilled water until free from chlorides. It is unnecessary to transfer the whole to the filter; the greater part may be washed by decantation. Such of the crystals as are upon the filter are now washed back into the basin (best by the aid of hot water) and the whole quantity is dissolved by heating to boiling with 1 c.c. of 10-per-cent. sodium carbonate solution and as much distilled water as the basin will safely hold.

The solution is transferred to a $\frac{1}{4}$-litre Erlenmeyer flask, which should be marked roughly at 100 c.c. The solution is made up to this mark with distilled water, and cooled to the temperature of the room.

Twenty c.c. of strong sulphuric acid are added to the contents of the flask, and the mixture shaken and titrated with the standard permanganate solution.

During the addition of the standard solution the liquid in the flask should be kept in vigorous movement. It will be found that at first the disappearance of the pink colour is so rapid that each drop as it is added is decolorised before it has time to *diffuse* through the whole liquid. The first instantaneous appearance of a diffused flush throughout the solution indicates the end point of the reaction. This colour rapidly disappears, but it will be found that the effect of adding further quantities of permanganate after the end point has been passed is quite different from the effect before the end point was reached ; each drop is now able to diffuse throughout the fluid.

For each c.c. of the solution necessary to produce the end point just described 0·00375 gramme of uric acid is present. To the value so obtained 1 mgm. must be added for each 15 c.c. of the mother liquor from which the crystals separated. Thus the uric acid from 100 c.c. of a sample of urine used up 18·5 c.c. of the standard permanganate solution. The mother liquor filtered from crystals measured 25 c.c.

$$18{\cdot}5 \times {\cdot}00375 = {\cdot}0694 \text{ gr. nearly}$$
$$\cdot001 \times \frac{25}{15} = {\cdot}0017 \quad ,,$$
$$\text{Total} = \overline{{\cdot}0711}$$

The urine contained 71 mgms. uric acid per 100 c.c.

3. **Estimation of Creatinine.**—The crystalline compound which creatinine forms with zinc chloride is employed in estimating the quantity of creatinine in urine ; 100 parts of the compound correspond to 62·42 of creatinine.

Method.—Take 250 c.c. of urine. Add milk of lime and calcium chloride in excess to precipitate the phosphates. Filter, and evaporate the filtrate to a small bulk ; to this add 50 c.c. absolute alcohol, and let the mixture stand for six hours. Then add 10 or 15 drops of an alcoholic solution of zinc chloride ; the crystals form, and after two or three days' standing in a dark place may be collected on a weighed filter. Wash with 90 per cent. alcohol, dry and weigh, and thence calculate the percentage of creatinine.

Johnson's method is described on p 141.

LESSON XXVI

THE PIGMENTS OF THE URINE

The urinary pigments are numerous, and have from time to time been described under different names by various observers.

1. **Urochrome.**—This is the essential yellow pigment of the urine. The word was originally introduced by Thudichum, and the substance he obtained is now recognised to have been a mixture of several pigments, of which, however, the essential yellow pigment formed a large proportion. Garrod's method of separating it from the urine is as follows:—

The urine is saturated with ammonium sulphate and filtered. The filtrate contains the pigment; this is shaken with alcohol. The alcohol separates readily from the saline mixture, and as it does so dissolves out much of the urochrome. By repeated extraction all the pigment passes into solution in the alcohol. The alcoholic solution is diluted with water, and the mixture again saturated with ammonium sulphate. The alcohol containing the pigment in solution again separates out. The second alcoholic solution is made faintly alkaline with ammonia and evaporated to dryness. The residue is extracted once or twice with acetic ether, and then again dissolved in strong alcohol. Finally the alcohol is concentrated till it is deep orange in tint, and poured into an equal volume of ether. The pure pigment is by this means precipitated as an amorphous brown powder.

Urochrome shows no absorption bands. As already stated (p. 123), it is probably an oxidation product of urobilin.

2. **Urobilin.**—Urobilin is a derivative of the blood-pigment, and is identical with stercobilin (see pp. 83, 123). Probably both reduction and hydration occur in its formation. It is very like the substance named hydrobilirubin by Maly, which he obtained by the action of sodium amalgam on bilirubin. The following formulæ[1] show the relationship between these allied pigments:—

Hæmatin	$C_{32}H_{32}N_4O_4Fe$
Bilirubin	$C_{32}H_{36}N_4O_6$
Hydrobilirubin	$C_{32}H_{40}N_4O_7$

Urobilin is probably a further stage in reduction.

Normal urine contains but little urobilin; what is present is chiefly in the form of a colourless chromogen, which by oxidation is converted into urobilin. In numerous pathological conditions urobilin is abundant. The

[1] The formulæ given are those of Nencki and Sieber. They differ from those previously given (p. 101) by Hoppe-Seyler.

so-called 'pathological urobilin' described by previous writers is ordinary urobilin incompletely separated from various impurities.

The following are the methods introduced by Garrod and Hopkins for its separation from the urine :—

(*a*) The urine is first saturated with ammonium chloride, and the urate so precipitated is filtered off. The filtrate is then acidified with sulphuric acid and saturated with ammonium sulphate. This causes a precipitate of urobilin, which may be collected and dissolved in water. The aqueous solution is again saturated with ammonium sulphate, and the pigment is thus precipitated in a state of purity.

(*b*) The urates are first removed, then the urine is acidified and saturated with ammonium sulphate as before. The urobilin is then extracted from the mixture by shaking it with a mixture of chloroform and ether (1 : 2) in a large separating funnel. The ether-chloroform extract is then rendered faintly alkaline and shaken with distilled water, and the urobilin passes into solution in the water. The aqueous solution is now once more saturated with ammonium sulphate and slightly acidified; it then once more yields its pigment to ether-chloroform.

By means of either of these methods urobilin is obtained in a pure condition; even normal urine will give some, for the chromogen is partly converted into the pigment by the acid employed.

Urobilin dissolved in alcohol exhibits a green fluorescence, which is greatly increased by the addition of zinc chloride and ammonia. It shows a well-marked absorption band between *b* and F, slightly overlapping the latter (fig. 63, spectrum 4).

Urobilin, like most animal pigments, shows acidic tendencies and forms compounds with bases; it is liberated from such combinations by the addition of an acid.

If urobilin is dissolved in caustic potash or soda, and sufficient sulphuric or hydrochloric acid is added to render the liquid faintly acid, a turbidity is produced. This turbid liquid shows an additional band in the region of the E line (fig. 63, spectrum 6), which is probably due to the special light absorption exercised by fine particles of urobilin in suspension. It wholly disappears when the precipitate is filtered off, and when it is re-dissolved the ordinary band alone is visible.

3. **Uroerythrin.**—This is the colouring matter of pink urate sediments. It may be separated from the sediment as follows:—The deposit is washed with ice-cold water, dried, and placed in absolute alcohol. The alcohol, though a solvent for uroerythrin, does not extract it from the urates. The alcohol is poured off, and the deposit dissolved in warm water. From this solution the pigment is easily extracted by amylic alcohol.

Uroerythrin has a great affinity for urates, with which it appears to form a loose compound. Its solutions are rapidly decolorised by light. Spectroscopically it shows two rather ill-defined bands (fig. 63, spectrum 7). It gives a green colour with caustic potash, and red or pink with mineral acids. Uroerythrin appears to be a small but constant constituent of urine. Its origin and relationship to other pigments are unknown.

4. **Hæmatoporphyrin.**—This also occurs in small quantities in normal

urine. In some pathological conditions, especially after the administration of certain drugs (*e.g.* sulphonal), its amount is increased. Its amount is stated to increase when the urine stands; this points to the existence of a colourless chromogen. It may be separated from the urine as follows:—

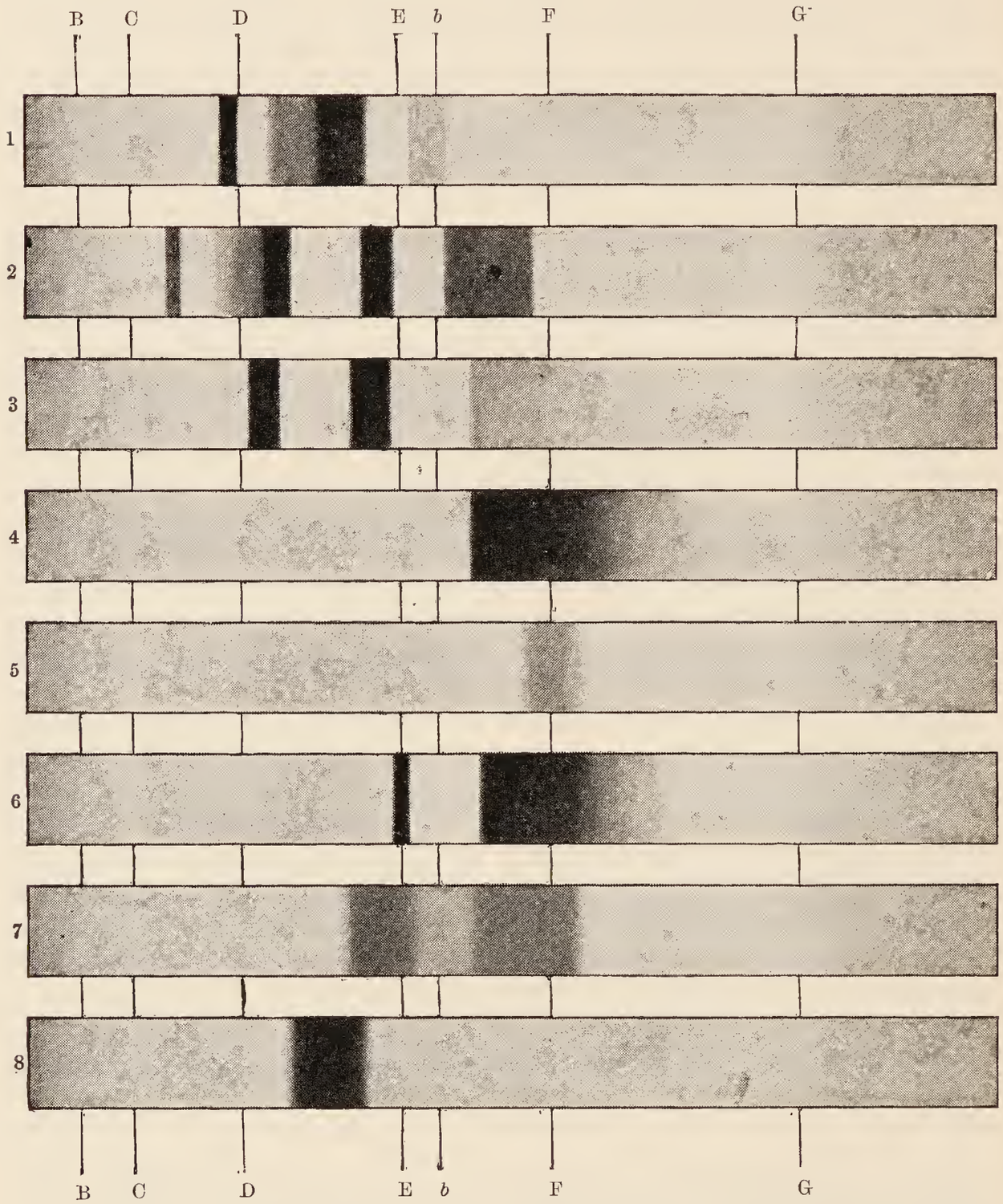


FIG. 63.—Chart of absorption spectra: 1, acid hæmatoporphyrin; 2, alkaline hæmatoporphyrin; 3, hæmatoporphyrin as found sometimes in urate sediments; 4, acid urobilin, concentrated; 5, acid urobilin, dilute; 6, the E band spectrum of urobilin; 7, uroerythrin; 8, urorosein concentrated—on dilution the band shrinks rapidly from redward end. (After F. G. Hopkins.)

Caustic alkali is added to the urine; this causes a precipitate of phosphates, which carries down the pigment with it: the pigment may be dissolved out with chloroform. The chloroform is evaporated, the residue washed with alcohol, and finally dissolved in acidified alcohol. Urines rich in the pigment yield it easily to acetic ether or to amylic alcohol.

When the urine is sufficiently rich in the pigment, the bands shown are those of alkaline hæmatoporphyrin (fig. 63, spectrum 2). On adding sulphuric acid the spectrum of acid hæmatoporphyrin is seen (fig. 63, spectrum 1). Occasionally urate sediments are pigmented with a form of the pigment which shows a two-banded spectrum, very like that of oxyhæmoglobin (fig. 63, spectrum 3); by treatment with dilute mineral acids this changes immediately to the spectrum of acid hæmatoporphyrin.

5. **Chromogens in Urine.**—In addition to the chromogens of urobilin and hæmatoporphyrin alluded to in the foregoing paragraphs there are others, of which the following may be mentioned:—(*a*) *Indoxyl.*—The origin of this substance from indole is mentioned on p. 133. It is easily oxidised to indigo-blue or indigo-red.

$$2C_6H_4{<}\overset{C.OH}{HN}{>}CH + O_2 = C_6H_4{<}\overset{CO}{NH}{>}C:C{<}\overset{CO}{NH}{>}C_6H_4 + 2H_2O.$$

[indoxyl] [indigo-blue]

Indigo-red is isomeric with indigo-blue, its structural formula being $C_6H_4{<}\overset{CO}{NH}{>}C:C{<}\overset{C.OH}{C_6H_4}{>}N$. It is very rare for the urine to be actually pigmented with indigo, for the urinary indoxyl is excreted as a conjugated sulphate which resists oxidation. When the urine is mixed with an equal volume of hydrochloric acid, indoxyl is liberated from the sulphate. A solution of a hypochlorite is then added drop by drop, when indigo-blue is formed, and on shaking the mixture with chloroform the indigo-blue passes into the chloroform. (Jaffé.) (*b*) *Skatoxyl.*—When skatoxyl is given by the mouth it passes into the urine, and yields skatoxyl-red on oxidation. (*c*) *Urorosein* is distinct from indigo-red. It is produced from its chromogen by the action of mineral acids. It frequently appears when urine is treated with strong hydrochloric acid and allowed to stand, but it appears more readily when an oxidising agent is added as well. It is readily soluble in amylic alcohol, but not in ether. The chromogen is precipitated by saturation with ammonium sulphate. The colour is destroyed by alkalis. It shows an absorption band between the D and E lines (fig. 63, spectrum 8).

6. **Pathological Pigments.**—The most frequently appearing of abnormal pigments are those of blood and bile. The urine may contain accidental pigments due to the use of drugs (rhubarb, senna, logwood, santonin); in carbolic acid poisoning pyrocatechin and hydrochinon are chiefly responsible for the greenish-brown colour of the urine, which increases on exposure to the air. The black or dark-brown pigment called melanin may pass into the urine in cases of melanotic sarcoma. For alcaptonuria see p. 149.

APPENDIX

HÆMACYTOMETERS

Gowers's Hæmacytometer.—The enumeration of the blood corpuscles is readily effected by the hæmacytometer of Gowers. This instrument consists of a glass slide (fig. 64, C), the centre of which is ruled into $\frac{1}{10}$ millimetre squares and surrounded by a glass rim $\frac{1}{5}$ millimetre thick. It is provided with measuring pipettes (A and B), a vessel (D) for mixing the blood with a saline solution (sulphate of soda of specific gravity 1015), a glass stirrer (E), and a guarded needle (F).

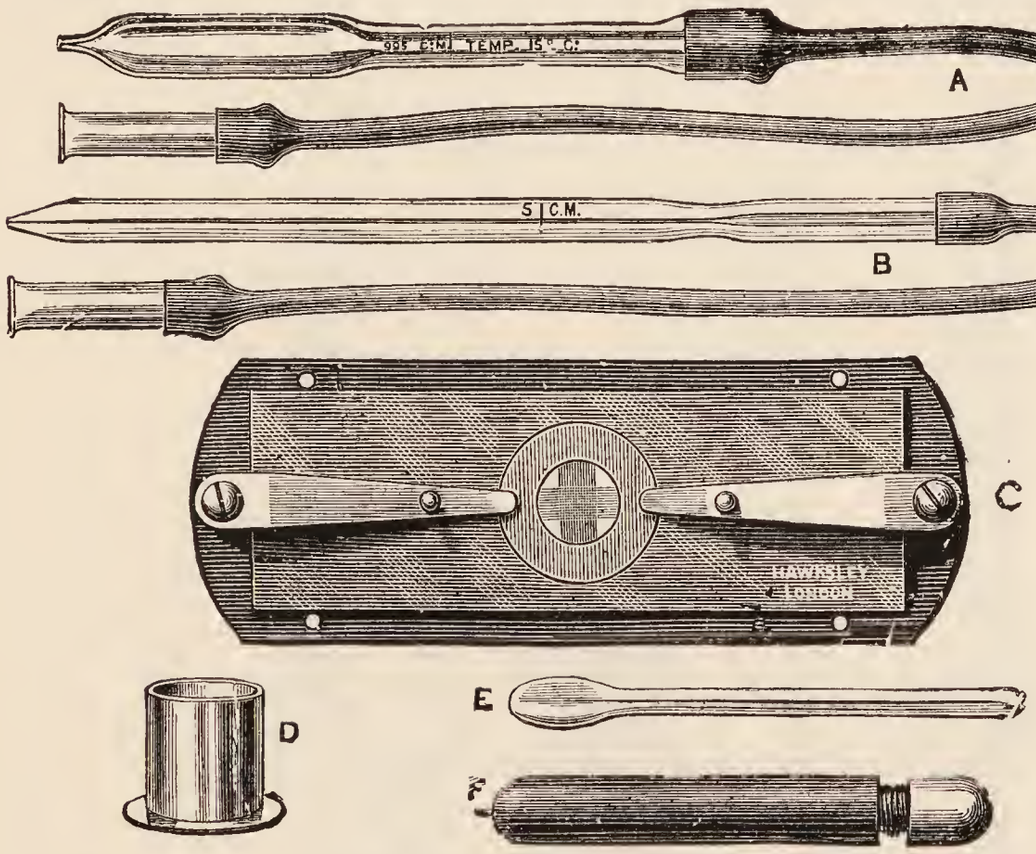


FIG. 64.—Hæmacytometer of Sir W. Gowers.

Nine hundred and ninety-five cubic millimetres of the saline solution are measured out by means of A, and then placed in the mixing jar; 5 cubic millimetres of blood are then drawn from a puncture in the finger by means of the pipette B, and blown into the solution. The two fluids are well mixed by the stirrer, and a small drop of this diluted mixture placed in the

centre of the slide C, a cover glass is gently laid on (so as to touch the drop, which thus forms a layer $\frac{1}{5}$ millimetre thick between the slide and cover glass), and pressed down by two brass springs. In a few minutes the corpuscles have sunk to the bottom of the layer of fluid, and rest on the squares. The number on ten squares is then counted, and this multiplied by 10,000 gives the number in a cubic millimetre of blood. The average number of red corpuscles in each square ought therefore in normal human blood to be 45–50.

Differential counts to show the relative proportions of the varieties of leucocytes are made in appropriately stained specimens.

Oliver's Hæmacytometer.—The following method, devised by Dr. George

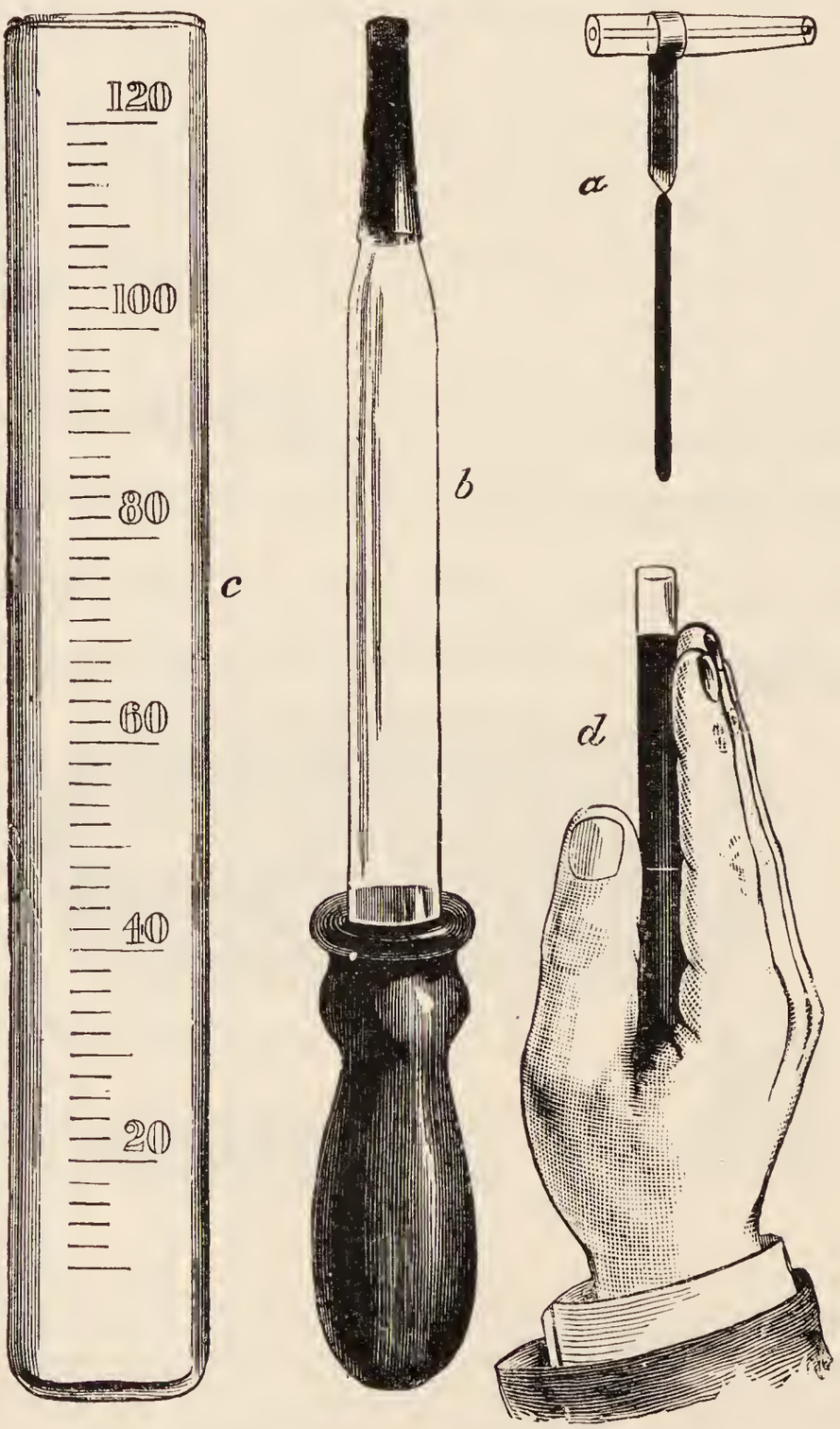


Fig. 65.—Oliver's hæmacytometer.

Oliver, is a ready way of determining the total number of corpuscles. It is, however, not possible to determine the relative proportion of red and white corpuscles by this means.

The finger is pricked, and the blood allowed to flow into the small capillary pipette (fig. 65, *a*) until it is full. This is washed out by the dropping tube *b* into a graduated flattened test-tube, *c*, with Hayem's fluid.[1] The graduations of the tube are so adjusted that with normal blood containing 5,000,000 coloured corpuscles per cubic millimetre, the light of a small wax candle placed at a distance of 9 feet from the eye in a dark room is just transmitted as a fine bright line when looked at through the tube held edgeways between the fingers (*d*) and filled up to the 100 mark of the graduation. If the number of corpuscles is less than normal, less of the diluting solution is required for the light to be transmitted; if above the normal, more of the Hayem's fluid must be added. The tube is graduated, so as to indicate in percentages the decrease or increase of corpuscles per cubic millimetre as compared with the normal standard of 100 per cent.

HÆMOGLOBINOMETERS

Gowers's Hæmoglobinometer.—The apparatus consists of two glass tubes, C and D, of the same size. D contains glycerin jelly tinted with carmine to a standard colour—viz., that of normal blood diluted 100 times with distilled water. The finger is pricked and 20 cubic millimetres of blood are measured

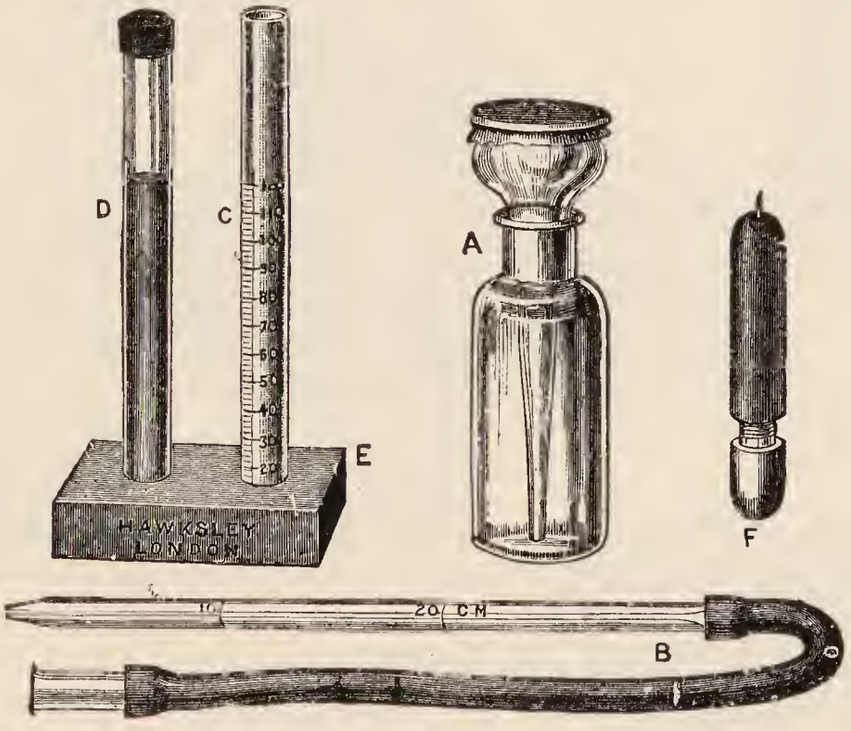


FIG. 66.—Hæmoglobinometer of Sir W. Gowers.

out by the capillary pipette, B. This is blown out into the tube C, and diluted with distilled water, added drop by drop from the pipette stopper of the bottle, A, until the tint of the diluted blood reaches the standard colour. The tube C is graduated into 100 parts. If the tint of the diluted blood is the

[1] Sodium sulphate 5 grammes, sodium chloride 1 grm., mercuric chloride 0·5 grm., distilled water 200 c.c.

same as the standard when the tube is filled up to the graduation 100, the quantity of oxyhæmoglobin in the blood is normal. If it has to be diluted more largely, the oxyhæmoglobin is in excess; if to a smaller extent, it is less than normal. If the blood has, for instance, to be diluted up to the graduation 50, the amount of hæmoglobin is only half what it ought to be—50 per cent. of the normal—and so for other percentages.

Haldane's Hæmoglobinometer is more frequently used. Instead of tinted gelatin, the standard of comparison is a sealed tube filled with a solution of carbonic oxide hæmoglobin of known strength. This keeps unchanged for years. A stream of coal gas is passed through the blood to be examined. This converts all the hæmoglobin present into carboxyhæmoglobin; this is then diluted with water to match the standard.

Von Fleischl's Hæmometer.—The apparatus (fig. 67) consists of a stand bearing a white reflecting surface (S) and a platform. Under the platform is

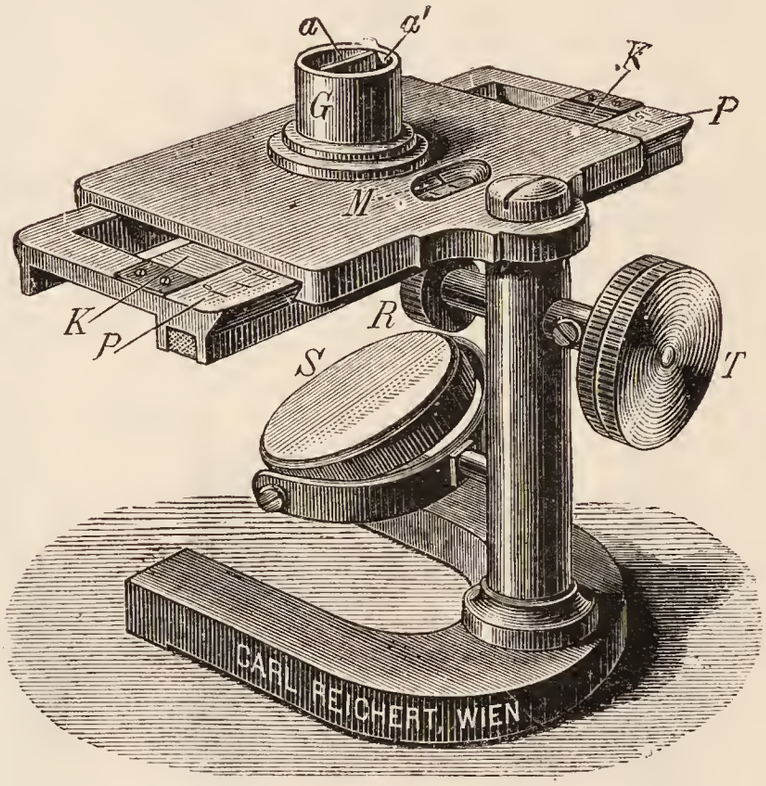


Fig. 67.—Von Fleischl's hæmometer.

a slot carrying a glass wedge stained red (K) and moved by a wheel (R). On the platform is a small cylindrical vessel divided vertically into two compartments, *a* and *a'*.

Fill with a pipette the compartment *a'* over the wedge with distilled water. Fill about a quarter of the other compartment (*a*) with water.

Prick the finger and fill the short capillary pipette provided with the instrument with blood. Dissolve this in the water in compartment *a*, and fill it up with distilled water. Having arranged the reflector (S) to throw *artificial* light vertically through both compartments, look down through them, and move the wedge of glass by the milled head (T) until the colour in the two is identical. Read off the scale, which is so constructed as to give the percentage of hæmoglobin.

Oliver's Hæmoglobinometer.—This method consists in comparing a speci-

men of blood, suitably diluted with water in a shallow white palette, with a number of standard tests very carefully prepared by the use of

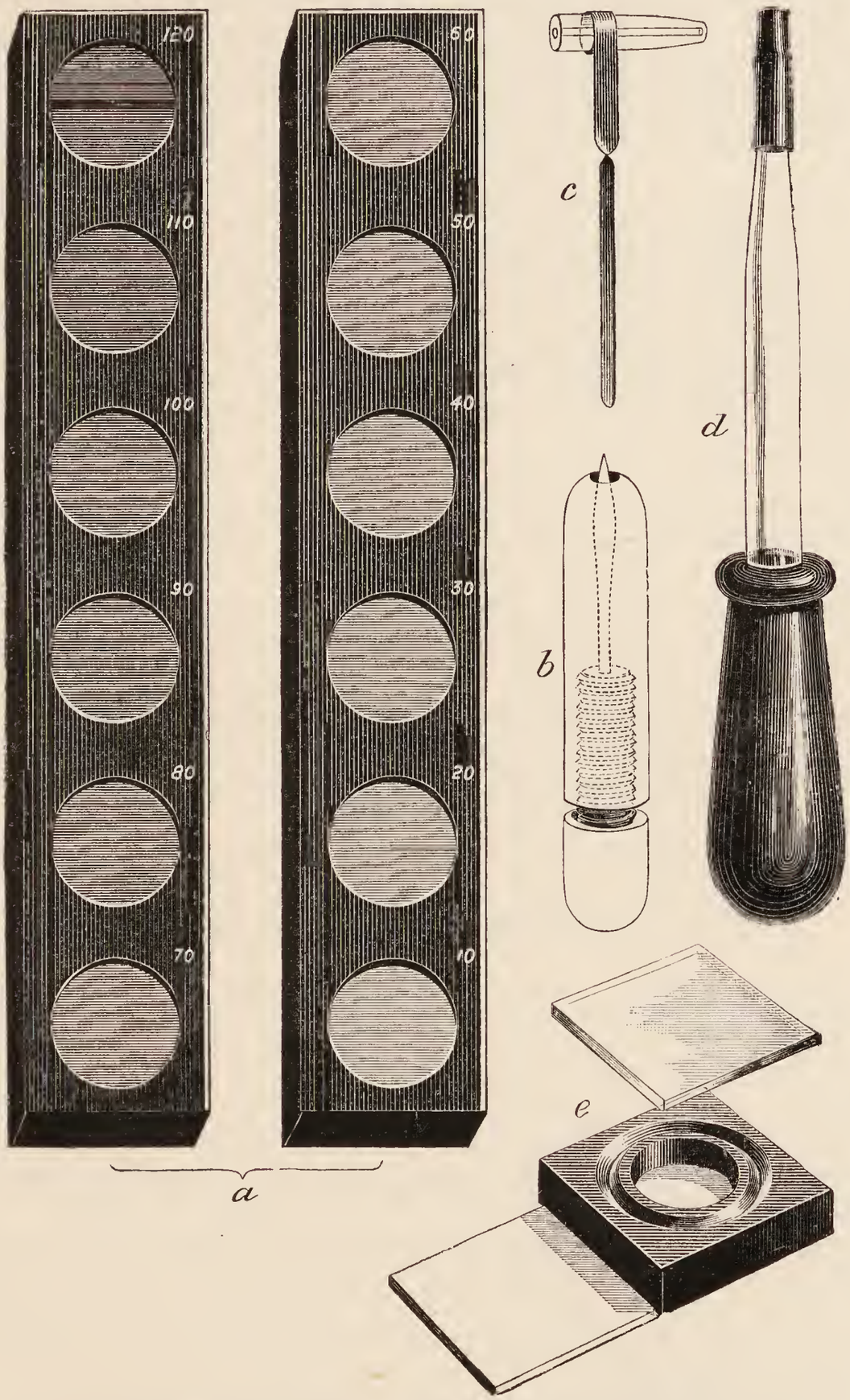


FIG. 68.—Oliver's hæmoglobinometer: *a*, standard gradations; *b*, lancet; *c*, capillary measuring pipette; *d*, mixing pipette; *e*, blood cell and cover glass.

Lovibond's coloured glasses. The capillary pipette *c* (fig. 68) is first filled with blood obtained by pricking the finger. This is washed with water

by the mixing pipette *d* into the blood cell *e*; the cell is then just filled with water, and the blood and water thoroughly mixed by the handle of *c* being used as a stirrer. The cover glass is then adjusted, when a small bubble should form, a clear sign that the cell has not been overfilled. The cell is then placed by the side of the standard gradations, and the eye quickly recognises its approximate position on the scale. The camera tube provided with the instrument will more accurately define it. Artificial light should be used.

If it is proved that the blood solution is matched in depth of colour by one of the standard grades, the observation is at an end; but if the tint is higher than one grade, but lower than another, the blood cell is placed opposite to the former, and riders (not shown in the illustration) are added to complete the observation. The standard gradations are marked in percentages, 100 per cent. being taken as normal.

'Worth' of the Corpuscles.—If the percentage of hæmoglobin is 100, and the percentage number of corpuscles is 100 also, then the quotient $\frac{100}{100} = 1$ is taken as the normal. This varies in health from 0·95 to 1·05 in men, and from 0·9 to 1 in women. This quotient has been termed the worth of the corpuscle.

Specific Gravity of Blood.—Of the numerous methods introduced for taking the specific gravity of fresh blood, that of Hammerschlag is the simplest. A drop of blood from the finger is placed in a mixture of chloroform and benzene. If the drop falls, add chloroform till it just begins to rise; if the drop rises, add benzene till it just begins to fall. The fluid will then be of the same specific gravity as the blood. Take the specific gravity of the mixture in the usual way with an accurate hydrometer.

Schmalz's capillary pycnometer is more accurate.

POLARISATION OF LIGHT

If an object, such as a black dot on a piece of white paper, be looked at through a crystal of Iceland spar, two black dots will be seen; and if the crystal be rotated, one black dot will move round the other, which remains stationary. That is, rays of light entering such a crystal are split into two rays, which travel through the crystal with different velocities, and consequently one is more refracted than the other. One ray travels just as it would through glass; this is the *ordinary ray*, the ray which gives the stationary image; the other ray gives the movable image when the crystal is rotated; the ordinary laws of refraction do not apply to it, and it is called the *extraordinary ray*. Both rays are of equal brilliancy. In one direction, however, that of the optic axis of the crystal, a ray of light is transmitted without double refraction.

Ordinary light, according to the wave theory, is due to vibrations occurring in all planes transversely to the direction of the propagation of the wave. Light is said to be plane polarised when the vibrations take place all in one plane. The two rays produced by double refraction are both polarised, one in one plane, the other in a plane at right angles to this one. Doubly refract-

ing bodies are called *anisotropous*; singly refracting bodies, *isotropous*. The effect of polarisation may be very roughly illustrated by a model.

If a string be stretched as in the figure, and then touched with the finger, it can be made to vibrate, and the vibrations will be free to occur from above down, or from side to side, or in any intermediate position. If, however, a disc with a vertical slit be placed on the course of the string, the vibrations will be all obliged to take place in a vertical plane, any side to side movement being stopped by the edges of the slit [1] (fig. 69).

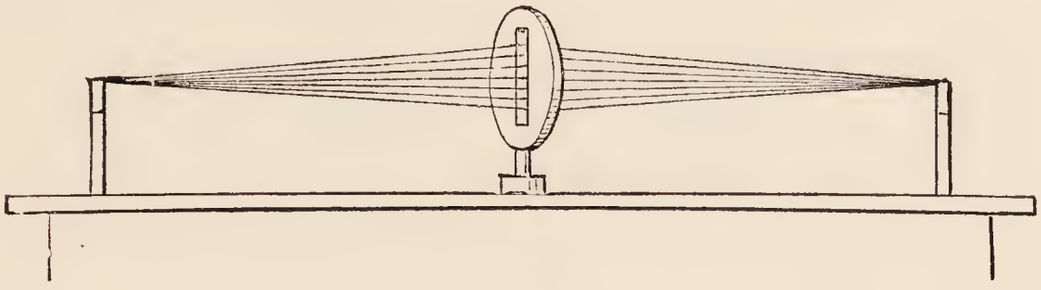


FIG. 69

Light can be polarised not only by the action of crystals, but by reflection from a surface at an angle which varies for different substances (glass 54° 35′, water 52° 45′, diamond 68°, quartz 57° 32′, &c.). It is also found that certain non-crystalline substances, like muscle, cilia, &c., are doubly refracting.

Nicol's Prism is the *polariser* usually employed in polariscopes; it consists of a rhombohedron of Iceland spar divided into two by a section through its obtuse angles. The cut surfaces are polished and cemented together in their former position with Canada balsam. By this means the

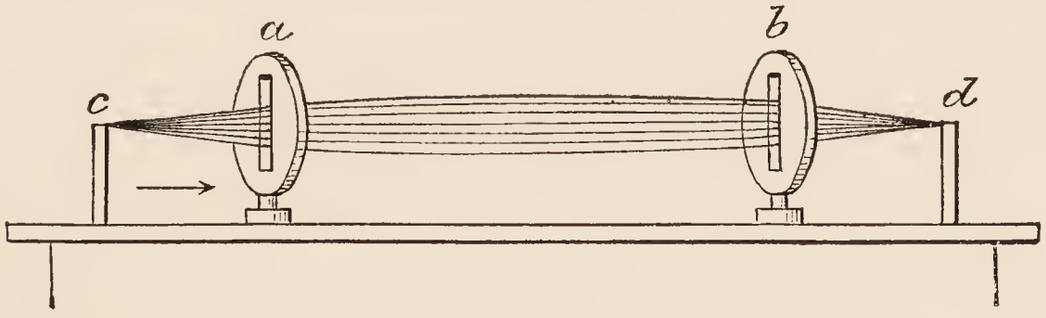


FIG. 70.

ordinary ray is totally reflected through the Canada balsam; the extra-ordinary ray passes on and emerges in a direction parallel to the entering ray. In this polarised ray there is nothing to render its peculiar condition visible to the naked eye; but if the eye is aided by a second Nicol's prism, which is called the *analyser*, it is possible to detect the fact that it is polarised.

This may be again illustrated by reference to our model (fig. 70).

Suppose that the string is made to vibrate, and that the waves travel in the direction of the arrow. From the fixed point *c* to the disc *a*, the string

[1] Such a model is, of course, imperfect; it does not, for instance, represent the splitting of the ray into two, and moreover the polarisation takes place on each side of the slit; whereas, in regard to light, it is only the rays on one side of a polariser, viz. those that have passed through it, which are polarised.

is theoretically free to vibrate in any plane;[1] but after passing through the vertical slit in *a*, the vibrations must all be vertical also; if a second similar disc *b* be placed further on, the vibrations will also pass on freely to the other extremity of the string *d*, if as in the figure (fig. 70) the slit in *b* be also placed vertically. If, however, *b* be so placed that its slit is horizontal (fig. 71), the vibrations will be extinguished on reaching *b*, and the string between *b* and *d* will be motionless.

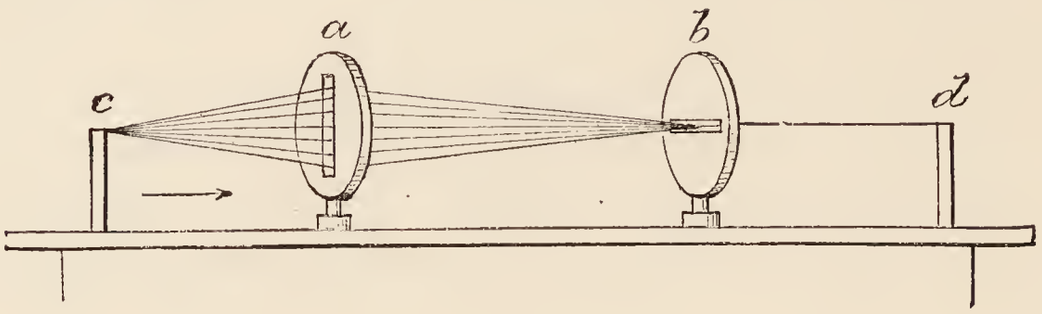


Fig. 71.

c here represents a source of light; the vibrations of the string represent the undulations which by the Nicol's prism *a* are polarised so as to occur in one plane only; if the second Nicol's prism or the analyser *b* is parallel to the first, the vibrations will pass on to the eye, which is represented by *d*; but if the planes of the two nicols are at right angles, the vibrations allowed to pass through the first are extinguished by the second, and so no light reaches the eye. In intermediate positions, *b* will allow only some of the light to pass through it. It must be clearly understood that a Nicol's prism contains no actual slits, but the arrangement of its molecules is such that their action on the particles of æther may be compared to the action of slits in a diaphragm to vibrations of more tangible materials than æther.

The Polarising Microscope consists of an ordinary microscope with certain additions; below the stage is the polarising nicol; in the eye-piece is the analysing nicol; the eye-piece is so arranged that it can be rotated; thus the directions of the two nicols can be made parallel, and then the field is bright; or crossed, and then the field is dark. The stage of the microscope is arranged so that it can also be rotated.

The polarising microscope is used to detect doubly-refracting substances. Let the two nicols be crossed, so that the field is dark; interpose between the two, that is, place upon the stage of the microscope, a doubly-refracting plate of which the principal plane is parallel to the first prism or polariser; the ray from the first prism is unaffected by the plate, but will be extinguished by the second; the field therefore still remains dark. If the plate is parallel to the second nicol the field is also dark; but in any intermediate position, the light will be transmitted by the second nicol. In other words, if between two crossed nicols, which consequently appear dark, a substance be interposed which in certain positions causes the darkness to give place to illumination, that substance is doubly refractive. How this takes place may be explained as follows:

Let N_1N_1 (fig. 72) represent the direction of the principal plane of the first nicol, and N_2N_2 that of the second. They are at right angles, and so

[1] The imperfection of the model has been explained in the preceding footnote.

the ray transmitted by the first will be extinguished by the second. Let PP represent the principal plane of the interposed doubly-refractive plate. The extraordinary ray transmitted by N_1N_1 vibrates in the plane N_1N_1, and falls obliquely on the plate PP; it is by this plate itself split into two rays, an ordinary and an extraordinary one, at right angles to one another, one vibrating in the plane PP, the other in the plane P^1P^1. These two rays meet the second nicol, which can only transmit vibrations in the plane N_2N_2. The vibrations in PP can be resolved into a vibration in N_1N_1 and a vibration in N_2N_2; the former is extinguished, the latter transmitted. Similarly the vibration in P^1P^1 can be resolved into two sub-rays in N_1N_1 and N_2N_2 respectively, the latter only being transmitted. The illumination is thus due to two sub-rays, one of the vibrations in PP, the other of those in P^1P^1 which have been made to vibrate in N_2N_2.

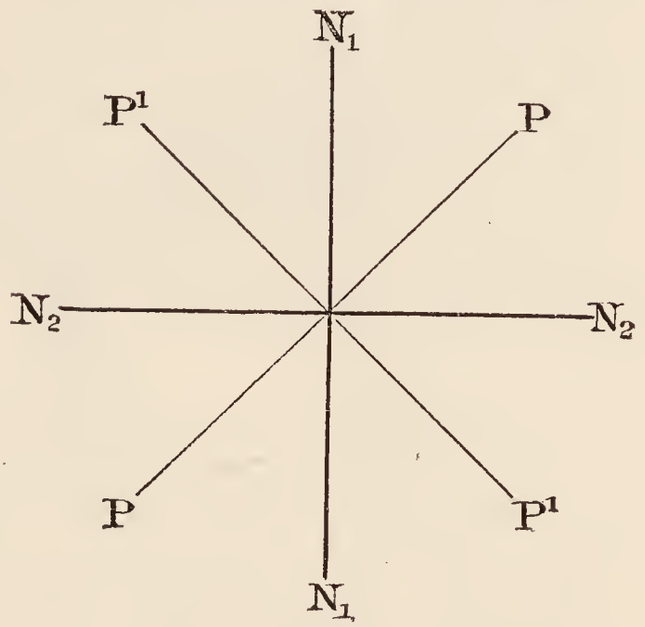


FIG. 72.

Now, although these two sub-rays vibrate in the same plane, they are of different velocities; hence the phases of the vibrations do not coincide, and thus the phenomena of interference are obtained. If we have two sets of vibrations fused, the crest of one wave may coincide with the crest of the other, in this case the wave will be higher; or the crest of one may coincide with the hollow of the other, that is, the undulation would be extinguished; in other intermediate cases, the movement would be interfered with, either helped or hindered, more or less. Interference in the case of many kinds of doubly-refracting substances (Iceland spar is in this an exception) shows itself in the extinction of certain rays of the white light, and the light seen through the second nicol is white light *minus* the extinguished rays; those extinguished and those transmitted will together form white light, and are thus complementary. Moreover, the rays extinguished in one position of the plate will be transmitted in one at right angles and *vice versa*; thus a crystal showing these phenomena of *pleochromatism*, as it is termed, will transmit one colour in one position, and the complementary colour in a position at right angles to the first; blue and yellow, and red and green, are the pairs of colours most frequently seen in this way.

Rotation of the Plane of Polarisation.—Certain crystals such as those of quartz, and certain fluids such as the essence of turpentine, aniseed, &c., and solutions of certain substances like sugar, and albumin, have the power of rotating the plane of polarised light to the right or left. The polarisation of light that is produced by a quartz crystal is different from that produced by a rhombohedron of Iceland spar. The light that passes through the latter is plane polarised; the light that passes through the former (quartz) is circularly polarised, *i.e.* the two sub-rays are made up of vibrations

which occur not in a plane, but are curved. The two rays are circularly polarised in opposite directions, one describing circles to the left, the other to the right; these unite on issuing from the quartz plate; and the net result is a plane polarised ray with the plane rotated to right or left according as the right circularly polarised ray or the left proceeded through the quartz with the greater velocity. There are two kinds of quartz, one which rotates the plane to the right (dextro-rotatory), the other to the left (lævo-rotatory).

Gordon explains this by the following mechanical illustration. Ordinary light may be represented by a wheel travelling in the direction of its axle, and the vibrations composing it executed along any or all of its spokes (*a*). If the vibrations all take place in the same direction, *i.e.* along one spoke, and the spoke opposite to it (*b*), the light is said to be plane polarised. The two spokes as they travel along in the direction of the arrow will trace out a plane (see fig. 73) between *b* and *b'*. If this polarised beam be made to

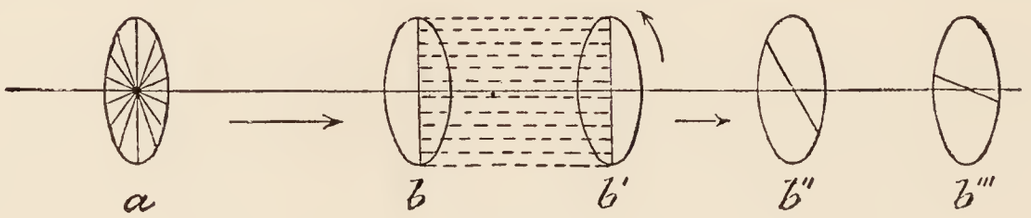


FIG. 73.

travel now through a solution of sugar, the net result is that the plane so traced out is twisted or rotated; the two spokes, as in *bb'*, do not trace out a plane, but we must consider that they rotate as they travel along, as though guided by a spiral or screw thread cut on the axis, so that after a certain distance the vibrations take place as in *b''*; later in *b'''*, and so on. This effect on polarised light is due to the molecules in solution, and the amount of rotation will depend on the strength of the solution, and on the length of the column of the solution through which the light passes, or, in the case of a quartz plate, on its thickness.

If a plate of quartz be interposed between two nicols, the light will not be extinguished in any position of the prisms, but will pass through various colours as rotation is continued. The rotation produced for different kinds of light being different, white light is split into its various constituent colours; and the angle of rotation that causes each colour to disappear is constant for a given thickness of quartz plate, being least for the red and greatest for the violet. These facts are made use of in the construction of polarimeters. Polarimeters are instruments for determining the strength of solutions of sugar, albumin, &c., by the direction and amount of rotation they produce on the plane of polarised light. They are often called saccharimeters, as they are specially useful in the estimation of sugar.

POLARIMETERS

Soleil's Saccharimeter.—This instrument (see fig. 74) consists of a Nicol's prism, *d*, called the polariser: this polarises the light entering it, and the polarised beam then passes through a quartz plate (*b* in fig. 74), 3·75 mm.

thick, one half of which (*d* in fig. 75) is made out of dextro-rotatory, the other half (*g* in fig. 75) of lævo-rotatory quartz.

The light then passes through the tube containing the solution in the position of the dotted line in fig. 74, then through a quartz plate cut perpendicularly to its axis (*q* in fig. 75), then through an arrangement called a compensator (*r* in fig. 75), then through a second nicol called the analyser, and lastly through a telescope (L in fig. 75).

The compensator consists of two quartz prisms (RR′, fig. 75) cut perpendicularly to the axis, but of contrary rotation to the plate just in front of

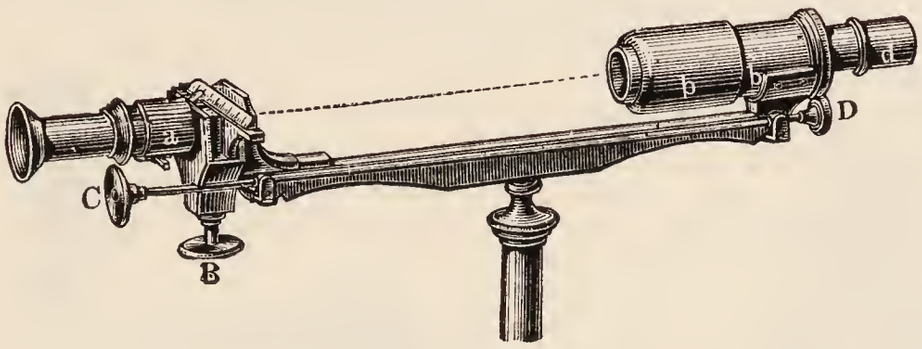


Fig. 74.—Soleil's saccharimeter.

them. These are wedge-shaped, and slide over each other, the sharp end of one being over the blunt end of the other. By a screw the wedges may be moved from each other, and this diminishes the thickness of quartz interposed; if moved towards each other the amount of quartz interposed is increased.

The effect of the quartz plate (*d*, *g*) next to the polariser (*i* in fig. 75) is to give the polarised light a violet tint when the two nicols are parallel to each other. But if the nicols are not parallel, or if the plane of the polarised light has been rotated by a solution in the tube, one half the field will change

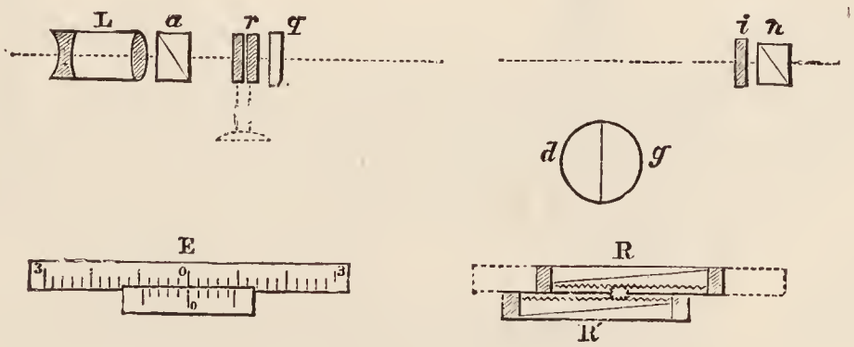


Fig. 75.—Diagram of optical arrangements in Soleil's saccharimeter.

in colour to the red end, the other to the violet end of the spectrum, because the two halves of the quartz act in the opposite way.

The instrument is first adjusted with the compensator at zero, and the nicols parallel, so that the whole field is of one colour. The tube containing the solution is then interposed; and if the solution is optically inactive the field is still uniformly violet. But if the solution is dextro-rotatory the two halves will have different tints; a certain thickness of the compensating quartz plate which is lævo-rotatory must be interposed to make the tint of the two halves of the field equal again; the thickness so interposed can be read off in amounts corresponding to degrees of a circle by means of a vernier

and scale (E in fig. 75) worked by the screw which moves the compensator. If the solution is lævo-rotatory, the screw must be turned in the opposite direction.

Zeiss's polarimeter is in principle much the same as Soleil's; the chief difference is that the rotation produced by the solution is corrected, not by a quartz compensator, but by actually rotating the analyser in the same direction, the amount of rotation being directly read off in degrees of a circle.

Laurent's polarimeter is a more valuable instrument. Instead of using daylight, or the light of a lamp, monochromatic light (a sodium flame pro-

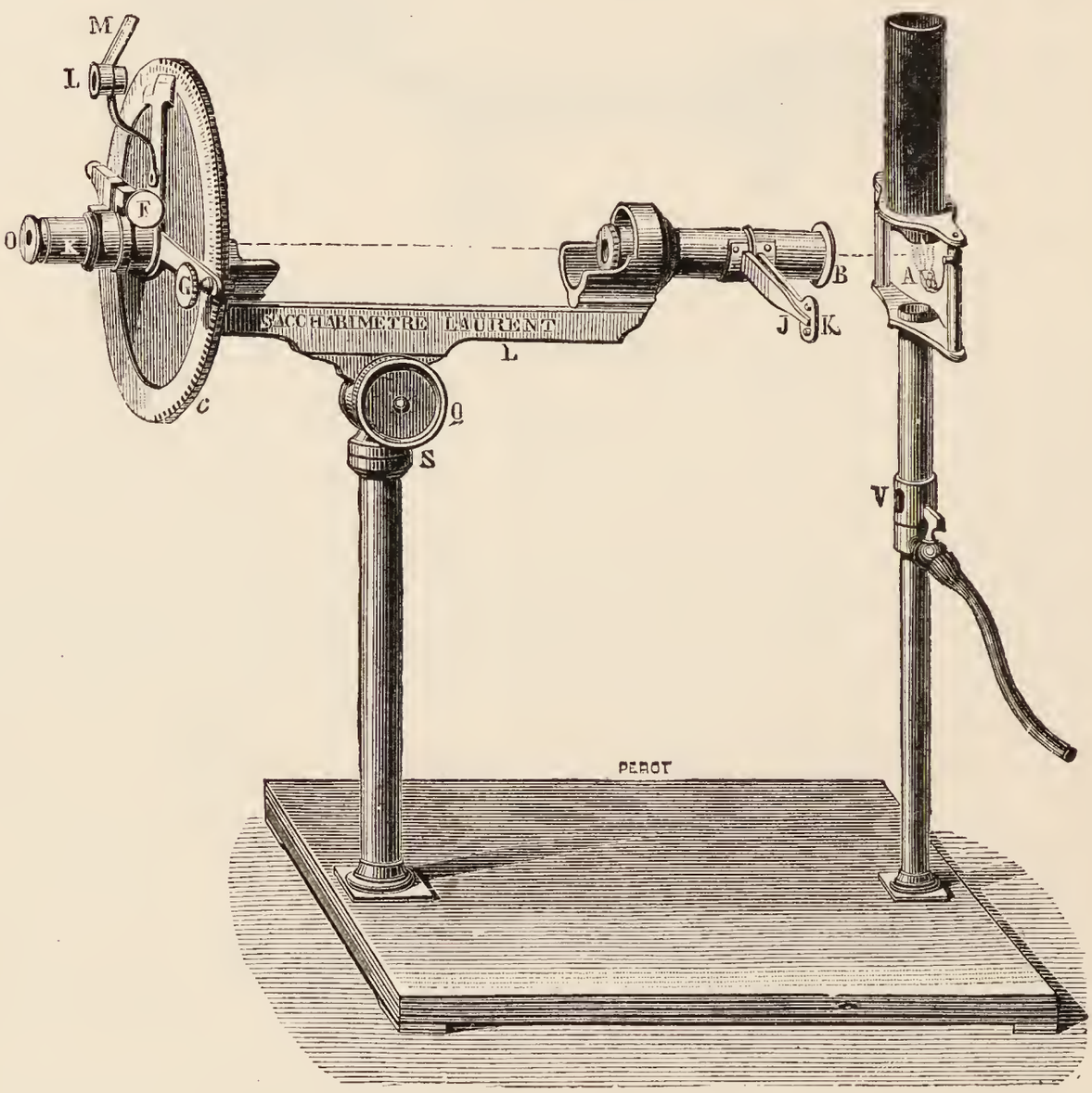


FIG. 76.—Laurent's polarimeter.

duced by volatilising common salt in a colourless gas flame) is employed; the amount of rotation varies for different colours; and observations are recorded as having been taken with light corresponding to the D or sodium line of the spectrum. The essentials of the instrument are, as before, a polariser, a tube for the solution, and an analyser. The polarised light before passing into the solution traverses a quartz plate, which, however, covers only half the field, and retards the rays passing through it by half a wave-length. In the 0° position the two halves of the field appear equally

illuminated: in any other position, or if rotation has been produced by the solution when the nicols have been set at zero, the two halves appear unequally illuminated. This is corrected by means of a rotation of the analyser that can be measured in degrees by a scale attached to it.

Specific rotatory power of any substance is the amount of rotation in degrees of a circle of the plane of polarised light produced by 1 gramme of the substance dissolved in 1 c.c. of liquid examined in a column 1 decimetre long.

If a = rotation observed.
w = weight in grammes of the substance per cubic centimetre.
l = length of tube in decimetres.
$(a)_D$ = specific rotation for light with wave-length corresponding to the D line (sodium flame).

Then $(a)_D = \pm \frac{a}{wl}$.

In this formula + indicates that the substance is dextro-rotatory, − that it is lævo-rotatory.

If, on the other hand, $(a)_D$ is known, and we wish to find the value of w, then

$$w = \frac{a}{(a)_D \times l}.$$

THE SPECTRO-POLARIMETER.

This instrument is one in which a spectroscope and polarising apparatus are combined for the purpose of determining the concentration of substances

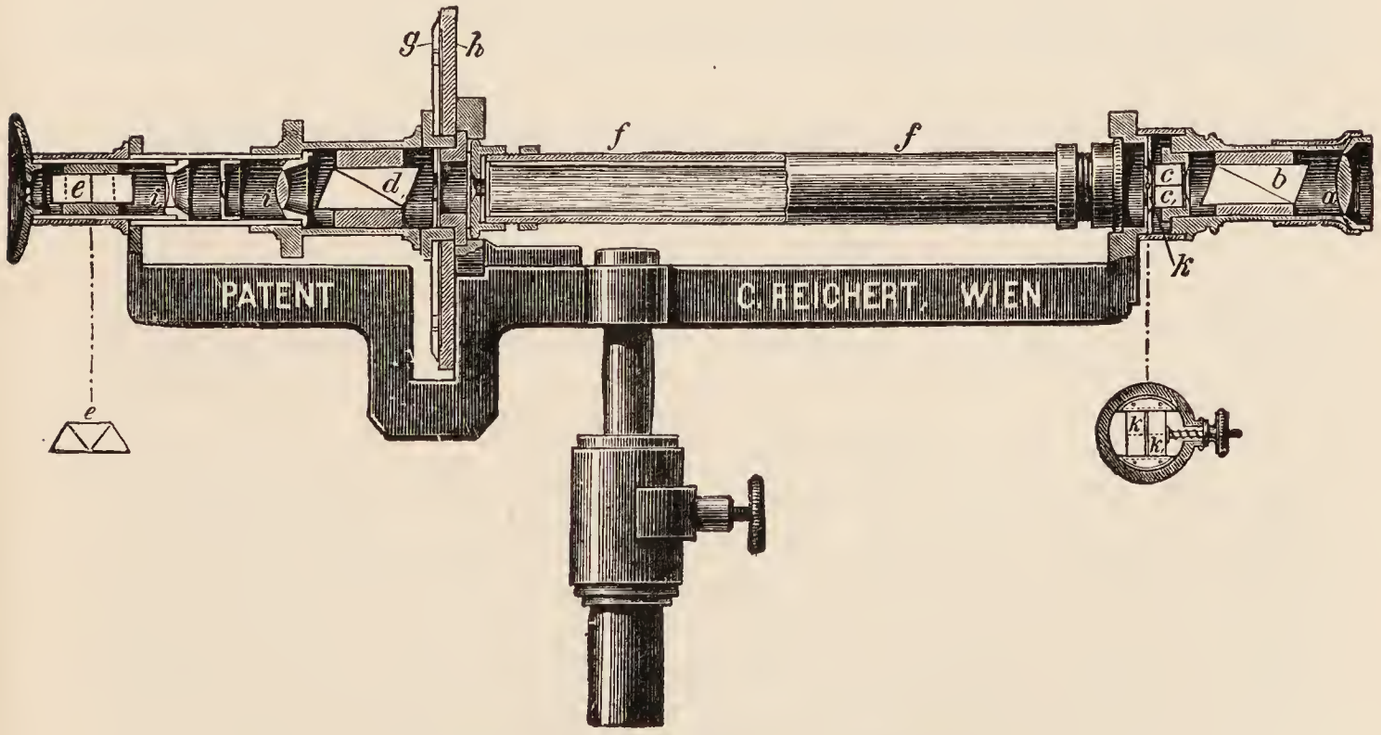


FIG. 77.—Spectro-polarimeter of v. Fleischl.

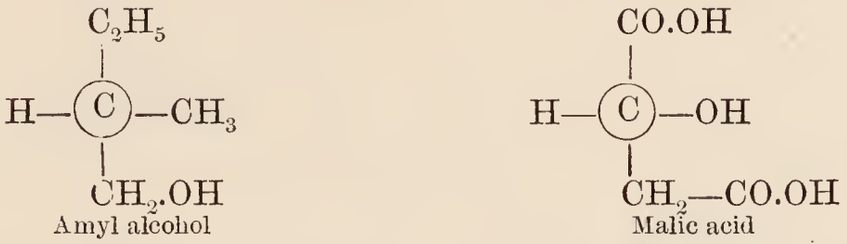
which rotate the plane of polarised light. It was invented by E. v. Fleischl for the estimation of sugar in diabetic urine. Its chief advantage is that no difficulty arises in forming a judgment as to the identity of two coloured

surfaces, as in Soleil's saccharimeter, or of two shades of the same colour, as in Laurent's instrument. The light enters at the right-hand end of the instrument, is polarised by the Nicol's prism *b*, and then passes through two quartz plates, *cc*, placed horizontally over each other. One of these plates is dextro-, the other lævo-rotatory, and they are of such a thickness (7·75 mm.) that the green rays between the E and *b* lines of the spectrum are circularly polarised through an angle of 90°, the one set passing off through the upper quartz to the left, the other through the lower to the right. The light then continues through a long tube, *ff*, which contains 15 c.c. of the solution under examination. It then passes through an analysing nicol *d*, and finally through a direct-vision spectroscope, *e*. On looking through the instrument, the tube *ff* being empty or filled with water or some other optically inert substance, two spectra are seen, one over the other, but each shows a dark band between E and *b* owing to the extinction of these rays by the circular polarisation, produced by the quartz. The analyser can be rotated: a vernier, *g*, is attached to, and moves with it, round a circular disc (seen in section at *h*) graduated in degrees. The two bands in the spectra coincide when the zeros of vernier and scale correspond. If now the tube *f* is filled with an optically active substance like sugar, the bands are shifted, one to the right, the other to the left, according to the direction of rotation of the substance in *f*. The rotation is corrected by rotating the analyser into such a position that the two bands exactly coincide once more as to vertical position. The number of degrees through which it is thus necessary to move the analyser measures the amount of rotation produced by the substance in *f*, and is a measure of the concentration of the solution. The degrees marked on the circular scale are not degrees of a circle, but an arbitrary degree of such a length that each corresponds to 1 per cent. of sugar in the given length of the column of fluid in *ff* (177·2 mm.).

RELATION BETWEEN CIRCULAR POLARISATION AND CHEMICAL CONSTITUTION

The first work in this direction was performed by Pasteur, and it was his publications on this subject that brought him into prominence. He found that racemic acid, which is optically inactive, can be decomposed into two isomerides, one of which is common tartaric acid which is dextro-rotatory, and the other tartaric acid differing from the common variety in being lævo-rotatory. The salts of tartaric acid usually exhibit hemihedral faces, while those of racemic acid are holohedral. Pasteur found that, although all the tartrate crystals were hemihedral, the hemihedral faces were situated on some crystals to the right, and on others to the left hand of the observer, so that one formed, as it were, the reflected image of the other. These crystals were separated, purified by recrystallisation, and those which exhibited dextro-hemihedry possessed dextro-rotatory power, whilst the others were lævo-rotatory. Pasteur further showed that if the mould *Penicillium glaucum* is grown in a solution of racemic acid, dextro-tartaric acid first disappears, and the lævo- acid alone remains. The subject remained in this condition for many years; it was, however, conjectured that probably there is some molecular condition corresponding to the naked-eye crystalline appearances which

produces the opposite optical effects of various substances. What this molecular structure is, was pointed out independently by two observers—Le Bel in Paris, and Van't Hoff in Holland—who published their researches within a few days of each other. They pointed out that all optically active bodies contain one or more asymmetric carbon atoms, *i.e.* one or more atoms of carbon connected with four dissimilar groups of atoms, as in the following examples:—



Amyl alcohol — Malic acid

The question, however, remained—do all substances containing such atoms rotate the plane of polarised light? It was found that they do not; this is explained by Le Bel by supposing that these, like racemic acid, are compounds of two molecules—one dextro-, the other lævo-rotatory; that this

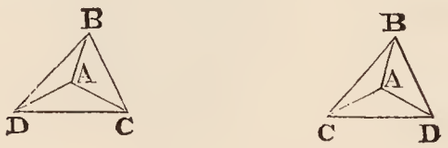


FIG. 78.

was the case was demonstrated by growing moulds, the fermenting action of which is to separate the two molecules in question. Then the other question—how is it that two isomerides, which in chemical characteristics, in graphic as well as empirical formulæ, are precisely alike, differ in optical properties?—is explained ingeniously by Van't Hoff. He compares the carbon atom to a tetrahedron with its four dissimilar groups, A, B, C, D, at the four corners. The two tetrahedra represented in fig. 78 appear at first sight precisely alike; but if one be superimposed on the other, C in one and D in the other could never be made to coincide. This difference cannot be represented in any other graphic manner, and probably represents the difference in the way the atoms are grouped in the molecule of right- and left-handed substances respectively.

MERCURIAL AIR PUMPS

Pflüger's Pump.—*l* is a large glass bulb filled with mercury; from its lower end a straight glass tube, *m*, about 3 feet long, extends, which is connected by an indiarubber tube, *n*, with a reservoir of mercury, *o*, which can be raised or lowered as required, by a simple mechanical arrangement. From the upper end of the bulb, *l*, a vertical tube passes; above the stopcock, *k*, this has a horizontal branch, which can be closed by the stopcock, *f*. The vertical part is continued into the bent tube, which dips under mercury in the trough, *h*. A stopcock, *j*, is placed on the course of this tube. Beyond *f* the horizontal tube leads into a large double glass bulb, *a b*; a mercurial gauge, *e*, and a drying-tube, *d*, filled with pieces of pumice-stone moistened with sulphuric acid, are interposed. *a* is called the blood-bulb, and the

blood is brought into it by the tube *c*; the gases, as they come off, cause the blood to froth, and the bulb, *b*, is called the froth-chamber, as it intercepts the froth, preventing it from passing into the rest of the apparatus.

The pump is used in the following way: *l* is filled with mercury, the level in *l* and *o* being the same; *k* is closed; *o* is then lowered, and when it

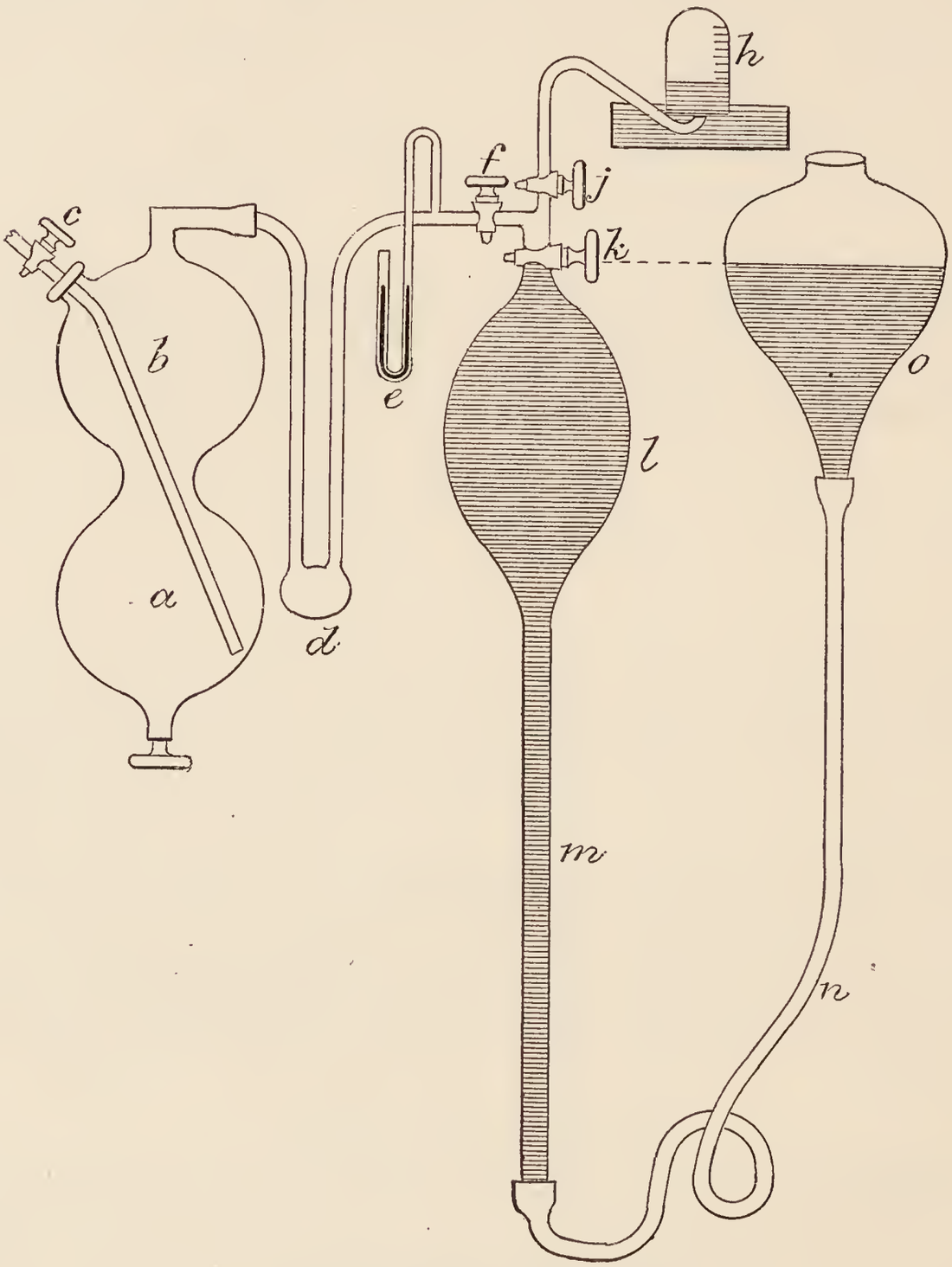


Fig. 79.—Diagram of Pflüger's pump.

is 30 inches lower than the stopcock, *k*, the mercury in *l* falls also, leaving that bulb empty: *j* being closed and *f* open, *k* is then opened, and the air in *a*, *b*, *d*, &c. rushes into the Torricellian vacuum in *l*; *f* is closed and *j* opened; the reservoir, *o*, is raised; the mercury in *l* rises also, pushing the air before it, and it bubbles out into the atmosphere through the mercury (the tube, *h*, is not at this stage in position). When *l* is full of mercury, *k* and *j* are once

more closed and *o* is again lowered; when *l* is thus rendered once more a vacuum, *k* and *f* are opened and more of the air remaining in *a*, *b*, *d*, &c. rushes into the vacuum; *f* is closed, *j* is opened, and this air is expelled as before. The process is repeated as often as is necessary to make *a*, *b*, *d*, &c. as complete a vacuum, as indicated by the mercury in the gauge, *e*, as is obtainable.

a being now empty and the stopcock, *f*, closed, blood is introduced by the tube *c*; it froths and gives forth all its gases, especially if heated to 40-45° C. In the case of serum, acid has to be added to disengage the more firmly combined carbonic acid.[1] The bulb, *l*, is once more rendered a vacuum, and *k* and *f* are opened, *j* being closed. The gas from *a* and *b* rushes into the bulb *l*, being dried as it passes through *d*; *f* is then closed and *j* opened; the reservoir *o* is raised, and as the mercury in *l* rises simultaneously, it pushes the gases into the cylinder, *h*, which is filled with mercury and inverted over the end of the bent tube. This gas can be consequently analysed. By alternately raising and lowering *o*, and regulating the stopcocks in the manner already described, all the gas from the quantity of blood used can be ultimately expelled into *h*.

A good grease for the stopcocks is a mixture of two parts of vaseline to one of white wax.

Alvergniat's pump has the advantage over Pflüger's of fewer connections, and all of these are surrounded by mercury, which effectually prevents leakage; it has the disadvantage of a rather small bulb in place of *l*, and thus it is more labour to obtain a vacuum.

Leonard Hill's Pump.—This is a simpler instrument, and is sufficient for most purposes. It consists of three glass bulbs (B.B. in fig. 80), which we will call the *blood bulb*; this is closed above by a piece of tubing and a clip, *a*; this is connected by good indiarubber tubing to another bulb, *d*. Above *d*, however, there is a stopcock with two ways cut through it: one by means of which B.B. and *d* may be connected, as in the figure; and another seen in section, which unites *d* to the tube *e*, when the stopcock is turned through a right angle. In intermediate positions the stopcock cuts off all communication from *d* to all parts of the apparatus above it; *d* is connected by tubing to a receiver, R, which can be raised or lowered at will. At first the whole apparatus is filled with mercury, R being raised. Then, *a* being closed, R is lowered, and when it is more than the height of the barometer (30 inches) below the top of B.B. the mercury falls and leaves the blood bulb empty; by lowering R still further, *d* can also be rendered a vacuum. A few drops of

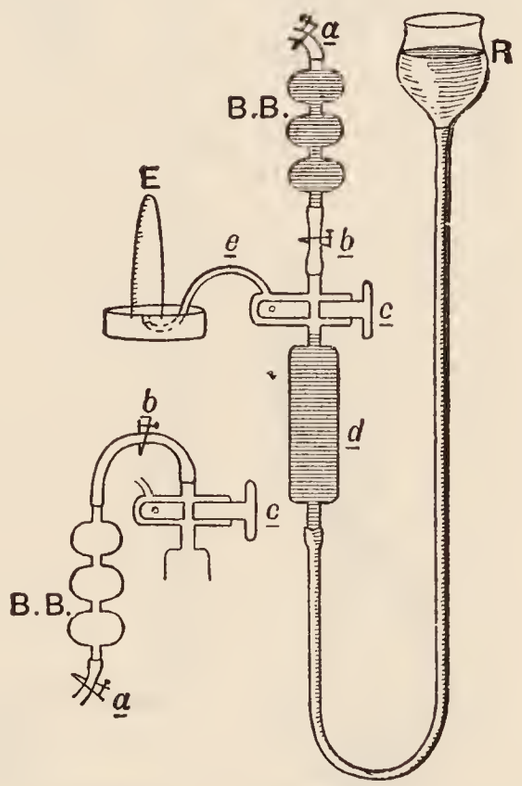


FIG. 80.—L. Hill's air-pump.

[1] Phosphoric acid is usually employed.

mercury should be left behind in B.B. B.B. is then detached from the rest of the apparatus and weighed, the clips, *a* and *b*, being tightly closed. Blood is then introduced into it by connecting the tube with the clip *a* on it to a cannula filled with blood inserted in an artery or vein of a living animal. Enough blood is withdrawn to fill about half of one of the bulbs. This is defibrinated by shaking it with the few drops of mercury left in the bulb. It is then weighed again; the increase of weight gives the amount of blood which is being investigated. B.B. is then once more attached to the rest of the apparatus, hanging downwards, as in the side drawing in fig. 80, and the blood gases boiled off; these pass into *d*, which has been made a vacuum; and then, by raising R again, the mercury rises in *d*, pushing the gases in front of it through the tube *e* (the stopcock being turned in the proper direction) into the eudiometer, E, which has been filled with, and placed over, mercury. The gas can then be measured and analysed.

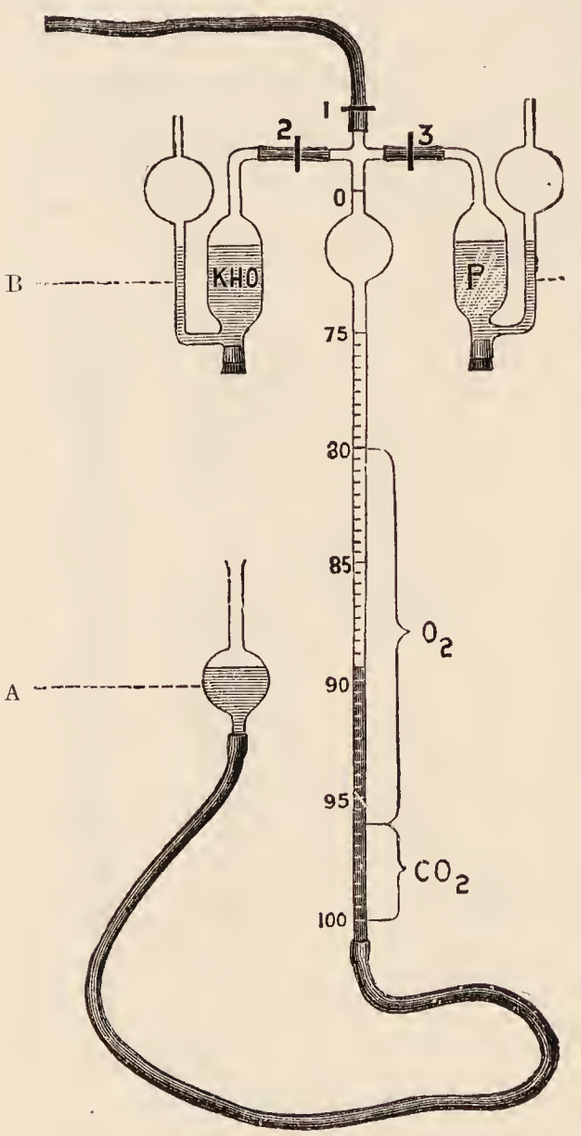


FIG. 81.—Waller's apparatus for gas analysis.

ANALYSIS OF GASES

Waller's modification of Zuntz's more complete apparatus will be found very useful in performing gas analysis, say of the expired air or blood gases: a 100 c.c. measuring tube graduated in tenths of a cubic centimetre between 75 and 100; a filling bulb and two gas pipettes are connected up as in the diagram.

It is first charged with acidulated water up to the zero mark by raising the filling bulb A, tap 1 being open. It is then filled with 100 c.c. of expired air, the filling bulb being lowered till the fluid in the tube has fallen to the 100 mark. Tap 1 is now closed. The amount of carbonic acid in the expired air is next ascertained; tap 2 is opened, and the air is expelled into the gas pipette B, containing strong caustic potash solution, by raising the filling bulb until the fluid has risen to the zero mark of the measuring tube. Tap 2 is closed, and the air left in the gas pipette for a few minutes, during which the carbonic acid is absorbed by the potash. Tap 2 is then opened and the air drawn back into the measuring tube by lowering the filling bulb. The volume of air (*minus* the carbonic acid) is read, the filling bulb being adjusted so that its contents are at the same level as the fluid in the measuring tube. The amount of oxygen is next ascertained in a precisely similar manner by sending the air into the other gas pipette, which contains sticks of phosphorus in water, and measuring the loss of volume (due to

absorption of oxygen) in the air when drawn back into the tube. The remaining gas is nitrogen.

KJELDAHL'S METHOD OF ESTIMATING NITROGEN

This simple method can be used in connection with most substances of physiological importance. Briefly, it consists in converting all the nitrogen present into ammonia by means of sulphuric acid; then rendering alkaline with soda, and distilling over the ammonia with standard acid, the diminution in acidity of which, measures the amount of ammonia present.

The following is Bernard Dyer's modification of the original method:

About 1 gramme of the substance under investigation (or in the case of urine when one wishes to make an estimation of total nitrogen for 5 to 10 c.c. of that fluid) is placed in a round-bottomed Jena flask of about 250 c.c. capacity, and 20 c.c. of pure concentrated sulphuric acid added. The flask should be provided with a loose balloon stopper. The flask is arranged in a sloping direction over a small flame. A drop of mercury and a piece of hard paraffin about the size of a pea are also added. The mixture is heated slowly until it boils. In about twenty minutes the fluid becomes colourless or nearly so. Then 10 grammes of ignited potassium sulphate in powder are added, and the boiling continued for another 45 minutes. By this time all the nitrogen will be in combination as ammonia.

After cooling, the fluid is washed into a litre flask of Jena glass (fig. 82, A), and water added until the total volume of the fluid is about 400 c.c. 10 c.c. of a 10-per-cent. solution of sodium sulphide are added to decompose the mercury compound (mercury is originally put in to act as an oxygen carrier and hasten the action), also 10 or 12 pieces of granulated zinc to avoid bumping in the subsequent distillation, and a bit of hard paraffin to avoid frothing. Excess of 40 per cent. soda is then run in from the funnel B through the tap C. The cork of the flask A is also perforated by a long leaden tube D placed as in the figure; this leads into the flask E which contains a measured amount (50 or 100 c.c.) of standard sulphuric acid; $\frac{1}{5}$ normal acid is a convenient strength to use. The pear-shaped adapter F shown in the figure avoids any regurgitation, and its end should just dip below the surface of the acid. The mixture in the flask is now boiled for about half an hour, when all the ammonia will have distilled over: the use of a condenser around the tube D is unnecessary. The acidity of the standard acid is then determined by titrating with standard alkali, a few drops of methyl orange being added to

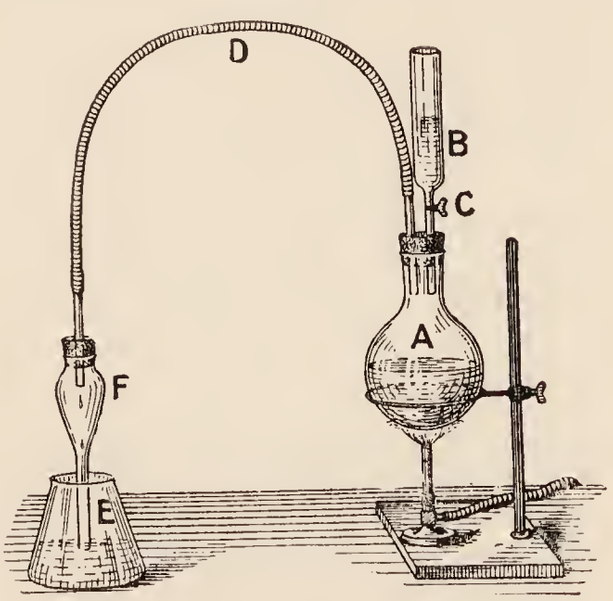


FIG. 82.—Kjeldahl's method: distilling apparatus.

act as the indicator of the end of the reaction (this gives a pink colour with acid, yellow with alkali).

Example.—Suppose 1 gramme of a nitrogenous substance is taken, and the ammonia distilled over into 100 c.c. of $\frac{1}{5}$ normal sulphuric acid (= 20 c.c. normal acid). This is then titrated with a corresponding solution of soda, and it is found that the neutral point is reached when 60 c.c. of the soda solution have been added. The other 40 c.c. must therefore have been neutralised by the ammonia derived from the substance under investigation. This 40 c.c. of acid = 8 c.c of normal acid = 8 c.c. of normal ammonia = 8 × 0·017 = 0·136 gramme of ammonia. One gramme of the substance analysed therefore yields 0·136 gramme of ammonia, and this contains 0·112 gramme of nitrogen; 100 grammes will therefore contain 11·2 grammes of nitrogen. If the strength of the acid is that just recommended, each c.c. corresponds to 0·0028092 of nitrogen.

SOLUTIONS. DIFFUSION. DIALYSIS. OSMOSIS.

The investigations of physical chemists during recent years have given us new conceptions of the meaning of the words that stand at the head of this article. I propose to state what these new conceptions are, and briefly to indicate the bearing they have on the elucidation of physiological problems.

Solutions.—Water is the fluid in which soluble materials are usually dissolved, and at ordinary temperatures it is a fluid the molecules of which are in constant movement; the hotter the water the more active are the movements of its molecules, until, when at last it is converted into steam, the molecular movements become much more energetic. Perfectly pure water consists of molecules with the formula H_2O, and these molecules undergo practically no dissociation into their constituent atoms, and it is for this reason that pure water is not a conductor of electricity.

If a substance like sugar is dissolved in the water, the solution still remains incapable of conducting an electrical current. The sugar molecules in solution are still sugar molecules; they do not undergo dissociation.

But if a substance like salt is dissolved in the water, the solution is then capable of conducting electrical currents, and the same is true for most acids, bases, and salts. These substances do undergo dissociation, and the simpler materials into which they are broken up in the water are called **ions**. Thus if sodium chloride is dissolved in water a certain number of its molecules become dissociated into sodium ions, which are charged positively with electricity, and chlorine ions, which are charged negatively with electricity. Similarly a solution of hydrochloric acid in water contains free hydrogen ions and free chlorine ions. Sulphuric acid is decomposed into hydrogen ions and ions of SO_4. The term ion is thus not equivalent to atom, for an ion may be a group of atoms, like SO_4, in the example just given.

Further, in the case of hydrochloric acid, the negative charge of the chlorine ion is equal to the positive charge of the hydrogen ion; but in the case of the sulphuric acid, the negative charge of the SO_4 ion is equal to the positive charge of two hydrogen ions. We can thus speak of monovalent, divalent, trivalent, &c. ions.

Ions positively charged are called *kat-ions* because they move towards

the kathode or negative pole; those which are negatively charged are called *an-ions* because they move towards the anode or positive pole. The following are some examples of each class:

Kat-ions. Monovalent:—H, Na, K, NH_4, &c.
Divalent:—Ca, Ba, Fe (in ferrous salts), &c.
Trivalent:—Al, Bi, Sb, Fe (in ferric salts), &c.
An-ions. Monovalent:—Cl, Br, I, OH, NO_3, &c.
Divalent:—S, Se, So_4, &c.

Roughly speaking, the greater the dilution the more nearly complete is the dissociation, and in a very dilute solution of such a substance as sodium chloride we may consider that the number of ions is double the number of molecules of the salt present.

The ions liberated by the act of dissociation are, as we have seen, charged with electricity, and when an electrical current is led into such a solution it is conducted through the solution by the movement of the ions. Substances which exhibit the property of dissociation are known as electrolytes.

The conception of electrolytes which we owe to Arrhenius is extremely important in view of the question of osmotic pressure which we shall be considering immediately; because the act of dissociation increases the number of particles moving in the solution and so increases the osmotic pressure, for in this relation the ion plays the same part as a molecule.

The liquids of the body contain electrolytes in solution, and it is owing to this fact that they are able to conduct electrical currents.

Another physiological aspect of the subject is seen in a study of the action of mineral salts in solution on living organisms and parts of organisms. Many years ago Ringer showed that contractile tissues (heart, cilia, &c.) continue to manifest their activity in certain saline solutions. Indeed, as Howell puts it, the cause of such rhythmical action is the presence of these inorganic substances in the blood or lymph which usually bathes them. In the case of the heart, the sinus, or venous end of the heart, is peculiarly susceptible to the stimulus of the inorganic salts, and the rhythmical peristaltic waves so started travel thence over the rest of the heart muscle.

Loeb and his fellow workers have confirmed these statements, but interpret them now as ionic action. Contractile tissues will not contract in pure solutions of non-electrolytes (like sugar, urea, albumin). But different contractile tissues differ in the nature of the ions which are most favourable stimuli. Thus cardiac muscle, cilia, amœboid movement, karyokinesis, cell division are all alike in requiring a proper adjustment of ions in their surroundings if they are to continue to act, but the proportions must be different in individual cases. Ions affecting the rhythmical contractions may be divided into three classes: (1) Those which produce such contractions; of these the most efficacious is Na. (2) Those which retard rhythmical contractions; for instance, Ca and K. (3) Those which act catalytically, that is, they accelerate the action of Na, though they do not themselves produce rhythmical contractions directly: for instance, H and OH. In spite of the antagonistic effect of Ca, a certain amount of it must be present if contractions are to continue for any length of time. Ions produce rhythmical contraction

only because they affect either the physical condition of the colloidal substances (proteid, &c.) in protoplasm, or the rapidity of chemical processes.

Loeb has even gone so far as to consider that the process of fertilisation is mainly ionic action. He denies that the nuclein in the head of the spermatozoon is essential, but asserts that all the spermatozoon does is to act as the stimulus in the due adjustment of the proportions of the surrounding ions. He supports this view by numerous experiments on ova, in which, without the presence of spermatozoa, he has produced larvæ (generally imperfect ones, it is true) by merely altering the saline constituents of the fluid that bathes them. Whether such a notion will stand the test of further verification must be left to the future. So also must the equally important idea that the basis of a nervous impulse is electrolytic action.

Gramme-molecular Solutions.—From the point of view of osmotic pressure a convenient unit is the gramme-molecule. A gramme-molecule of any substance is the quantity in grammes of that substance equal to its molecular weight. A gramme-molecular solution is one which contains a gramme-molecule of the substance per litre. Thus a gramme-molecular solution of sodium chloride is one which contains 58·5 grammes of sodium chloride (Na = 23·05 ; Cl = 35·45) in a litre. A gramme-molecular solution of grape sugar ($C_6H_{12}O_6$) is one which contains 180 grammes of grape sugar in a litre. A gramme-molecule of hydrogen (H_2) is 2 grammes by weight of hydrogen, and if this were compressed to the volume of a litre it would be comparable to a gramme-molecular solution. It therefore follows that a litre containing 2 grammes of hydrogen contains the same number of molecules of hydrogen in it, as a litre of a solution containing 58·5 grammes of sodium chloride, or one containing 180 grammes of grape sugar has in it of salt or sugar molecules respectively. To put it another way, the heavier the weight of a molecule of any substance the more of that substance must be dissolved in the litre to obtain its gramme-molecular solution. Or still another way : if solutions of various substances are made all of the same strength per cent., the solutions of the materials of small molecular weight will contain more molecules of those materials than the solutions of the materials which have heavy molecules. We shall see that the calculation of osmotic pressure depends on these facts.

Diffusion, Dialysis, Osmosis.—If two gases are brought together within a closed space, a homogeneous mixture of the two is soon obtained. This is due to the movements of the gaseous molecules within the confining space, and the process is called *diffusion*. In a similar way diffusion will effect in time a homogeneous mixture of two liquids or solutions. If water is carefully poured on to the surface of a solution of salt, the salt or its ions will soon be equally distributed throughout the whole. If a solution of albumin or any other *colloidal* substance is used instead of salt in the experiment, diffusion will be found to occur much more slowly. If, instead of pouring the water on to the surface of a solution of salt or sugar, the two are separated by a membrane made of such a material as parchment paper, a similar diffusion will occur, though more slowly than in cases where the membrane is absent. In time, the water on each side of the membrane will contain the same quantity of sugar or salt. Substances which pass through such membranes

are called crystalloids. Substances which have such heavy molecules (starch, proteid, &c.) that they will not pass through such membranes are called *colloids*. Diffusion of substances in solution in which we have to deal with an intervening membrane is usually called **dialysis**. The process of **filtration** (*i.e.* the passage of materials through the pores of a membrane under the influence of mechanical pressure) may be excluded in such experiments by placing the membrane (M) vertically as shown in the diagram (fig. 83), and the two fluids A and B on each side of it. Diffusion through a membrane is not limited to the molecules of water, but it may occur also in the molecules of certain substances dissolved in the water. But very few or no membranes are equally permeable to water and to molecules of the substances dissolved in the water. If in the accompanying diagram the compartment A is filled with pure water, and B with a sodium chloride solution, the liquids in the two compartments will ultimately be found to be equal in bulk as they were at the start, and each will be a solution of salt of half the original strength of that in the compartment B. But at first the volume of the liquid in compartment B increases, because more water molecules pass into it from A than salt molecules pass from B into A. The term **osmosis** is generally limited to the stream of water molecules passing through a membrane, while the term **dialysis** is applied to the passage of the molecules in solution in the water. The osmotic stream of water is especially important, and in connection with this it is necessary to explain the term **osmotic pressure**. At first, then, osmosis (the diffusion of water) is more rapid than the dialysis (the diffusion of the salt molecules or ions). The older explanation of this was that salt attracted the water, but we now express the fact differently by saying that the salt in solutions exerts a certain osmotic pressure : the result of the osmotic pressure is that more water flows from the water side to the side of the solution than in the contrary direction. The osmotic pressure varies with the amount of substance in solution, and is also altered by variations of temperature occurring more rapidly at high than at low temperatures.

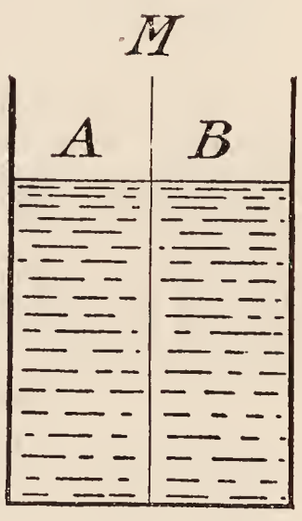


FIG. 83.

If we imagine two masses of water separated by a permeable membrane, as many water molecules will pass through from one side as from the other, and so the volumes of the two masses of water will remain unchanged. If now we imagine the membrane M is not permeable except to water, and the compartment A contains water, and the compartment B contains a solution of salt or sugar; under these circumstances water will pass through into B, and the volume of B will increase in proportion to the osmotic pressure of the sugar or salt in solution in B, but no molecules of sugar or salt can get through into A from B, so the volume of fluid in A will continue to decrease, until at last a limit is reached. The determination of this limit, as measured by the height of a column of fluid or mercury which it will support, will give us a measurement of the osmotic pressure. Membranes of this nature are called **semi-permeable**. One of the best kinds of semi-permeable membrane is

ferrocyanide of copper. This may be made by taking a cell of porous earthenware and washing it out first with copper sulphate and then with potassium ferrocyanide. An insoluble precipitate of copper ferrocyanide is thus deposited in the pores of the earthenware. If such a cell is filled with a 1-per-cent. solution of sodium chloride, water diffuses in till the pressure registered by a manometer connected to it registers the enormous height of 5,000 millimetres of mercury. Theoretically it is possible to measure osmotic pressure by a manometer in this way, but practically it is seldom done, and some of the indirect methods of measurement described later are used instead. The reason for this is that it has been found difficult to construct a membrane which is absolutely semi-permeable.

Many explanations of the nature of osmotic pressure have been brought forward, but none is perfectly satisfactory. The following simple explanation is perhaps the best, and may be rendered most intelligible by an illustration. Suppose we have a solution of sugar separated by a semi-permeable membrane from water; that is, the membrane is permeable to water molecules, but not to sugar molecules. The streams of water from the two sides will then be unequal; on one side we have water molecules striking against the membrane in what we may call normal numbers, while on the other side both water molecules and sugar molecules are striking against it. On this side, therefore, the sugar molecules take up a certain amount of room, and do not allow the water molecules to get to the membrane; the membrane is, as it were, screened against the water by the sugar, therefore fewer water molecules will get through from the screened to the unscreened side than *vice versa*. This comes to the same thing as saying that the osmotic stream of water is greater from the unscreened water side to the screened sugar side than it is in the reverse direction. The more sugar molecules that are present, the greater will be their screening action, and thus we see that the osmotic pressure is proportional to the number of sugar molecules in the solution: that is, to the concentration of the solution.

Osmotic pressure is, in fact, equal to that which the dissolved substance would exert if it occupied the same space in the form of a gas (Van't Hoff's hypothesis). The nature of the substance makes no difference; it is only the number of molecules which causes osmotic pressure to vary. The osmotic pressure, however, of substances like sodium chloride, which are electrolytes, is greater than what one would expect from the number of molecules present, This is because the molecules in solution are split into their constituent ions, and an ion plays the same part as a molecule, in questions of osmotic pressure. In dilute solutions of sodium chloride ionisation is more complete, and as the total number of ions is then nearly double the number of original molecules, the osmotic pressure is nearly double what would have been calculated from the number of molecules.

The analogy between osmotic pressure and the partial pressure of gases is very complete, as may be seen from the following statements:—

1. At a constant temperature osmotic pressure is proportional to the concentration of the solution (Boyle-Mariotte's law for gases).

2. With constant concentration, the osmotic pressure rises with and is proportional to the temperature (Gay-Lussac's law for gases).

3. The osmotic pressure of a solution of different substances is equal to the sum of the pressures which the individual substances would exert if they were alone in the solution (Henry-Dalton law for partial pressure of gases).

4. The osmotic pressure is independent of the nature of the substance in solution, and depends only on the number of molecules or ions in solution (Avogadro's law for gases).

Calculation of Osmotic Pressure.—We may best illustrate this by an example, and to simplify matters we will take an example in the case of a non-electrolyte like sugar. We shall then not have to take into account any electrolytic dissociation of the molecules into ions. We will suppose we want to calculate the osmotic pressure of a 1-per-cent. solution of cane sugar.

One gramme of hydrogen at atmospheric pressure and 0° C. occupies a volume of 11·19 litres; two grammes of hydrogen will therefore occupy a volume of 22·38 litres. A gramme-molecule of hydrogen—that is, 2 grammes of hydrogen—when brought to the volume of 1 litre will exert a gas pressure equal to that of 22·38 litres compressed to 1 litre—that is, a pressure of 22·38 atmospheres. A gramme-molecular solution of cane sugar, since it contains the same number of molecules in a litre, must therefore exert an osmotic pressure of 22·38 atmospheres also. A gramme-molecular solution of cane sugar ($C_{12}H_{22}O_{11}$) contains 342 grammes of cane sugar in a litre. A 1-per-cent. solution of cane sugar contains only 10 grammes of cane sugar in a litre; hence the osmotic pressure of a 1-per-cent. solution of cane sugar is $\frac{10}{342} \times 22{\cdot}38$ atmospheres, or 0·65 of an atmosphere, which in terms of a column of mercury = 760 × 0·65 = 494 mm.

It would not be possible to make such a calculation in the case of an electrolyte, because we should not know how many molecules had been ionised. In the liquids of the body, both electrolytes and non-electrolytes are present, and so a calculation is here also impossible.

We have seen the difficulty of directly measuring osmotic pressure by a manometer; we now see that mere arithmetic often fails us; and so we come to the question to which we have been leading up, viz. how osmotic pressure is actually determined.

Determination of Osmotic Pressure by means of the Freezing-point.—This is the method which is almost universally employed. A very simple apparatus (Beckmann's differential thermometer) is all that is necessary. The principle on which the method depends is the following:—The freezing-point of any substance in solution in water is lower than that of water; the lowering of the freezing-point is proportional to the molecular concentration of the dissolved substance, and that, as we have seen, is proportional to the osmotic pressure.

When a gramme-molecule of any substance is dissolved in a litre of water, the freezing-point is lowered by 1·87° C., and the osmotic pressure is, as we have seen, equal to 22·38 atmospheres: that is, 22·38 × 760 = 17,008 mm. of mercury.

We can, therefore, calculate the osmotic pressure of any solution if we know the lowering of its freezing-point in degrees Centigrade; the lowering of the freezing-point is usually expressed by the Greek letter Δ.

$$\text{Osmotic pressure} = \frac{\Delta}{1{\cdot}87} \times 17{,}008.$$

For example, a 1-per-cent. solution of sugar would freeze at $-0{\cdot}052°$ C.; its osmotic pressure is therefore $\frac{{\cdot}052 \times 17{,}008}{1{\cdot}87} = 473$ mm., a number approximately equal to that we obtained by calculation.

Mammalian blood serum gives $\Delta = 0{\cdot}56°$ C. A 0·9-per-cent. solution of sodium chloride has the same Δ; hence serum and a 0·9-per-cent. solution of common salt have the same osmotic pressure, or are *isotonic*. The osmotic pressure of blood serum is $\frac{{\cdot}56 \times 17{,}008}{1{\cdot}87} = 4{,}987$ mm. of mercury, or a pressure of nearly 7 atmospheres.

The osmotic pressure of solutions may also be compared by observing their effect on red corpuscles, or on vegetable cells such as those in *Tradescantia*. If the solution is *hypertonic*, i.e. has a greater osmotic pressure than the cell contents, the protoplasm shrinks and loses water, or, if red corpuscles are used, they become crenated. If the solution is *hypotonic*, e.g. has a smaller osmotic pressure than the material within the cell-wall, no shrinking of the protoplasm in the vegetable cell occurs, and if red corpuscles are used they swell and liberate their pigment. *Isotonic* solutions produce neither of these effects, because they have the same molecular concentration and osmotic pressure as the material within the cell-wall.

Physiological Applications.—It will at once be seen how important all these considerations are from the physiological standpoint. In the body we have aqueous solutions of various substances separated from one another by membranes. Thus we have the endothelial walls of the capillaries separating the blood from the lymph; we have the epithelial walls of the kidney tubules separating the blood and lymph from the urine; we have similar epithelium in all secreting glands; and we have the wall of the alimentary canal separating the digested food from the blood-vessels and lacteals. In such important problems, then, as lymph-formation, the formation of urine and other excretions and secretions, and absorption of food, we have to take into account the laws which regulate the movements both of water and of substances which are held in solution by the water. In the body osmosis is not the only force at work, but we have also to consider filtration: that is, the forcible passage of materials through membranes, due to differences of mechanical pressure. Further complicating these two processes we have to take into account another force, namely, the secretory or selective activity of the living cells of which the membranes in question are composed. This is sometimes called by the name *vital action*, which is an unsatisfactory and unscientific expression. The laws which regulate filtration, imbibition, and osmosis are fairly well known and can be experimentally verified. But we have undoubtedly some other force, or some other manifestation of force, in the case of living membranes. It probably is some physical or chemical property of living matter which has not yet been brought into line with the known chemical and physical forces which operate in the inorganic world. We cannot deny its existence, for it sometimes operates so as to neutralise the known forces of osmosis and filtration.

The more one studies the question of lymph-formation, the more convinced one becomes that mere osmosis and filtration will not explain it entirely.

The basis of the action is no doubt physical, but the living cells do not behave like the dead membranes of a dialyser; they have a selective action, picking out some substances and passing them through to the lymph, while they reject others.

The question of gaseous interchanges in the lungs has been another battlefield of a similar kind. Some maintain that all can be explained by the laws of diffusion of gases; others assert that the action is wholly vital. Probably those are most correct who admit a certain amount of truth in both views; the main facts are explicable on a physical basis, but there are also some puzzling data which show that the pulmonary epithelium is able to exercise some other force as well which interferes to some extent with the known physical process. Take again the case of absorption. The object of digestion is to render the food soluble and diffusible; it can hardly be supposed that this is useless; the readily diffusible substances will pass more easily through into the blood and lymph: but still, as Waymouth Reid has shown, if the living epithelium of the intestine is removed, absorption comes very nearly to a standstill, although from the purely physical standpoint removal of the thick columnar epithelium would increase the facilities for osmosis and filtration.

The osmotic pressure exerted by crystalloids is very considerable, but their ready diffusibility limits their influence on the flow of water in the body. Thus, if a strong solution of salt is injected into the blood, the first effect will be the setting up of an osmotic stream from the tissues to the blood. The salt, however, would soon diffuse out into the tissues, and would now exert osmotic pressure in the opposite direction. Moreover, both effects will be but temporary, because excess of salt is soon got rid of by the excretions.

Osmotic Pressure of Proteids.—It has been generally assumed that proteids, the most abundant and important constituents of the blood, exert little or no osmotic pressure. Starling, however, has claimed that they have a small osmotic pressure; if this is so, it is of importance, for proteids, unlike salt, do not diffuse readily, and their effect therefore remains as an almost permanent factor in the blood. Starling gives the osmotic pressure of the proteids of the blood-plasma as equal to 30 mm. of mercury. By others this is attributed to the inorganic salts with which proteids are always closely associated. Moore, for instance, finds that the purer a proteid is, the less is its osmotic pressure; the same is true for other colloidal substances. It really does not matter much, if the osmotic force exists, whether it is due to the proteid itself, or to the saline constituents which are almost an integral part of a proteid. It is merely interesting from the theoretical point of view. We should from the theoretical standpoint find it difficult to imagine that a pure proteid can exert more than a minimal osmotic pressure. It is made up of such huge molecules that, even when the proteids are present to the extent of 7 or 8 per cent., as they are in blood-plasma, there are comparatively few proteid molecules in solution. Still, by means of this weak but constant pressure it is possible to explain the fact that an isotonic or even a hypertonic solution of a diffusible crystalloid may be completely absorbed from the peritoneal cavity into the blood.

The functional activity of the tissue elements is accompanied by the

breaking down of their proteid constituents into such simple materials as urea (and its precursors), sulphates, and phosphates. These materials pass into the lymph, and increase its molecular concentration and its osmotic pressure; thus water is attracted (to use the older way of putting it) from the blood to the lymph, and so the volume of the lymph rises and its flow increases. On the other hand, as these substances accumulate in the lymph they will in time attain there a greater concentration than in the blood, and so they will diffuse towards the blood, by which they are carried to the organs of excretion.

But, again, we have a difficulty with the proteids; they are most important for the nutrition of the tissues, but they are practically indiffusible. We must provisionally assume that their presence in the lymph is due to filtration from the blood. The plasma in the capillaries is under a somewhat higher pressure than the lymph in the tissues, and this tends to squeeze the constituents of the blood, including the proteids, through the capillary walls. I have, however, already indicated that the question of lymph-formation is one of the many physiological problems which await solution by the physiologists of the future.

INDEX

In cases where several references are made to any subject, the figure in heavy type indicates where the principal matter in relation to the subject is to be found.

A

Abrin, 119
Absorption, 86; of carbohydrates, 87; of fats, 89; of proteids, 88
Absorption bands, 105
Absorption spectra of hæmoglobin and its derivatives, 107, 169; of myohæmatin, 178; of urinary pigments, 194
Accessories of food, 52
Acetic acid, 14, 24
Acetonæmia, 79
Acetone, 148
Acetyl, 25
Achroö-dextrin, **21**, 59, 159
Acid-albumin, 29, **35**, 38, 66, 72
Acid hæmatin, 168
Acid hæmatoporphyrin, 195; absorption bands of, 194
Acid, lactic, 47; test for, 165
Acid sodium phosphate, 29
Acid tide, 124
Acids, vegetable, 4, 124, 133
Acrolein, 24
Acute yellow atrophy of liver, 128, 129
Adamkiewicz's reaction, 27, 32
Adenine, 38, 138, **139**
Adipose tissue, 2
Aërobic micro-organisms, 57
Aërotonometer, 114
Agglutinating action, 119; agglutinins, 119
Air, expired and inspired, 110; pumps, mercurial, 212, 213
Alanine, 78
Albumin, acid, 29, 35; alkali, 28, 35
Albumin, action of acids and alkalis on, 28
Albuminates, 35
Albumin in urine, estimation of, 146; tests for, 146
Albumins, 30, **34**, 67, 163
Albuminoids, 4, **39**, 72
Albuminometer of Esbach, 146
Albumose, 30, 33, 66, **162**, 163
Albumoses and peptone, separation of, 27, 162; tests for, 27, 163
Alcapton, 149
Alcaptonuria, 149
Alcohol, action of, on proteids, 162
Alcoholic fermentation, 19; of milk, 19, 47
Alcohols, 4, 14, 24
Aldehyde, 14, 15, 24
Aldoses, 15
Aleurone grains, 32
Alkali-albumin, 28, **35**, 36, 72, 166
Alkaline hæmatin, 168
Alkaline hæmatoporphyrin, absorption bands of, 169
Alkaline tide, 124
Alkaloids, 52
Alloxan, 138
Alloxuric bases, 38, 138
Allyl alcohol, 24
Alvergniat's pump, 213
Amidulin, 20
Amines, 4
Amino-acetic acid, 77, 129, 141
Amino-acids, 73, 129, 130
Amino-caproic acid, 78, 129
Amino-ethyl-sulphonic acid, 82
Amino-hydroxybutyric acid, 80
Amino-iso-butyl-acetic acid, 77, 78
Amino-oxy-purine, 138
Amino-propionic acid, 78
Amino-purine, 138
Amino-pyrotartaric acid, 72
Amino-succinic acid, 72
Amino-valerianic acid, 40
Ammonia, 4, 72, 76, 128, 130, **131**, 134
Ammoniacal odour of putrid urine, 126

Ammonium carbonate and carbamate as urea precursors, 130, 131
Ammonium cyanate, 125
Ammonium sulphate, action of, on proteids, 27, 33, 34, 35, 41, 67, 93, **162**
Amœboid movements, 2
Amylolytic ferments, 56
Amylopsin, 19, 56, 69, 71, **73**
Amyloses, 16, 56
Anaërobic micro-organisms, 57
Analyses of gases, 214
Animal starch, 21
Anions, 217
Anisotropous bodies, 203
Anti- bodies, 68
Antipeptone, 72, 73
Antiseptics, 54
Antitoxic material, 116
Apparatus necessary for practical work, 4, 5, 6
Aqueous vapour, tension of, 8
Arginine, **40**, 72, 73, 129, 130
Aromatic compounds, 4, 18
Arterial blood, gases of, 110
Artificial gastric juice, 54
Aspartic acid, 72, 73, 129
Asphyxia, 111
Assimilation, 2
Atmosphere, 109, 110
Atomic weights, 8
Attraction sphere, 2
Avogadro's law, 110, 221

B

Bacilli, 56
Bacteria, 55
Bacterial action, 76
Bacterio-lysin, 117, 118
Barfoed's reagent, 158
Bath, warm, 28
Baumann and Wolkow on alcapton, 149
Beaumont, Dr., 61
Beef, 42, 48; beef-tea, 51
Beetroot, 18
Benger's liquor pancreaticus, 69; liquor pepticus, 53
Benzene, derivatives of, 4
Benzoic acid, 140
Bernard, Claude, on glycogen, 87
Bile, 69, **80**; amount secreted, 81; circulation, 81; characters of, 81; constituents, 81, 82, 85; mucin of, 82; pigments, 70, 81, 83; salts of, 81, 82; uses of, 84; action of, 91; in urine, 149
Biliary fistula, 80
Bilirubin, 81, **83**, 123, 192; of meconium, 86
Biliverdin, 81, **83**
Biuret, **32**, 67, 121, 163; reaction, 27, 32, 163
Blood, **92–120**; coagulation of, 93, 94, 95; corpuscles, 98; detection of, 116, 152; gases of, 110; specific gravity of, 202; tablets, 95; pigment, 102, 168; biological test for, 116; plasma and serum of, 92, 96; in urine, 149
Blood clot, 96
Bohr on absorption of oxygen, 114; on tension of carbonic dioxide, 114
Bone, composition of, 11, 39; marrow, 23
Bowman's capsule, 122
Bran, 49
Bread, 42, 50
Bright's disease, 148
Brown and Morris on digestion of starch, 59
Buffy coat, 94
Bunge on hæmatogens, 37; on milk, 44
Bush tea, 52
Butter, 4, 41
Butyric acid, 19
Butyrin, 47

C

Caffeine, 52, 138
Calcium, 3, 8; phosphate, 4
Calcium oxalate in urine, 135, 145
Calcium salts, importance of, in coagulation of blood, 95, 96, 174; of milk, 46, 47, 161
Cane sugar, 13, 16, **18**, 75, 157; in urine, 18; tests for, 13, 18
Caproic acid, 24
Caproin, 47
Caprylin, 47
Carbamide, 125. See also Urea
Carbohydrates, 4, **13–21**, 156, 158; absorption of, 87; classification of, 16; definition of, 14; tests for, 13–21, 151
Carbolic acid poisoning and urine, 195
Carbon, 3, 8; tests for, 9
Carbonates in blood, 112, 113; in urine, 133
Carbonic acid in air, 109, 110; in blood, 112
Carbonic oxide hæmoglobin, 102, 108, 115, 168
Cardiac glands, 61, 62, 63
Carmine-stained fibrin, 164
Cartilage hyaline, 39
Casein, 41, **45**, 46, 129, 161

Caseinogen, 41, **45**, 46, 161
Caseinogenate of calcium, of sodium, of potassium, 46, 47
Cell, definition of, 2 ; diagram of, 37
Cells, differentiation of, 1
Cellulose, 16, 21, 56, 86
Centigrade scale, 8
Central cells of cardiac glands, 61
Centrifugal machine, 174
Centrosome, 2
Cerebrin, 179
Cerebrosides, 179
Cerebrospinal fluid, 182 ; functions of, 182
Chemical physiology, 1
Chemical sediments of urine, 145
Chemical structure of protoplasm, 2
Chitin, 39
Chlorides of urine, 132 ; estimation of, 185, 186 ; tests for, 121
Chlorine, 3, 8
Chlorophyll, 86
Cholalic acid, 85
Cholesterin, 3, 4, 26, 70, 81, **84**, 85, 86, 99, 179 ; tests for, 84
Choletelin, 83
Choline, 26, 77, **180** ; tests for, 180
Chondrin, 39
Chorda tympani, 58
Chromatin, 37
Chromogens, 107 ; of urine, 195
Chyle, 89
Chyme, 80, 85
Ciliary movement, 2
Circular polarisation and chemical constitution, 210
Circulation of bile, 81, 85
Cirrhosis of liver, 128
Citric acid, 46, 124
Clark's essence of rennet, 51
Clotting of blood, 93, 94, 95, 174
Clupeine, 40
Coagulated proteids, 35
Coagulation, 33
Coagulation of blood, **93–96**, 174 ; of hydrocele fluid, 175 ; of milk, **45**, 46, 161 ; of muscle, 176 ; of proteids, 27, 30, 172, 173
Coagulative ferments, 33, 56
Cobalt sulphate, actions on proteids, 163
Cocaine, 52
Cocoa, 52, 138
Coffee, 52, 138
Coffin-lid crystals, 134
Cola nut, 52
Collagen, 29, 39, 72
Collimator, 104, 171
Colloid carbohydrates, salting out of, 21
Colloids, 30, 73, 219
Colostrum, 45
Colostrum corpuscles, 44
Commercial peptone, 95, 162
Complement, 119
Compound proteids, 36
Compounds found in the body, 4
Compounds of carbon, 4
Condiments, 52
Cooking of food, 50
Copper, 3, 8
Cream, 41, 47
Creatine, 4, 128, 129, 130 ; crystals, 142 ; in muscle, 178 ; as a urea precursor, 128
Creatinine, 122, 128, **141** ; crystals of, 142 ; detection of, 122 ; estimation of, 191 ; preparation of, 141
Crypts of Lieberkühn, 75
Crystallin, 34
Crystalline lens, 34
Crystallisable proteids, **31**, 100, 167, 171
Crystallisation of egg albumin, 160
Crystalloids, 31
Crystals from blood, 100, 102, 167
Curative inoculation, 116
Curd, 41, 45, 161
Curds and whey, 46
Cyclopterine, 40
Cyanamide, 129, 141
Cyanogen, 163
Cystin, 143 ; crystals of, 144
Cystinuria, 144

D

Decalcified blood, 92, 174
Decalcified milk, 41, 46, 161
Decomposition, products of fats, 25
Density of water, 8
Dentine, composition of, 39
Deposits in urine, 142
Desiccator, 178
Deutero-albumose, 66, 162, 166
Deutero-proteose, 66, 67
Dextrin, 13, 14, 16, **21**, 159 ; tests for, 14, 159
Dextro-rotatory, 17
Dextrose, 13, 14, 16, **17**, 19, 157 ; crystals, 17 ; in blood, 17 ; in urine, 146, 147
Diabetes mellitus, 17, 78, 148
Diabetic urine, 146, 148
Diacetin, 25
Dialyser, 31
Dialysis, **31**, 172, 216, 218, 219 ; of albumoses, 162 ; of serum, 172
Di-amino-caproic acid, 129
Di-amino-valerianic acid, 129

Diastase, 56
Diastatic ferments, 19, 159
Diet, 42
Diet tables, 43
Dietary, 43
Diffusion, 31, 113, 216, 218
Digestion, 164; gastric, 53, 61–68; intestinal, 75–78; pancreatic, 69, 70–75; salivary, 53, 58–61
Dimethylxanthine, 138
Dioxy-purine, 138
Diphtheria-toxin, 118
Direct vision spectroscope, 105, 106
Disaccharides, 16, 18
Dissociation, 217
Disuse atrophy, 181
Dough, 49
Drechsel on urea formation, 129
Dripping, 51
Dropsical effusions, 37
Dulcite, 15
Dumb-bell crystals, 149
Dupré's urea apparatus, 121
Dysalbumose, 162
Dyer's modification of Kjeldahl's method of estimating nitrogen, 215

E

Eck's fistula, 128
Egg-albumin, 27, 29, 30, 34; crystallisation of, 32, 160; in urine, 38
Egg-globulin, 27, 29, 30
Eggs, 48
Egg-white, **27**, 28, 53, 69
Egg yolk, 48
Ehrlich's experiments with methylene blue, 112
Elastic fibres, 39
Elastin, 39, 72
Elastoses, 66
Electrolytes, 217
Elements found in the body, 3; symbols and atomic weights of, 8
Emulsification, 23, 25, **89**, 90
Emulsion, 25, 73
Enamel, composition of, 39
English system of weights and measures, 6, 7
Entero-kinase, 76, 166
Envelope crystals, 135, 144
Enzymes, 55, 57
Epidermis, 2
Epithelium of intestine, 88, 89
Epsom salts, 132
Erepsin, 76, 89
Erlenmeyer flask, 191
Erythro-dextrin, **21**, 59, 159
Esbach's albuminometer, 146; reagent, 5; tube, 146
Estimation of chlorides, 185; of creatinine, 191; of dextrose, 147; of lactose, 159; of maltose, 159; of nitrogen, 215; of phosphates, 187; of sulphates, 188; of urea, 121, 183; of uric acid, 190
Ethereal sulphates in urine, 133; estimation of, 188
Ethyl alcohol, 24
Ethyl-diacetic acid, 148
Eu-globulin, 173
Expired air, 110
External respiration, 109
Extirpation of pancreas, 78
Extractives of blood, 98; of muscle, 51, 177, 178
Extraordinary ray, 202

F

Fæces, 86
Fahrenheit scale, 8
Fats, 4, **22–26**, 41
Fat, absorption of, 89–91; constitution of, 24; decomposition products of, 25; melting point of, 22, 23; tests for, 22
Fatty acids, 22
Fehling's test, 13, 20, **146**, 148, 159
Ferment coagulation, 33, 56
Ferment, invert, 75
Ferment of ferments, 76
Fermentation, 54
Fermentation test, 13, 149
Ferments, 4, **54–57**; of gastric juice, 63; of pancreatic juice, 71; of saliva, 59; of succus entericus, 75; organised, 55; unorganised, 59
Ferments, classification of, 55; diastatic, 159
Fibres, elastic, 39
Fibrin, **94**, 96, 97, 164
Fibrin ferment, 56, 92, 95, **98**, 175; preparation of, 98
Fibrin filaments, 94, 97
Fibrinogen, 34, 95, 96, **97**
Fibrino-globulin, 96, 97
Filtration, 217
Fistula, gastric, 61
Fleischl's hæmometer, 200; spectropolarimeter, 209
Flour, 42, 49
Fluorescent screens, 171
Fluorine, 8
Foods, 41–52; cooking of, 50
Formic acid, 24
Fractional heat coagulation of muscle proteids, 176; of serum proteids, 172, 173
Fraunhofer's lines, 93, 103, 169

Fredericq on oxygen tension, 111; on tension of carbon dioxide, 113
Free hydrochloric acid in gastric juice, tests for, 165
Fürth, v., on muscle plasma, 176

G

Galactose, 15, 16, **18**, 157
Gallstones, 84
Gamgee on secretion of hydrochloric acid, 64; on photographic spectra, 170, 171
Garrod on urochrome, 123
Garrod and Hopkins on urobilin and stercobilin, 193
Garrod's method of separating pigments from urine, 192
Gas analysis, 214
Gaseous interchange in lungs, 109, 113
Gases of blood, 110
Gastric digestion, 53
Gastric fistula, 61
Gastric juice, 53; action of, 66; composition of, 64; glands, 62, 63; properties of, 61; secretion of, 61
Gay-Lussac's law, 220
Gelatin, 4, 29, **39**, 164; tests for, 29
Gelatinisation, 29
Gelatoses, 66
Germ theory of diseases, 54
Globin, 101
Globulin, 30, 32, **34**, 67, 163, 179
Globulose, 66
Glossopharyngeal nerve, 58
Gluco-proteid, 36
Glucosamine, 36
Glucose, 16, **17**, 79, 147, 148
Glutaminic acid, 129
Gluten, 42, 49
Glyceric ethers, 24
Glycerides, 24
Glycerin, 15, 22, 25
Glycerol, 25
Glyceryl, 25
Glycine, 85, 129, 130, 140
Glycocholate of soda, 81
Glycocholic acid, 70, 82
Glycocine, *see* Glycine
Glycogen, 14, 16, **21**, 87; microscopical detection of, 157; preparation of, 156; tests for, 14, 21, 157
Glycol, 15
Glycolysis, 79
Glycuronic acid, 79, 149, 158
Glyoxylic acid, 5, 27
Gmelin's test, 70, 83
Gnezda on biuret reaction, 163
Gowers, Sir William, his hæmacytometer, 197; hæmoglobinometer, 199
Graham, Thomas, on colloids and crystalloids, 30, 31
Gram-molecular solutions, 218
Granulose, 20
Grape sugar, *see* Dextrose
Ground substance, 29, 36
Grützner's method of comparing digestive power of solutions of pepsin, 164
Guanine, 38, 138, 139
Guarana, 52
Gums, 21
Gunsberg's reagent, 165
Gürber on serum albumin crystals, 32

H

Hæmacytometer of Sir William Gowers, 197; of Oliver, 198
Hæmatin, 86, 101, 192; acid, 168; alkaline, 168; iron free, 171; of food, 123
Hæmatogen, 37
Hæmatoidin, 81; crystals, 80
Hæmatoporphyrin, 101, 102, 193; absorption bands of, 169, 194
Hæmatoscope of Hermann, 105
Hæmin, 93, 101; crystals, 102, 116; preparation of, 93
Hæmochromogen, 170, 177
Hæmoglobin, 36, 86, 93, 99, **100**, 123, 168; composition of, 101; derivatives of, 168, 169, 170
Hæmoglobinometer of Sir William Gowers, 199; of Haldane, 200; of Oliver, 200
Hæmoglobinuria, 117; paroxysmal, 150
Hæmolysins, 117
Hæmometer of von Fleischl, 200
Haldane on methæmoglobin, 108; on absorption of oxygen, 114
Haldane's hæmoglobinometer, 200
Halogens, action on proteid, 36
Hammarsten on blood coagulation, 95; method of precipitating serum globulin, 172
Hammerschlag's method of estimating the specific gravity of blood, 202
Haptophor group, 119
Hayem's fluid, 199
Hay's test for bile salts, 70, 149
Heat coagulation of proteids, 33, 172; of serum globulin and serum albumin, 172, 173
Heat rigor, 177
Heidenhain on secretion of gastric juice, 63; on pressure in bile duct, 80
Heller's nitric acid test, 146
Hemipeptone, 72
Henry-Dalton law, 221
Heptoses, 16

Hermann, hæmatoscope of, 105
Hetero-albumose, 66, 162 ; proteose, 72, 148
Hexone bases, **40**, 73, 76, 129
Hexoses, 16
Hill's air-pump, 213
Hill, Croft, on ferments, 57,
Hippuric acid, 140 ; crystals of, 140 ; tests for, 153
Histidine, 40, 73
Histone of Kossel, 101
Hofmeister on crystallisation of egg-albumin, 32
Homogentisinic acid, 149
Hopkins's method of estimating uric acid, 137, 190 ; on crystallisation of egg-albumin, 32, 160
Hoppe-Seyler on hæmatin, 101 ; on proteids, 29
Hot-air oven, 156
Hüfner's method of estimating urea, 121
Hyaline cartilage, 39
Hydrazone, 157
Hydrobilirubin, **83**, 123, 192
Hydrocarbon, 24
Hydrocele fluid, 96, 175
Hydrochinon, 195
Hydrochloric acid, 165 ; of gastric juice, 64 ; tests for, 165 ; test for cane sugar, 13
Hydrochloride of hæmatin, 101
Hydrogen, 3, 8, 9 ; tests for, 9
Hydrolysis, 19, 66
Hydrolytic ferments, 56
Hydrometer, 41, 45, 202
Hydroxybutyric acid, 148
Hydroxyl, 24
Hypertonic solutions, 222
Hypobromite of soda, action of, on urea, 121, **126**, 184
Hypotonic solutions, 222
Hypoxanthine, 38, 98, 138, **139**

I

Immune body, 119
Immunity, 116, 119; side chain, theory of, 119
Indican, 133
Indiffusibility of proteids, 30, 224
Indigo, 133 ; blue, 195 ; red, 195
Indole, 86, 133
Indoxyl, 133, 195
Indoxyl sulphate of potassium, 133, 195
Infection, 54, 55
Inoculation, curative, 116 ; protective, 116
Inorganic compounds, 4 ; salts, 4
Inosite, 14 ; crystals of, 18
Inspired air, 110
Internal respiration, 109
Internal secretion, 78, 79
Interstitial substance, 29, 36
Intestinal juice, 18, 75 ; digestion, 75
Intravascular coagulation, 175
Inversion, 17, **18**, 75
Inversion of cane sugar, 18, 56
Inversive ferments, 17, 56
Invertin, 56, 75
Iodine, 11
Iodine test, 159
Ions, 216
Iron, 8, 11, 100 ; in milk, 44
Iron-free hæmatin, 169, 171
Islets of Langerhans, 70, 79
Iso-cholesterin, 84
Isotonic solutions, 222
Isotropous bodies, 203

J

Jaffé on test for indoxyl, 195
Jaundice, 149
Jelly, Lieberkühn's, 36 ; Whartonian, 29
Johnson, G. S., on creatinine, 141 ; on sugar in urine, 147
Johnson, Sir G., on picric-acid test, 147
Juice, intestinal, 75
Junkets, 51

K

Kations, 216, 217
Kauder's method of precipitating serum globulin, 172
Kephalin, 179
Keratin, 2, 4, 11, **39**, 86
Ketone, 14, 15
Ketoses, 15
Kidney, 122
Kidneys, removal of part of, 78 ; removal of both, 127
Kjeldahl's method of estimating nitrogen, 215
Kossel on protamines, 40 ; on histone, 101
Koumiss, 47
Kühne on peptone, 72 ; on precipitation of pepsin, 65
Külz's method of extracting glycogen, 156
Kutscher on antipeptone, 73

L

Lactalbumin, 41, **45**, 161 ; properties of, 45
Lacteals, 87

Lactic acid, 4, 19, **47**, 51, 65, 86, 140, 165, tests for, 165
Lactic acid fermentation, 19, 75; in milk, 47, in muscle, 51, 176; organisms, 19, 47
Lacto-globulin, 161
Lactometer, 41
Lactose, 14, 16, **19**, 41, 148, 157; in urine, 148; tests for, 19, 141
Laky blood, 100
Lanoline, 84
Lard, 22
Lassaigne's test for nitrogen, 10
Lateritious deposit, 135, 137
Laurent's polarimeter, 208
Lead, 3, 8
Lecithin, 3, 4, 11, 25, 26, 76, 84, 99, 179, **180**
Leech extract, 95
Leucine, 40, 71, 72, 73, 76, **77**, 81, 130, 166; as a urea precursor, 129; in urine, 128; tests for, 166
Leucine crystals, 77; preparation of, 166
Levo-rotatory, 17
Levulose, 15, 16, **17**, 18, 157; in blood, 17; in muscle, 17; in urine, 17; reactions of, 17
Lieberkühn's crypts, 75; jelly, 36
Liebig's extract, 178; method of estimating chlorides, 186; urea, 183
Lime water, 29
Lipochrome, 47, 48; in muscle, 177
Lipolytic ferments, 56
Liquor pancreaticus, 69; pepticus, 53
Lithates, *see* Urates
Lithium, 3, 8
Litre, standard of capacity, 7
Liver, function of, in relation to urea, 128; uric acid, 137
Living test-tube experiment, 96
Loeb on ionic action, 217; on fertilisation, 218
Lungs, 109
Lysatinine, 129
Lysine, **40**, 72, 129

M

McKendrick on cholesterin, 84
MacMunn on stercobilin, 86
McWilliam's test for proteids, 163
Magnesium, 3, 8
Magnesium sulphate, action of, on proteids, 27, 34, 35
Malic acid, 124
Mallow, 18
Malpighian capillaries, 122
Maltase, 57
Malting ferment, 159
Maltose, 16, **19**, 75, 157
Malt upon starch, action of, 159; diastase, 159
Maly on the formation of hydrochloric acid, 63, 64
Manganese, 3, 8
Mannite, 15
Mannose, 15
Marchi reaction, 181
Marsh gas, 77
Measures of capacity, 7; of length, 7
Meat, 9, 42, 48; constituents of, 49
Meconium, 86
Melanin, 195
Mercurial air-pumps, 211
Mercuric nitrate, method of estimating chlorides, 186; urea, 183
Mercury compound of creatinine, 141
Metabolism, 2
Methæmoglobin, 102, **107**, 167, 168, 169; crystals of, 167; in urine, 150
Methane, 56
Methyl-glycine, 141
Methylene blue experiments, 112
Methyl-orange, 215
Metric system, 7
Mett's tubes, 164
Microchemical detection of glycogen, 157
Micrococcus urea, 57, 126, 144
Microspectroscope, 105
Milk, 41, **44**, 161; alcoholic fermentation of, 47; coagulation of, 45, 161; composition of, 45; fats of, 47; proteids of, 45; salts of, 48; souring of, 47
Milk-curdling ferment of pancreas, 71, 74; of stomach, 45, 66
Milk-sugar, 18, **19**, 41, 75; crystals, 19
Millon's reagent, 5; test, 27
Mineral compounds, 3
Mohr's method of estimating chlorides, 185
Moleschott's diet, 43
Monatomic alcohols, 25
Monoacetin, 25
Monochromatic light, 208
Monosaccharides, 16, **17**
Monoxypurine, 138
Moore and Rockwood on fat absorption, 90
Moore on osmotic pressure of proteids, 223
Moore's test for sugar, 13, 17
Mörner and Sjöqvist's method of estimating urea, 184
Mörner on hæmin, 102
Morris and Brown on starch digestion, 59
Mucic acid, 18, 158

Mucin, 2, 29, 36; in bile, 37, 70; in saliva, 29; in urine, 142; tests for, 29
Mucinogen, 58
Mucoids, 36, 37
Mucous glands, structure of, 60
Mucous membrane of frog's intestine, 89
Mucous salivary glands, 60
Munk on fat absorption, 90
Murexide test, 135, 136
Muscle, 176; clot, 176; pigments of, 177; plasma of, 176; extractives of, 178
Muscle fibrin, 176
Muscles, pale and red, 177; involuntary, 177
Muscle sugar, 18
Muscular movement, 2; exercise and urea, 127; exercise and carbonic acid, 101
Mutton, 43
Myogen, 176; myogen fibrin, 176
Myo-hæmatin, 177; absorption spectrum of, 178
Myosin, 4, 176, 177; fibrin, 176
Myosinogen, 176, 177
Myxœdema, 78

N

Nencki and Sieber on pigments, 192
Nencki's experiments on urea, 130
Nerve degeneration, chemistry of, 180
Nervous tissues, 179
Neurokeratin, 39, 179
Neutral fats, 22
Neutral salts, action of, on carbohydrates, 21; on proteids, 27, 30, 35, 41, 92, 97, **162**, 172, 173
Nickel sulphate, action of, on proteids, 163
Nissl's bodies, 179
Nicol's prism, 203
Nitrate of urea, 126, 135
Nitric oxide hæmoglobin, 102, **108**
Nitrogen, 9; estimation of, 215; tests for, 9, 10
Nitrogenous food, 48; glucosides, 179
Nitrous acid, action of, on urea, 126, 135
Non-electrolytes, 217
Nucleic acid, 37; decomposition of, 38
Nuclein, 3, 11, 37, **38**, 138, 139, 140; bases from, 138
Nucleo-proteid, 3, 11, **37**, 95, 179; in bile, 70, 82; decomposition of, 38; in muscle, 177; tests for, 153
Nucleus, functions of, 2

O

Oleic acid, 25
Olein, 23, 25
Olfactory nerve, 58
Oliver's hæmacytometer, 198; hæmoglobinometer, 200
Orcin reaction, 158
Ordinary ray, 202
Organic compounds, 4
Organised ferments, 55
Osazones, 157
Osborne, W. A., on coagulation of milk, 47
Osmic acid test, 22, **24**, 41
Osmosis, 31, 216, 218, 219
Osmotic pressure, 219; calculation of, 221; of proteids, 223; determination of, 221; physiological application of, 222
Ossein, 39
Osteomalacia, 148
Ovarian cyst fluid, 37
Ovo-mucoid, 37, 48
Oxalate of calcium, in milk, 41, 46, 47; plasma, 174; in urine, 135, 144
Oxalate of urea, 126, 135
Oxidases, 57
Oxygen, 3, 8; in blood, 111
Oxyhæmoglobin, 102, 107, 168; crystals, 100, 167; in muscle, 177
Oxyhæmoglobin, pure, preparation of, 171
Oxyntic cells of cardiac glands, 61
Oxyphenyl-amido-propionic acid, 78

P

Pale muscle, 177
Palmitic acid, 25.
Palmitin, 23, **25**, 47
Pancreas, extirpation of, 75; grafting the, 75; structure of, 70, 71
Pancreatic digestion, 69, 166
Pancreatic juice, 71; action of, 71; composition of, 71; secretion of, 74, 165
Panum's method of precipitating serum globulin, 172
Paraglobulin, *see* Serum globulin
Para-mucin, 37
Paramyosinogen, 34, 176, 177
Parapeptone, 66
Parietal cells of cardiac glands, 61, 62, 63
Parotid gland, 58
Paroxysmal hæmoglobinuria, 150
Partial pressure of gases, 110–114
Pasteur on circular polarisation, 210
Pathological urine, 146; pigments of, 195; pigments, 195

Pavy on composition of proteid, 36 ; on glycogenic function of liver, 187 ; on estimation of sugar, 147
Pavy's solution, 147
Pawlow on secretory nerve-fibres to the gastric glands, 65 ; to the pancreas, 74
Pentoses, 16, 158
Pepsin, 52, 56, **66**, 72, 73 ; solutions, activity of, 164 ; hydrochloric acid, 65, 73
Pepsinogen, 63
Peptic digestion, 53, 164
Peptone, 27, 30, **66**, 162, 163 ; precipitation of, 67, 162 ; in urine, 148 ; tests for, 27, 163
Peptonuria, 148
Perfect foods, 42
Pericardial fluids, 96
Pettenkofer's test for bile salts, 70, 82
Pflüger's mercurial air-pump, 211, 212
Phenol, 86, 133
Phenyl sulphate of potassium, 133
Phenyl-hydrazine test for sugars, 157
Phloridzin, 79
Phloroglucin reaction, 158
Phosphates of urine, 133, 134, 135 ; estimation of, 185 ; tests for, 121, 135
Phosphates, stellar, 144
Phosphates, triple, 144
Phospho-molybdic acid, 162
Phosphorised fat, 179
Phosphorus, 3, 8, 11 ; tests for, 10
Phospho-tungstic acid, 162
Photographic spectra, 170, 171
Physiological chemistry, 1
Physiological proximate principles, detection of, 151
Pickering on colour reactions of proteids, 163
Picramic acid, 17
Picric acid, test for sugar, 17, 147
Pigment of red corpuscles, 99 ; of muscle, 177 ; of urine, 192
Pigments, 4 ; pathological, 195
Pilocarpine, administration of, 167
Piotrowski's reaction, 27, 163
Plasma, constituents of, 97 ; gases of, 110 ; proteids of, 97
Plasma of blood, 92, 174 ; of muscle, 176
Pleochromatism, 205
Poisonous alkaloids, 76
Polarimeters, 206 ; of Laurent, 208 ; of Soleil, 207 ; of Zeiss, 208
Polarisation of light, 202, 205
Polariscopes, 206
Polarised light, 202 ; action on carbohydrates on, 16, 17, 18 ; of proteids, 32
Polarising microscope, 204
Polysaccharides, 16, **20**
Pork, 48
Portal vein, 80
Potash, method of extracting glycogen, 156 ; of showing zymogen granules, 167
Potassio-mercuric iodide, 162
Potassium, 3, 8
Potassium ferricyanide in preparation of methæmoglobin, 168
Potassium ferrocyanide in the estimation of phosphate, 187
Potassium permanganate in estimation of uric acid, 190
Potassium sulphocyanide in saliva, 53, 59
Precipitants of proteids, 32, 33, 162
Precipitation, 33
Precipitin, 120
Prévost and Dumas on formation of urea, 127
Primary albumoses, 66 ; proteoses, 66, 72
Principal cells of stomach, 61
Prisms, in direct vision spectroscope, 104, 105
Propeptone, 66, 67
Propionic acid, 24
Prosecretin, 75, 165
Protagon, 179
Protamines, 40
Proteids, 3, 4, 9, **27–40**, 179 ; absorption of, 68 ; classification of, 34 ; coagulated, 35 ; coagulation of, 30 ; colour reactions of, 32 ; composition of, 29 ; compound, 36 ; conjugated, 36 ; crystallisation of, 31 ; definition of, 29 ; digestion of, by gastric juice, 53 ; digestion of, by pancreatic juice, 69 ; molecule, 21 ; of blood plasma, 97 ; of milk, 41, 45 ; of muscle, 176, 177 ; of serum, 97 ; osmotic pressure of, 223 ; in urine, 148 ; precipitants of, 27, 32, 33 ; simple solubilities of, 30 ; tests for, 27, 30, 151–152
Proteid metabolism, 3
Proteid-sparing food, 39
Proteolytic ferments, 56 ; enzymes, 56, 72
Proteoses, 30, 66, **67**, **162**, 95, 163 ; in urine, 148
Prothrombin, 96
Proto-albumoses, 66, 162
Protones, 40
Protoplasm, chemical structure of, 3 ; properties of, 2
Protective inoculation, 116
Proximate principles, classification of, 4 ; of food, 42 ; scheme for detecting, 151–154

Pseudo globulin, 173, 176
Pseudo-mucin, 37
Pseudo-nuclein, 38, 46
Ptomaines, 54
Ptyalin, 19, 56, 59
Pulses, 50
Pumps, mercurial air, 211–215
Purine bases, 38, 138
Purpurate of ammonia, 135
Pus in urine, 150; tests for, 150
Putrefaction, 54, 55
Pyloric glands, 61, 62

Q

Quantitative estimation of albumin, 146; of chlorides, 185, 186; of creatinine, 191; of dextrose, 147; of glycogen, 156; of lactose, 159; of maltose, 159; of nitrogen, 215; of phosphates, 187; of sulphates, 188; of sugar, 147; of urea, 121, 183, 184; of uric acid, 190

R

Racemic acid, 147
Ranke's diet, 43
Reagents necessary for practical work, 4, 5
Réaumur's scale, 8
Receptor group, 119
Red blood corpuscles, 98, 119; composition of, 99; pigment of, 100
Red muscle, 177
Reflex action, 58
Rennet, 46, 51, 56
Rennin, 63
Reproduction, 2
Respiration, chemistry of, 109
Respiratory oxygen of hæmoglobin, 102
Respiratory pigments, 100
Respiratory quotient, 110
Reversible action, 57
Ricin, 119
Rigor mortis, 51, 176, 177
Ringer on contractile tissue, 217
Riva on urochrome, 123
Roberts, Sir William on estimation of sugar in urine, 149
Rockwood and Moore on fat absorption, 90

S

Saccharic acid, 18
Saccharimeters, 206, 207
Saccharoses, 16, 56
St. Martin, Alexis, case of, 61
Salicyl-sulphonic acid, action of, on proteids, 163
Saliva, 29, 53, **58**; action of, 59; composition of, 58; secretion of, 58
Salivary corpuscles, 58; glands, 60
Salkowski's reaction for cholesterin, 70; method of estimating sulphates, 189
Salmine, 40
Salted plasma, 174
Salted whey, 42
Salting-out of colloid carbohydrates, 21; of proteids, 33
Saponification, 22, **25**, 73, 89, 90
Sarcine, 139
Sarcolemma, 39
Sarcosine, 141
Schäfer on fat absorption, 88, 89
Schiff on bile circulation, 81; test for uric acid, 135
Schizomycetes, forms of, 55
Schmalz's capillary picnometer, 202
Schmidt, Alexander, on precipitating serum globulin, 172; on salts of plasma, 98; on preparation of fibrin-ferment, 98
Schröder's work on urea, 130
Schwann, white substance of, 84
Scombrine, 40
Sebum, 84
Secretin, 74, 165
Secretion, internal, 78, 79
Sediments in urine, 142–145
Semipermeable membranes, 219, 220
Serous glands, 60
Serum, 92, 96, 172; albumin, 97; crystallisation of, 32; heat coagulation, 173
Serum casein, 172
Serum globulin, 34, 93, 97; heat coagulation, 173
Serum, bactericidal, globulicidal, 119
Serum lutein, 92
Serum of blood, 92, 94, **96**; proteids of, 97
Serum proteids, 97; separation of, 92
Sheep's-wool fat, 84
Shell of eggs, 48
Side-chain theory, 119
Siegfried on antipeptone, 73
Silicon, 3, 8
Silver nitrate method of estimating chlorides, 185
Skatole, 86
Skatole-amino-acetic acid, 32, 72
Skatoxyl, 195; red, 195
Skimmed milk, 41
Smoky urine, 149
Snake venom, 55, 116, 118
Soap, 73, 74
Sodio-magnesium sulphate, action of, on proteids, 173

Sodium, 3, 8
Sodium bi-carbonate in blood, 112, 113
Sodium chloride, 3; action on proteids, 175; on nucleo-proteids, 37, 175
Sodium hypobromite, method of estimating urea, 121
Sodium phosphate, 29; acid, 29, 113
Sodium sulphate plasma, 92, 174
Solar spectrum, 107, 169
Soleil's saccharimeter, 206, 207
Soluble starch, 20
Solutions, 216
Sorbite, 15
Soret's band, 171
Soup, 52
Specific gravity of blood, 202; of milk, 41, 45; of urine, 121, 124
Specific rotatory power, 209
Spectra, 107; of hæmoglobin and its derivatives, 107, 169; photographic, of methæmoglobin, oxyhæmoglobin, and hæmoglobin, 170; of myohæmatin, 178; of urinary pigments, 194
Spectro-polarimeter, 209
Spectroscope, 103, 104
Spectrum, 107, 169
Spermatozoa, 40
Starch, 4, 13, 16, **20**; soluble, 20; action of malt on, 159; digestion of, 53, 59, 69, 73; test for, 20
Starling and Bayliss on pancreatic secretion, 74
Starling on osmotic pressure of proteids, 223
Steapsin, 25, 56, 69, **73**; action of, 69, 71
Stearic acid, 25
Stearin, 23, 25, 47
Steatolytic ferments, 56
Stellar phosphates, 144
Stercobilin, 83, 85, 86, 192
Stewart, G. N., dietary, 43
Stirling on preparation of pure oxyhæmoglobin, 171
Stokes's fluid, 102, 103
Stomach, glands of, 62, 63
Sturine, 40
Sublingual gland, 58
Submaxillary gland, 58, 60
Succus entericus, 18, 75
Sucroses, 16
Sudan III. stain, 24
Sugar, 4, 13–21; cane, 13, 18; in blood, 87, 98; in urine, 147, 148; in muscle, 18; tests for, **13–21**, 157
Sugar, maple, 18
Sulphates of urine, 148; estimation of, 188; tests for, 121
Sulphonal poisoning, 194
Sulphur, 3, 8, 82; tests for, 10
Suprarenal gland, removal of, 79
Suspending medium, 23
Swim-bladder of fishes, 114
Symbols and atomic weights, 4
Syntonin, 29, **35**, 66, 176

T

Tannin, 162
Tapetum, 139
Tartaric acid, 124
Taurine, 85
Tauro-carbamic acid, 85
Taurocholate of soda, 81
Taurocholic acid, 82
Tchistovitch's experiment, 120
Tea, 52, 138
Teichmann's crystals, 101
Tendon, 29
Tension of aqueous vapour, 8; of gases, 111, 113, 115
Testis, removal of, 78
Tetroses, 16
Theine, 52
Theobromine, 52, 138
Theophylline, 138
Thermometric scales, 8
Thrombin, 95, 96
Thudichum on urochrome, 192
Thyroid gland, 11; removal of, 78
Tissue-fibrinogens, 37, 175
Tissue respiration, 109, 114
Tomes on enamel, 39
Tonsils, 58
Töpfer's test, 165
Tooth, 11
Torricellian vacuum, 102, 212
Torula ureæ, 125
Torulæ, 54, 55, 125
Transitory glucosuria, 148
Triacetin, 25
Triatomic alcohol, 25
Trichloracetic acid as a precipitant of proteids, 163
Trimethyl-xanthine, 138
Triolein, 25
Trioses, 16
Trioxy-purine, 138
Tripalmitin, 25
Triple phosphate, 134, 135, 144
Tristearin, 25
Trommer's test, **13**, 17, 18, 19
Tropæolin test, 165
Trypsin, 56, 69, 70, 71, **72**, 89, 166; action of, 71, **72**, 75
Trypsinogen, 70, 75, 166
Tryptophan, 32, 72; test for, 166
Tunicates, 21
Tyrosine, 40, 71, 72, 73, 76, **77**, 129, 149, 166; crystals, 77; in urine, 128; tests for, 166

U

Umbilical cord, 29
Uncrystallisable sugar, 17
Unorganised ferments, 55, 59
Uræmia, 127
Uranium acetate in estimation of phosphates, 187
Uranium nitrate in estimation of phosphates, 187
Urate, acid ammonium, deposit of, 143; acid sodium, deposit of, 143
Urates, 135, 143, 145
Urea, 4, 12, 121, **125**, 141, 144, composition and compounds, 126; crystals of, 124; decomposition of, 126, 144; estimation of, 121, 184; mode and site of formation, 127, 128; preparation of, 184; quantity excreted, 126; tests for, 121, 125; where formed, 127, 128, 129
Urea nitrate, 125, 126, 135; preparation of, 184
Urea oxalate, 125, 126, 135
Uric acid, 98, 129, 135, **136–140**, 145, **190**; crystals of, 136; estimation of, 137, 190; origin of, 137; preparation of, 136, 190; tests for, 135
Urina potus, 124
Urinary deposits, 142
Urine, **121**; composition of, 125; inorganic constituents of, 121, 132; pigments of, 123, 192; tests for abnormal constituents of, 146, 154; tests for constituents, 121, 153
Urinometer, 124
Urobilin, 83, 85, **123**, 149, 192, 193; absorption bands of, 194
Urobilinogen, 123
Urochrome, 123, 192; uro-erythrin, 143, 193; absorption bands of, 194
Urorosein, 195; absorption bands of, 194

V

Valeric acid, 24
Van't Hoff on polarisation of light, 211; hypothesis, 222
Vegetable acids, 4, 133; food, composition of, 50; parchment, 31
Vegetables, composition of, 50; green, 52
Venous blood, gases of, 110
Villus, section of, 88
Vinegar, 54
Vital action, 222
Vitellin, 32, **37**, 38, 48
Vitellose, 66
Vitreous humour, 29

W

Waller's modification of Zuntz's gas apparatus, 214
Warm bath, 28
Water in protoplasm, 3, 4; density of, 8
Wave theory of light, 202
Weights and measures, 6, 7
Whartonian jelly, 29
Whey, 41, 46; salted, 41
Whey-proteid, 46
White blood corpuscles, 95, 96, 98
White of egg, 3, 27, 28, 30, **48**, 53, 69
Whole flour, 49
Witte's peptone, 27, 162, 163
Wöhler, preparation of urea, 125
Wooldridge on tissue-fibrinogen, 37, 95, 175
'Worth' of corpuscles, 202

X

Xanthine, 71, 98, 138, **139**
Xanthine group, 38, 138
Xanthoproteic test, 27, 32
Xylose, 158

Y

Yeast, action of, 17, 19; cells, 54; in bread-making, 50; in testing for sugar, 149
Yolk of eggs, 48

Z

Zeiss's polarimeter, 208
Zinc chloride creatinine, 141
Zuntz's gas apparatus, 214
Zymogen, **63**, 70, 98; granules, 167

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